



Characterizing In Vitro Bioavailability of Acyclovir and Metronidazole Topical Products, and In Vitro – In Vivo Correlation Results with Transdermal Systems

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Topical Dose Administration Techniques

- Highly variable among labs, researchers, and patients
 - Methods of dispensing formulation
 - Duration of rubbing
 - Force used for rubbing
 - Loss of formulation during rubbing



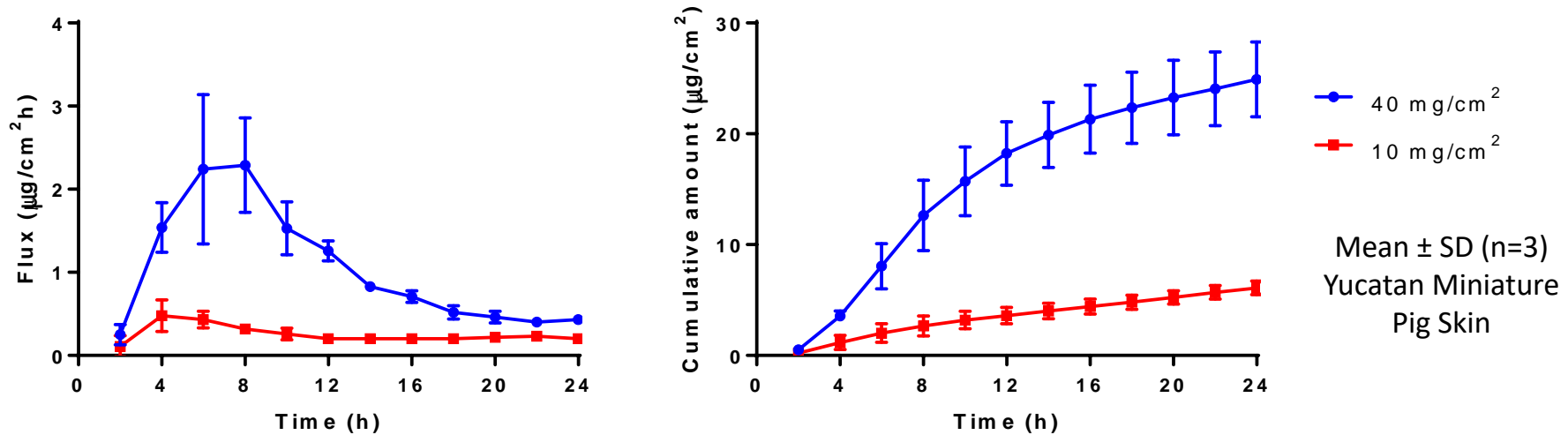
- Need a reproducible, clinically-relevant, and practical technique for IVPT

Image from <http://www.telegraph.co.uk/expat/expatlife/10441983/Pale-and-interesting.html>

IVPT Results Variability

Importance of Dose Application – Voltaren® gel example

Dose Test and Reference Products the Same

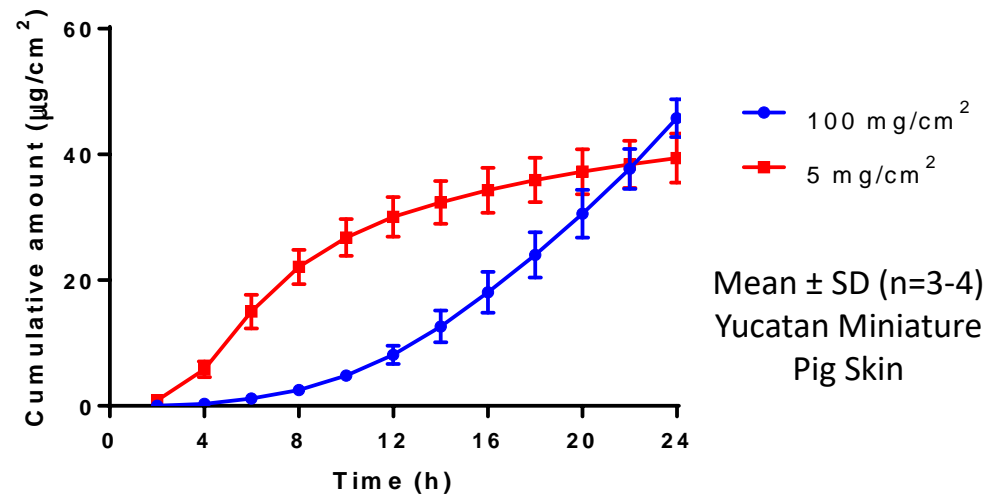
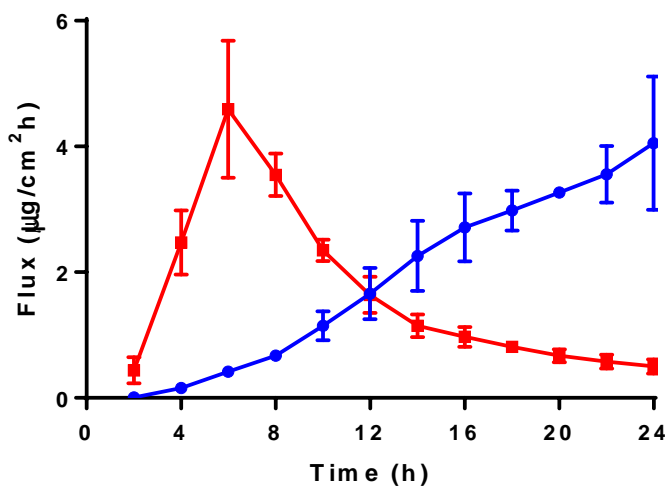


| | $J_{\max} \pm \text{SD}$ ($\mu\text{g}/\text{cm}^2/\text{h}$) | T_{\max} (h) | Cumulative Amount $\pm \text{SD}$ ($\mu\text{g}/\text{cm}^2$) |
|----------------------------|---|----------------|---|
| 40 mg/cm^2 | 2.29 ± 0.57 | 8 | 24.91 ± 3.38 |
| 10 mg/cm^2 | 0.48 ± 0.19 | 4 | 6.10 ± 0.61 |

HPLC vial rubbing application technique

IVPT Results Variability

Importance of Dose Application – Pennsaid® 2% Dose Test and Reference Products the Same

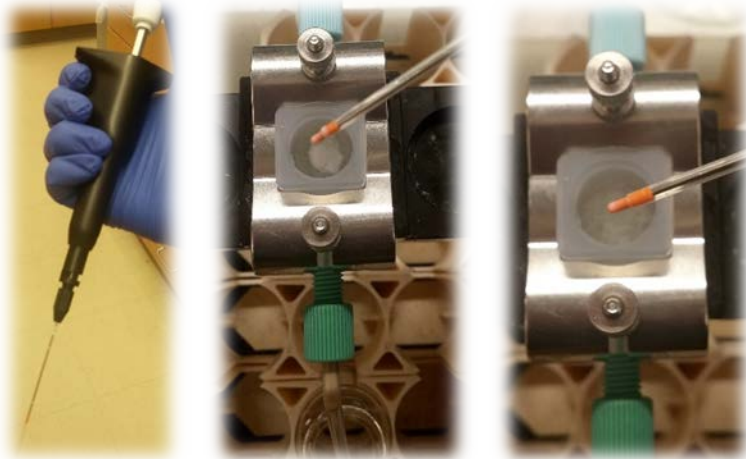


| | $J_{\text{max}} \pm \text{SD}$ ($\mu\text{g}/\text{cm}^2/\text{h}$) | T_{max} (h) | Cumulative Amount \pm SD ($\mu\text{g}/\text{cm}^2$) |
|------------------------|---|----------------------|--|
| 100 mg/cm ² | 4.05 \pm 1.06 | 24 | 45.79 \pm 3.00 |
| 5 mg/cm ² | 4.59 \pm 1.09 | 6 | 39.43 \pm 3.90 |

HPLC vial rubbing application technique

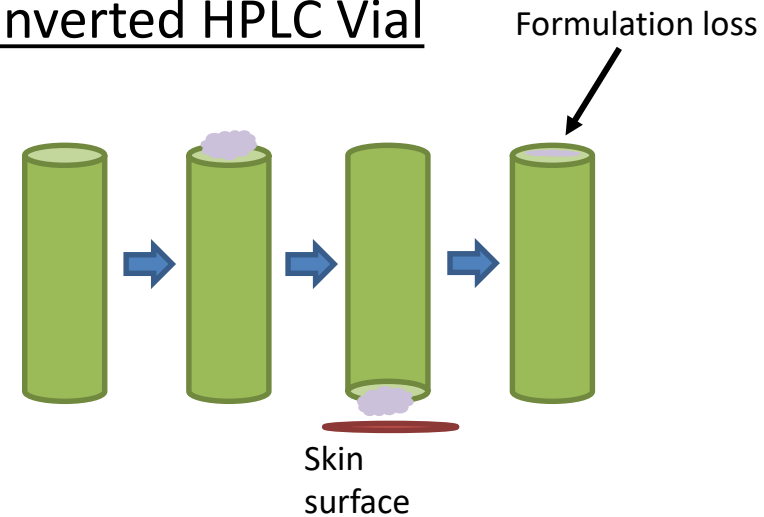
Dose Administration Techniques

Positive Displacement Pipette



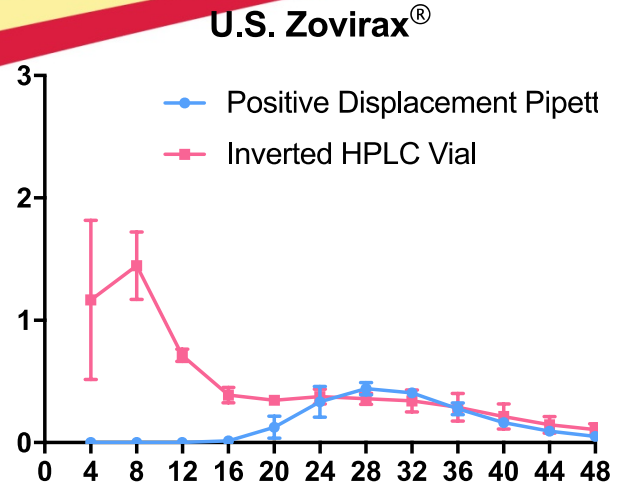
- Quick, convenient, low variability
- Minimal formulation loss
- Lack of rubbing effect

Inverted HPLC Vial



- Time-consuming, more variability
- Some formulation loss
- Simulates clinically-relevant rubbing effect

Dose Administration Techniques

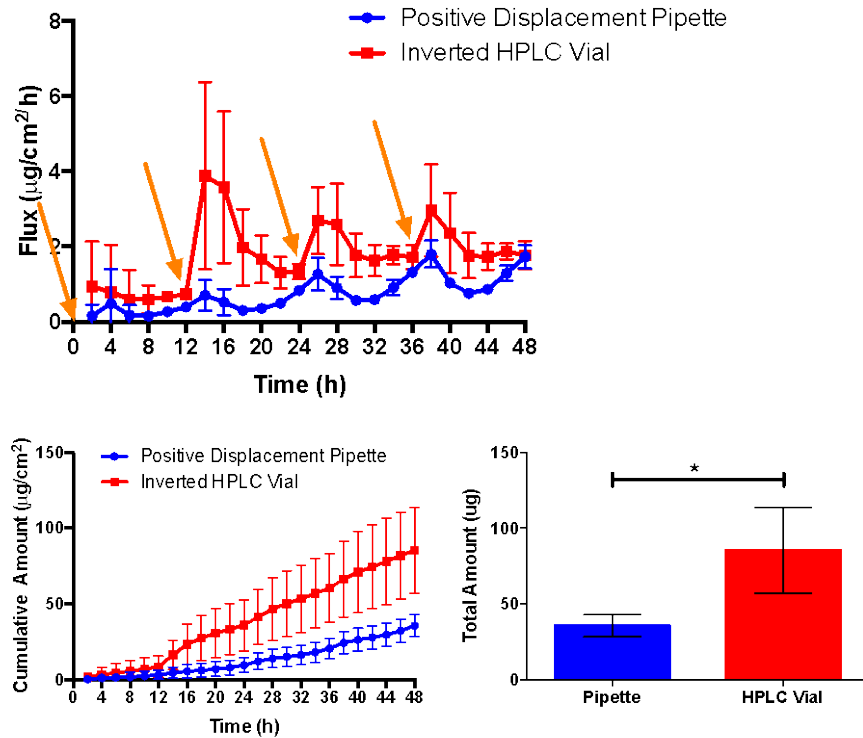


Ex vivo human skin
Mean \pm SD (n=3-4 for each technique)

/cm²)

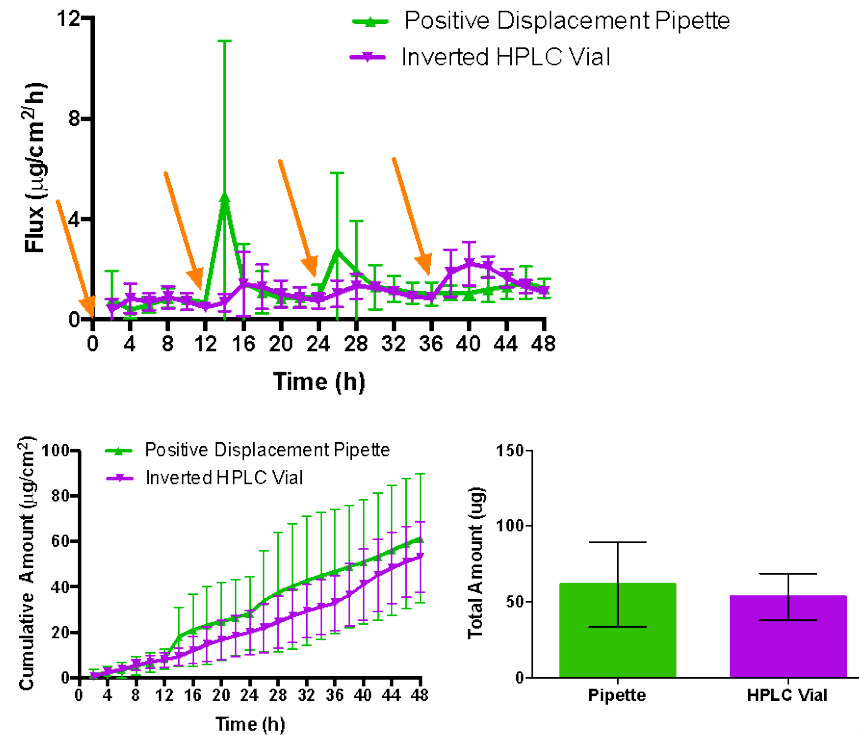
Preliminary: Dose Administration Techniques

Pennsaid® 2% (more viscous)



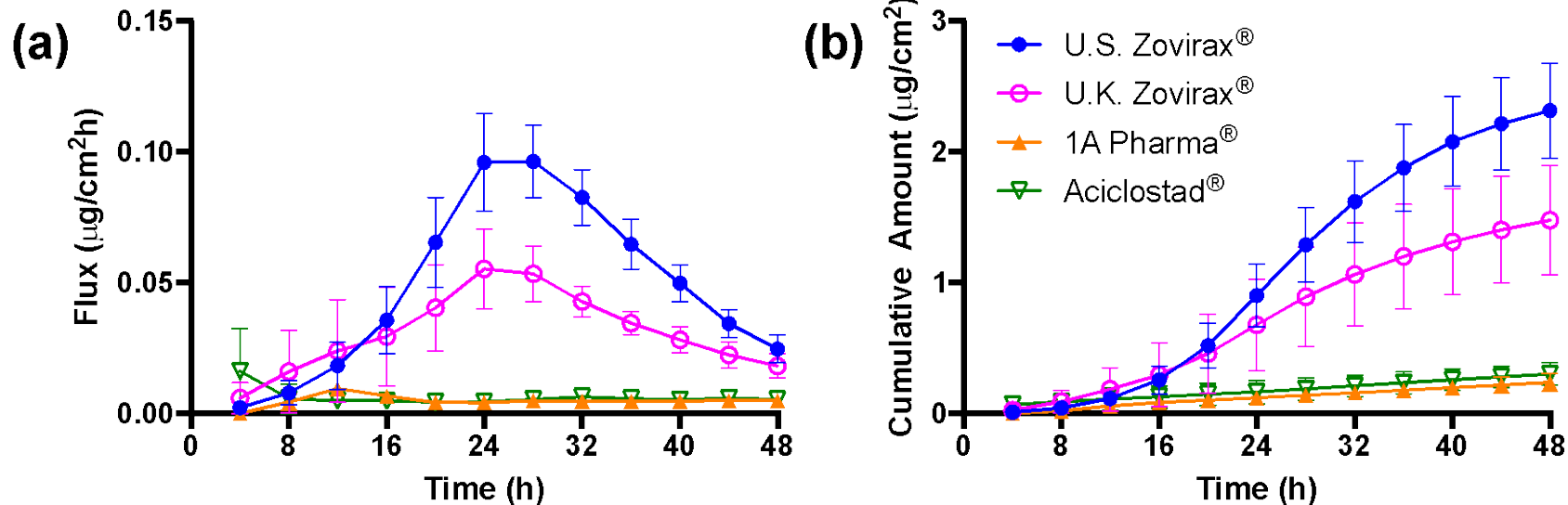
Orange Arrow: dosing ($\sim 5 \text{ mg}/\text{cm}^2$ of formulation)

Pennsaid® 1.5%



Mean \pm SD (n=3-4)
Yucatan Miniature Pig Skin

Four Acyclovir Cream Products

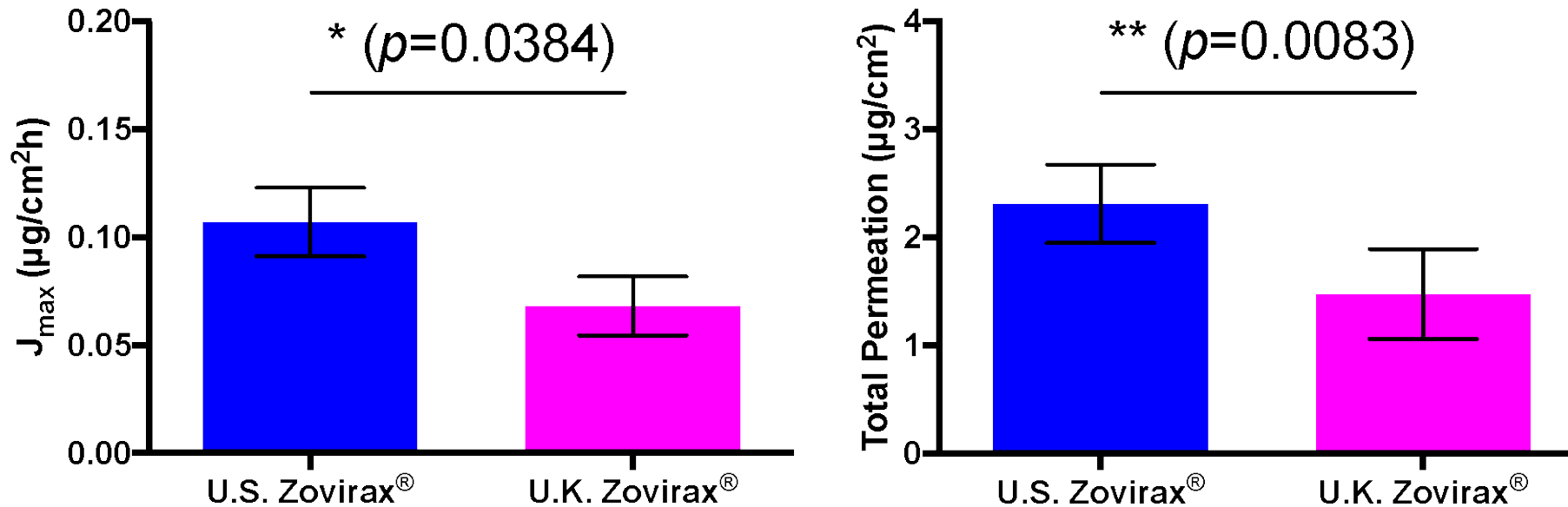


(Mean \pm SEM, n= 6 donors with 4-7 replicates per donor for Zovirax® creams and n = 2 donors with 3-4 replicates per donor for non-Zovirax® creams)

****The IVPT method was able to discriminate the Reference and Test acyclovir products, based on Jmax and the total amount of acyclovir permeated over 48 h**

Positive displacement pipette application

J_{\max} and the total amount of acyclovir permeated over 48h between Reference and Test

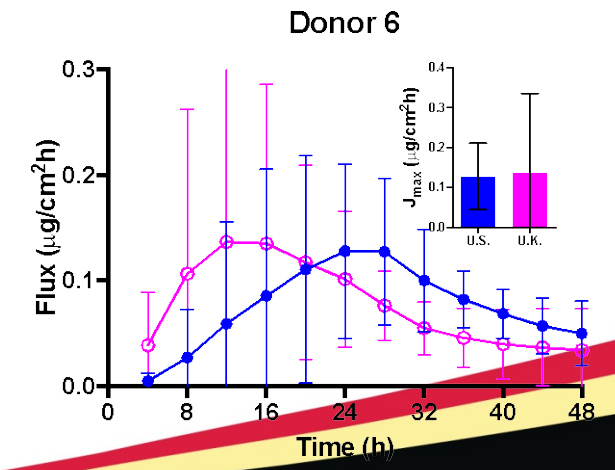
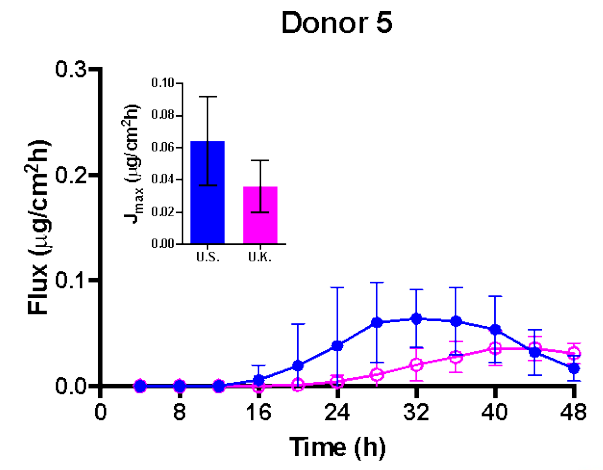
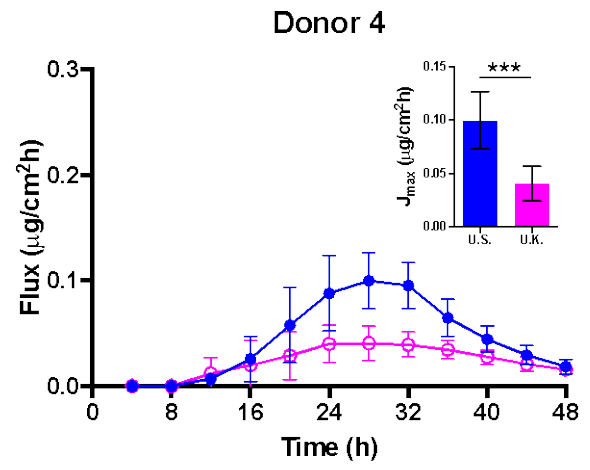
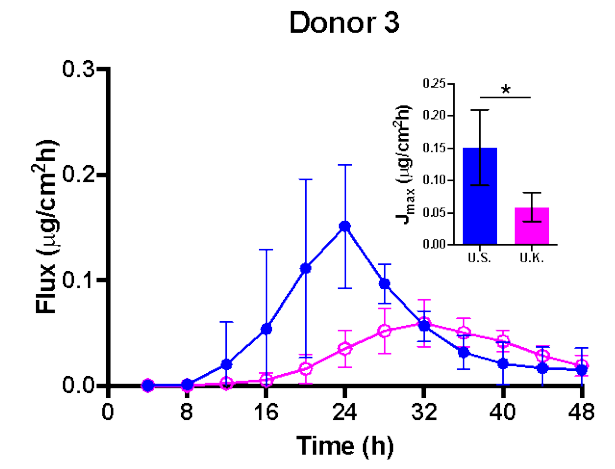
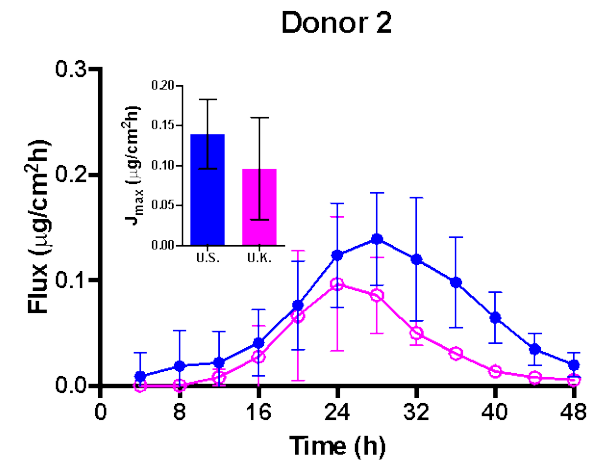
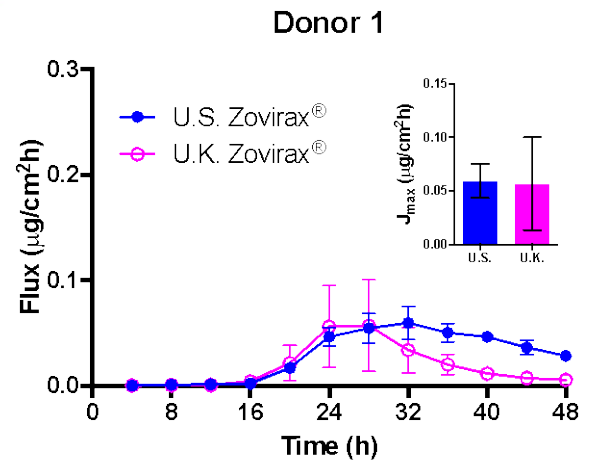


Positive displacement pipette application

Comparisons of products (Mean \pm SEM, n= 6 donors

with 4-7 replicates per donor)

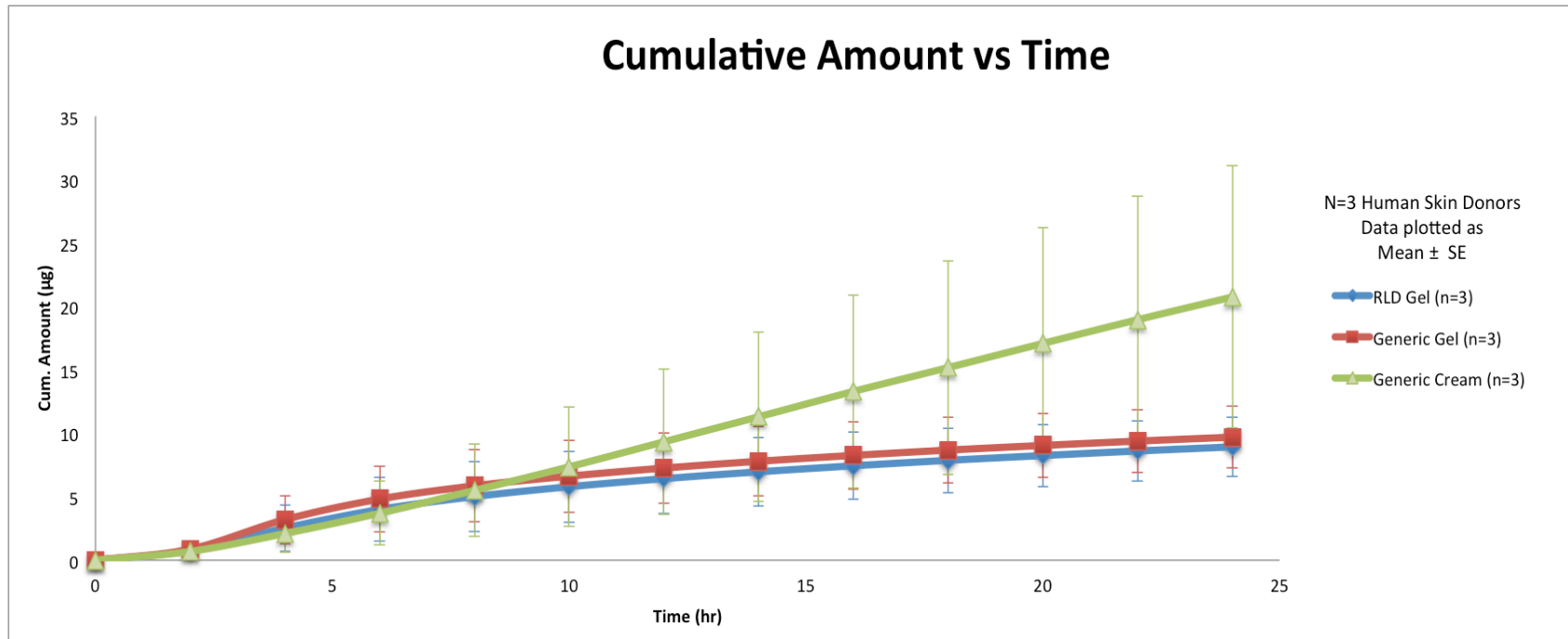
U.S. vs. U.K. Zovirax[®] creams per donor



Positive displacement pipette application

Metronidazole RLD Gel & Generic vs. Generic Cream

| Product Name | Cumulative Cutaneous Absorption (μg) |
|---------------------|---|
| RLD Gel (n=3) | 8.93 ± 2.33 |
| Generic Gel (n=3) | 9.70 ± 2.42 |
| Generic Cream (n=3) | 21.0 ± 10.32 |



Cumulative absorption from RLD gel, generic metronidazole gel and generic metronidazole cream over 24-h study duration.

Dosing Technique: Inverted HPLC vial

Target dose: 10 mg/cm^2

Flow rate: 1.0 mL/h

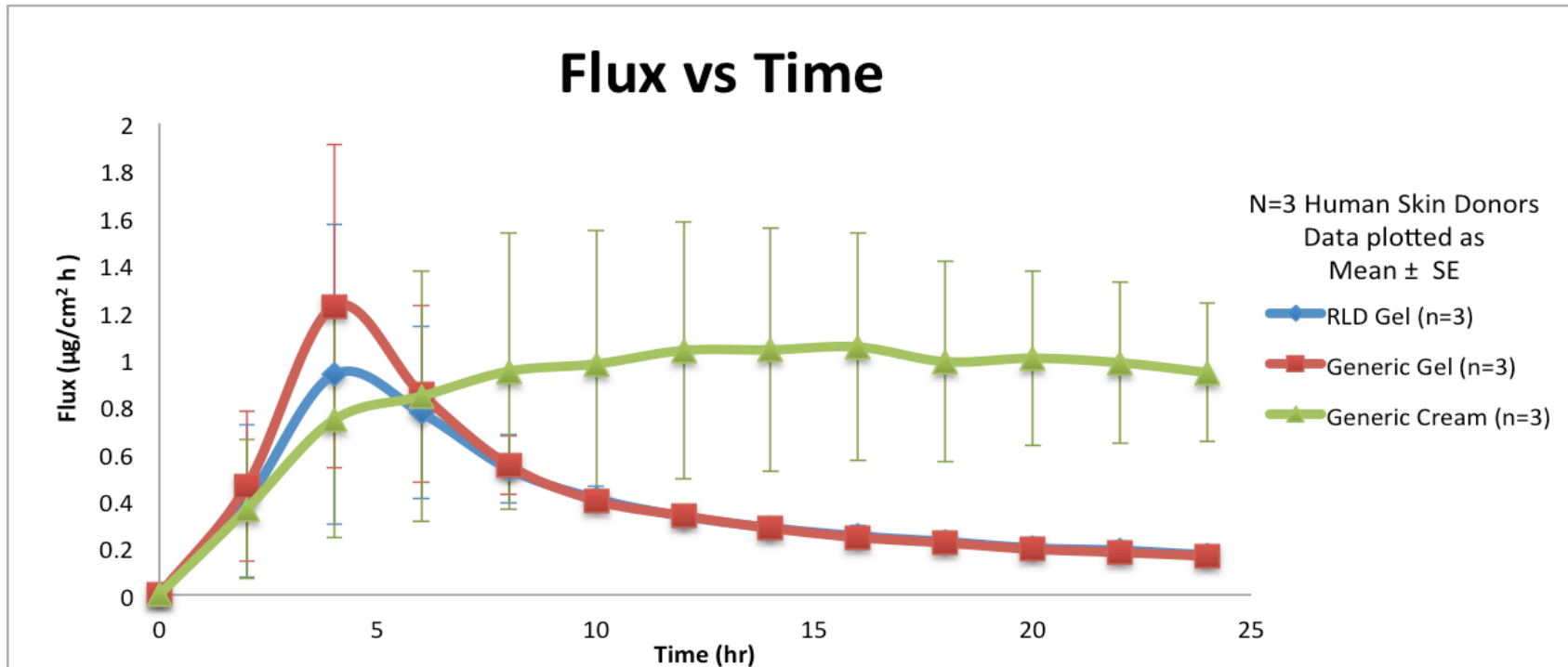
Skin surface temperature: $32 \pm 2^\circ\text{C}$ (circulating water bath)

Receiver solution: Isotonic phosphate buffer ($\text{pH } 7.4 \pm 0.1$)

Skin: human abdominal skin from three donors with four replicate skin sections per donor per product

Metronidazole RLD Gel & Generic vs. Generic Cream

| Product | Maximum Flux ($\mu\text{g}/\text{cm}^2/\text{h}$) |
|---------------------|---|
| RLD Gel (n=3) | 0.93 ± 0.63 |
| Generic Gel (n=3) | 1.22 ± 0.69 |
| Generic Cream (n=3) | Observed at ≥ 12 h |



Flux profile from RLD gel, generic metronidazole gel and generic metronidazole cream over 24-h study duration.

Conclusion: Metronidazole IVPT results

- IVPT studies may have utility to help support an evaluation of bioequivalence for topical drug products
 - RLD and generic gels
 - Positive controls for bioequivalence relative to each other
 - Had a similar rate and extent of metronidazole delivery
 - Discriminated the cutaneous bioavailability from the cream as being different from that for both gels
 - Generic cream
 - Negative control for bioequivalence relative to the reference gel
 - Distinct rate and extent of metronidazole delivery with respect to both gels
- Consistent with the expectation that differences in physical and structural critical quality attributes between topical semisolid drug products (e.g., between a gel and a cream) can alter the bioavailability of metronidazole



Can the *in vitro* permeation test (IVPT) predict the performance of TDS (patch) and heat effects on drug delivery and absorption *in vivo*?

Model Drugs: **Nicotine** & Fentanyl

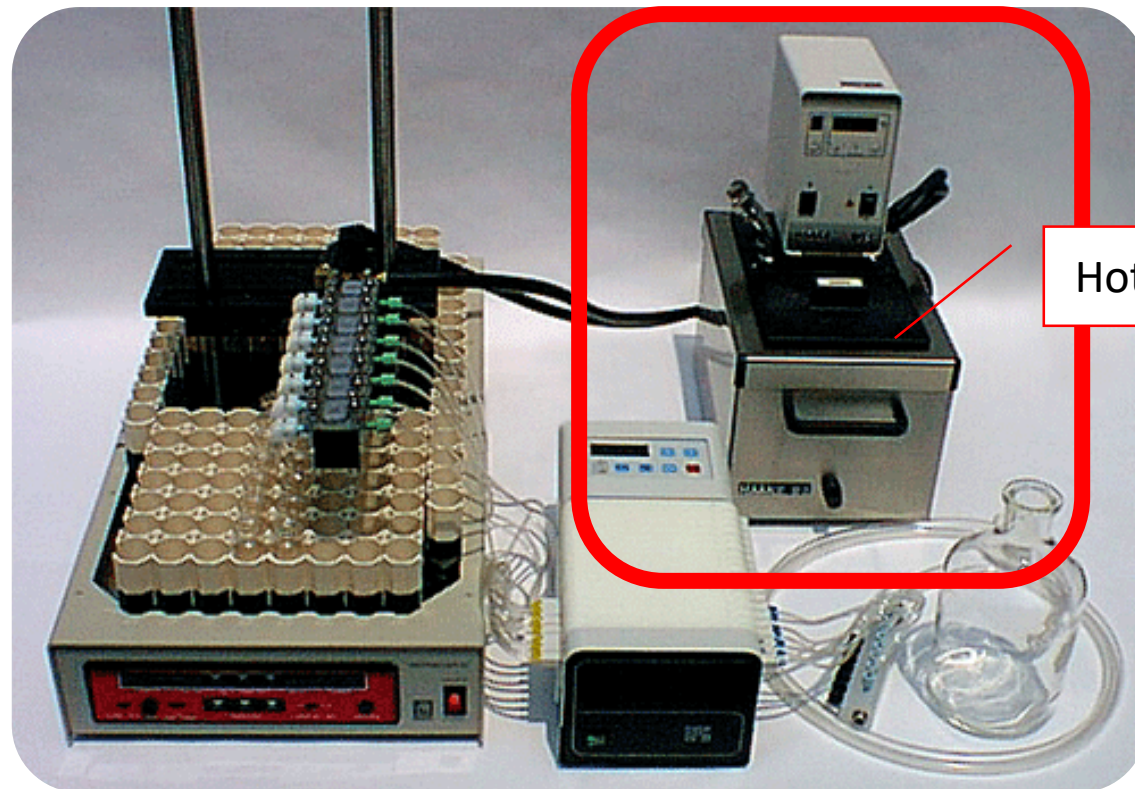
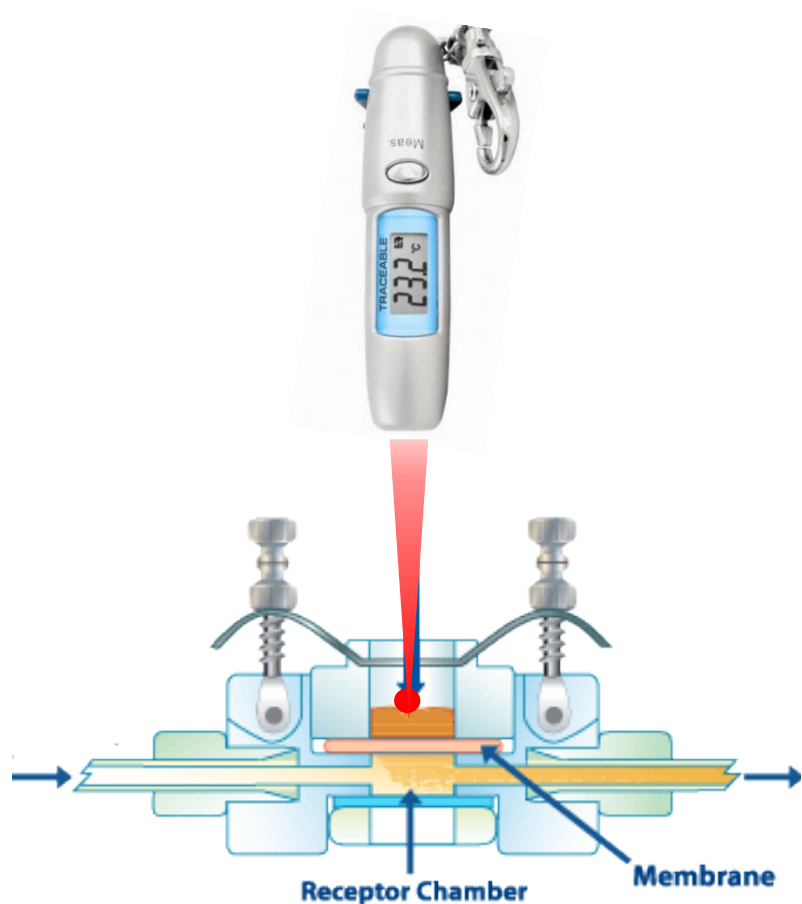
- I. Evaluation of the influence of transient heat (1 h) on the release and permeation of drug from TDS using the *in vitro* permeation test (IVPT)
- II. Evaluation of the influence of transient heat (1 h) on the TDS pharmacokinetics *in vivo* by conducting PK studies in human subjects
- III. Evaluation of preliminary *in vitro* and *in vivo* correlations (IVIVC) of TDS

***This TDS project is informative for topical drug product evaluation since many provide quantifiable blood levels of drug**



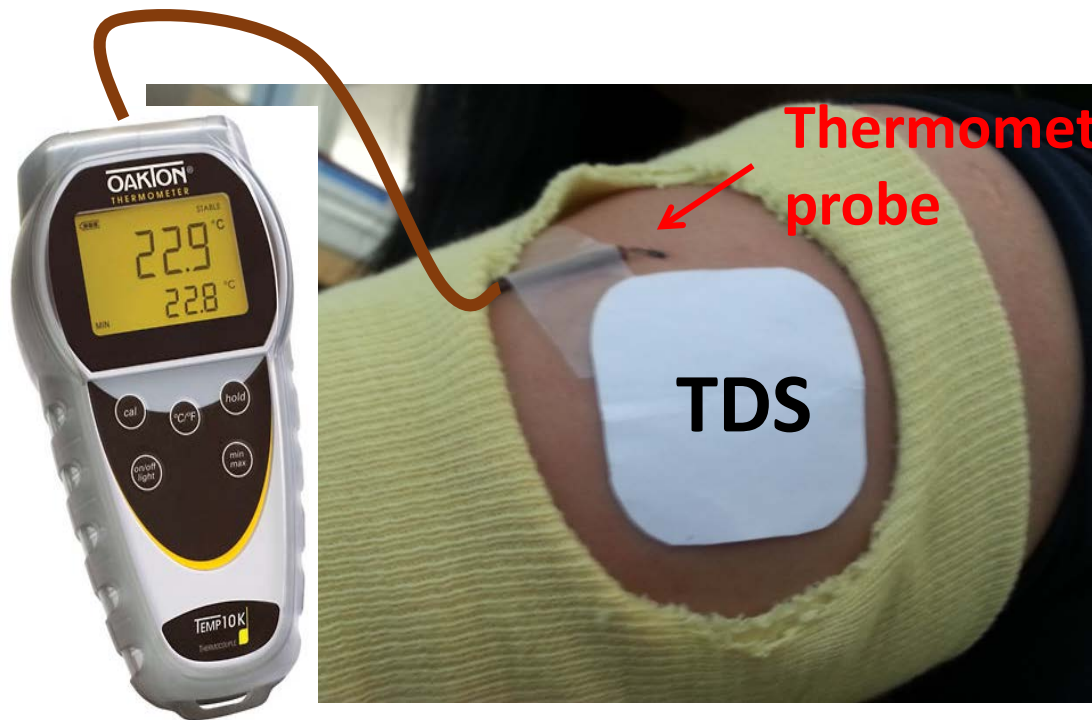
Temperature Monitoring & Heat Application *In Vitro*

Infrared Thermometer





Temperature Monitoring & Heat Application *In Vivo*

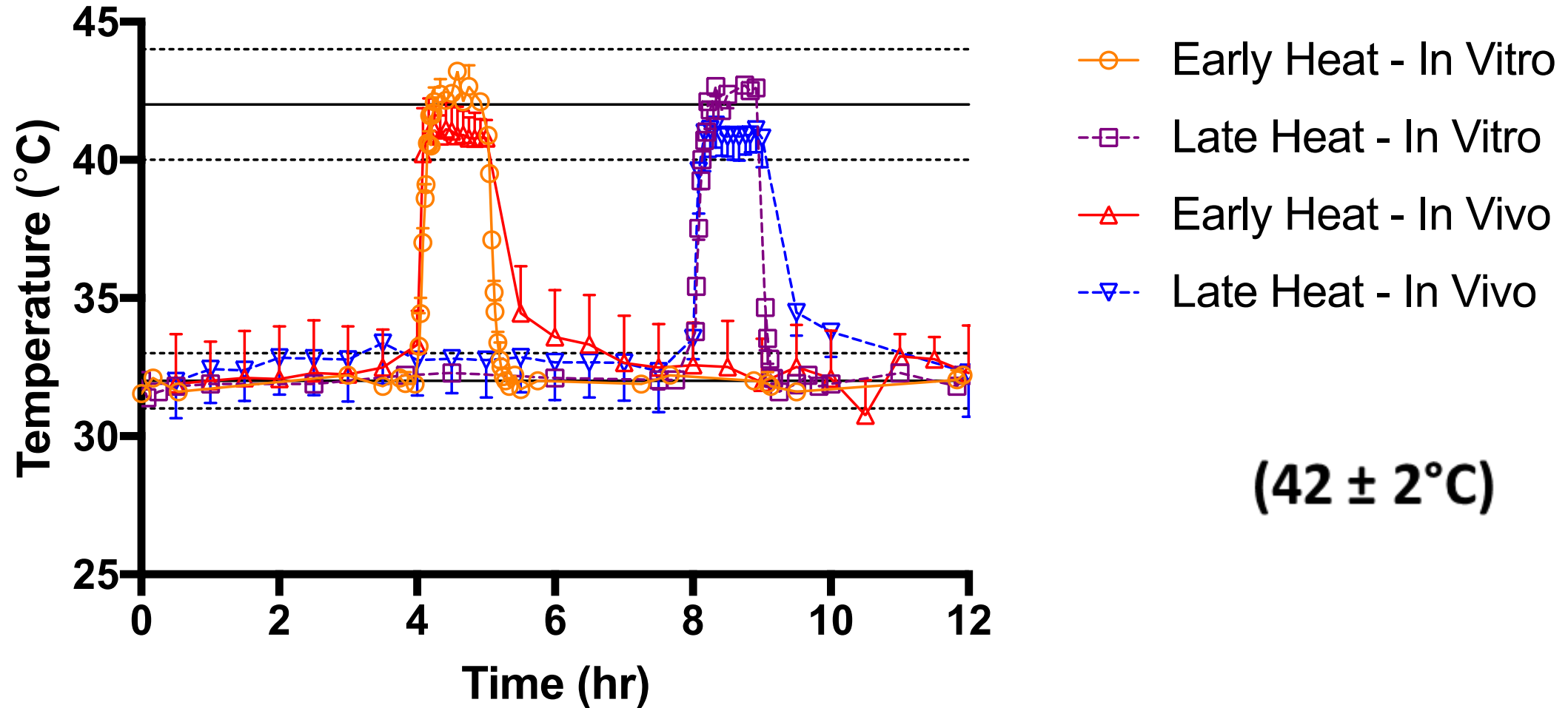


- Kevlar sleeve with an opening to expose TDS, while protecting skin outside the dosing area
- Thermometer probe adjacent to TDS

- Pre-heated heating pad
- ACE™ Bandage to ensure good contact between TDS and heating pad

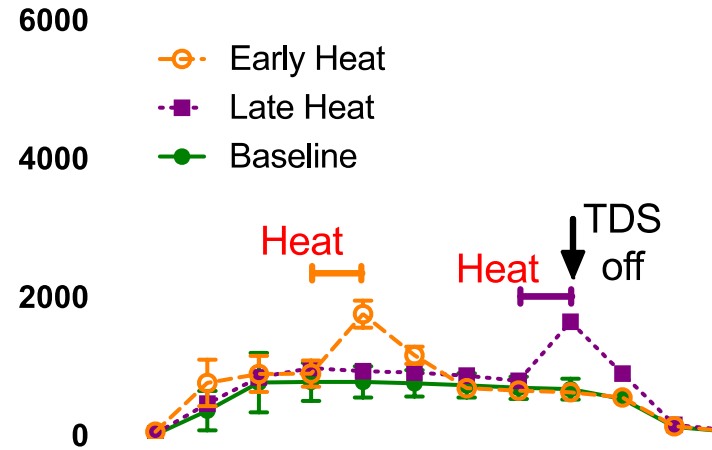
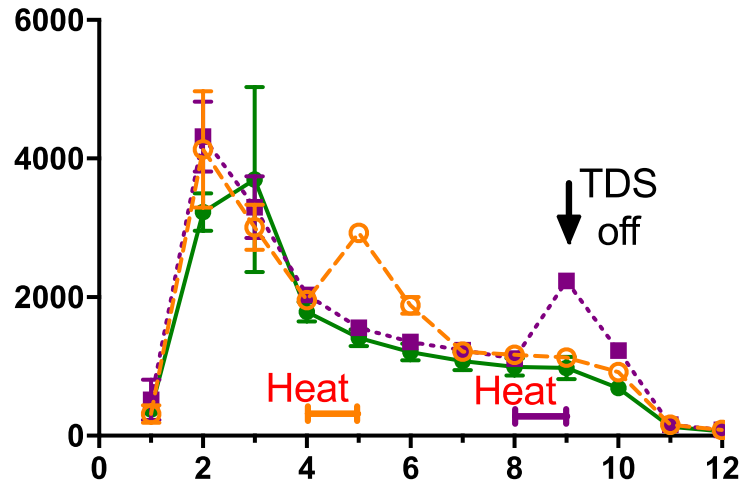


Temperature: *In Vitro* & *In Vivo*

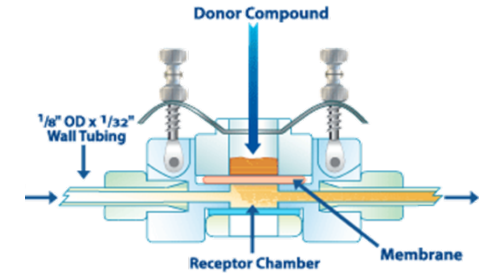




IVPT Results

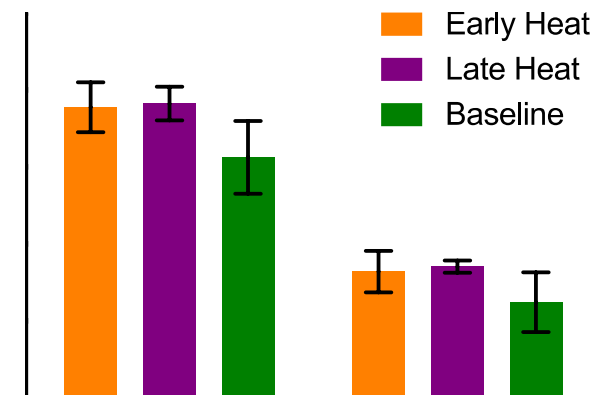
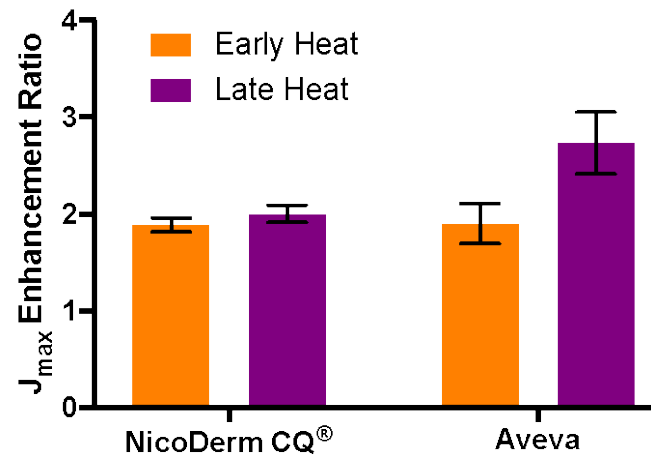
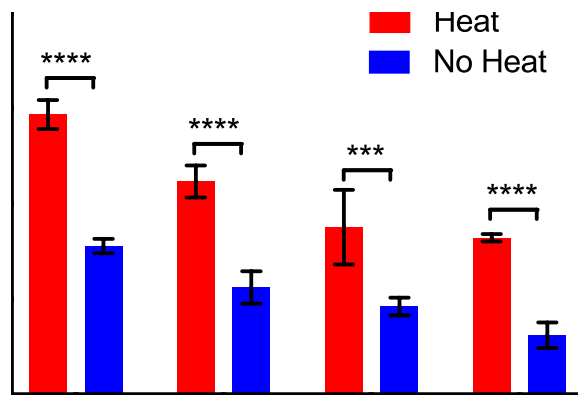


Human Skin Data



Mean \pm SEM from 4 donors for Early Heat and Late Heat, 2 donors for Baseline with n=4 per donor

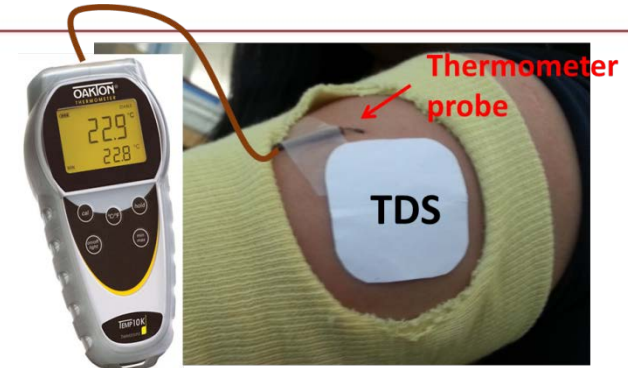
Flux+ = Flux value multiplied by TDS size to account for the whole TDS



Two-way ANOVA followed by Bonferroni's post-hoc multiple comparisons

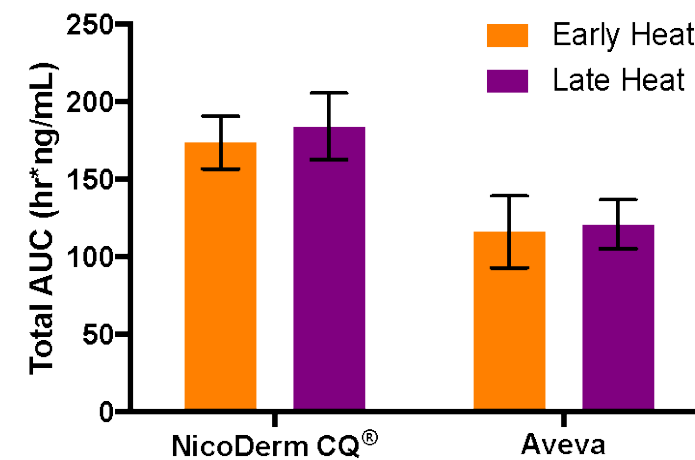
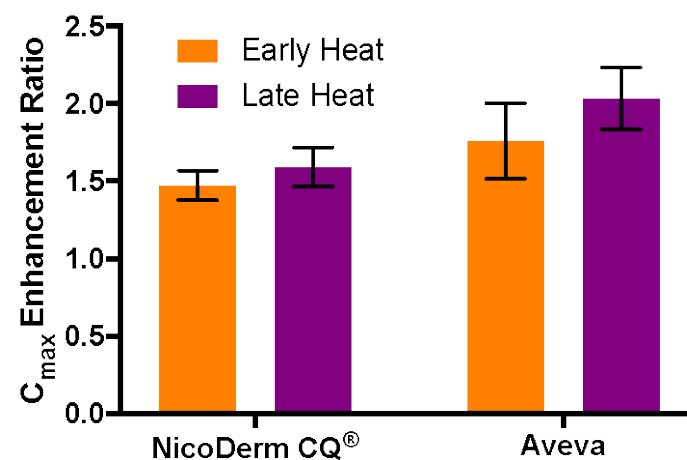
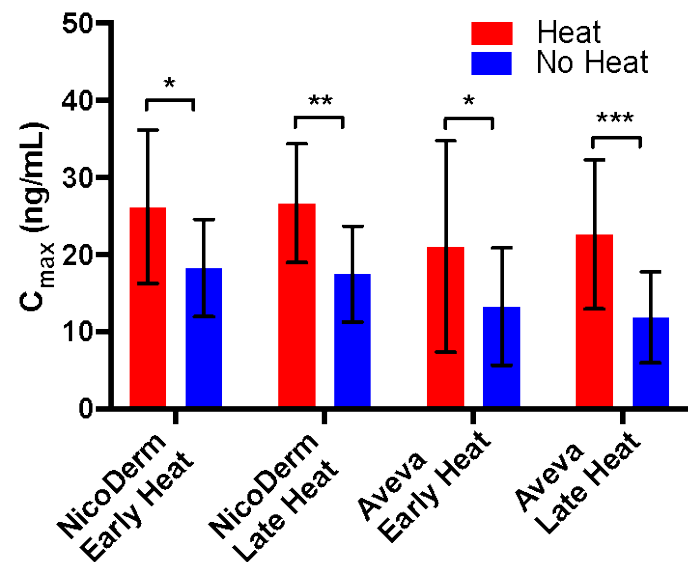
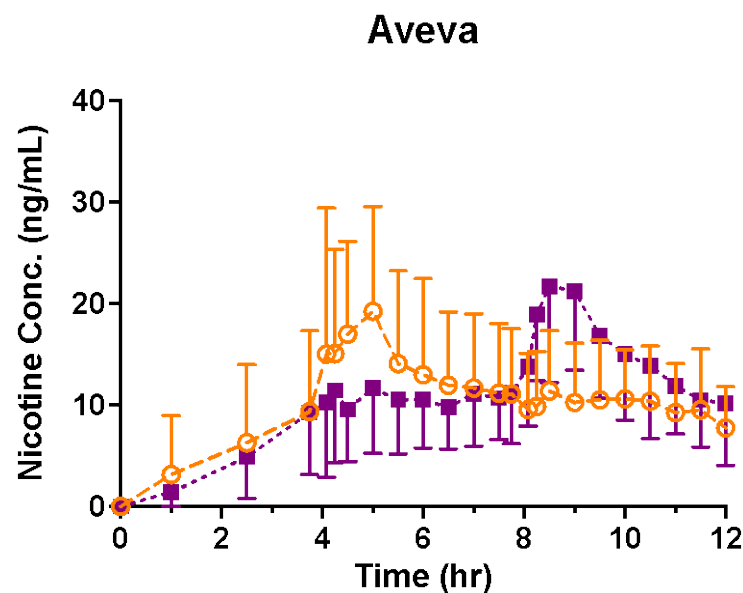
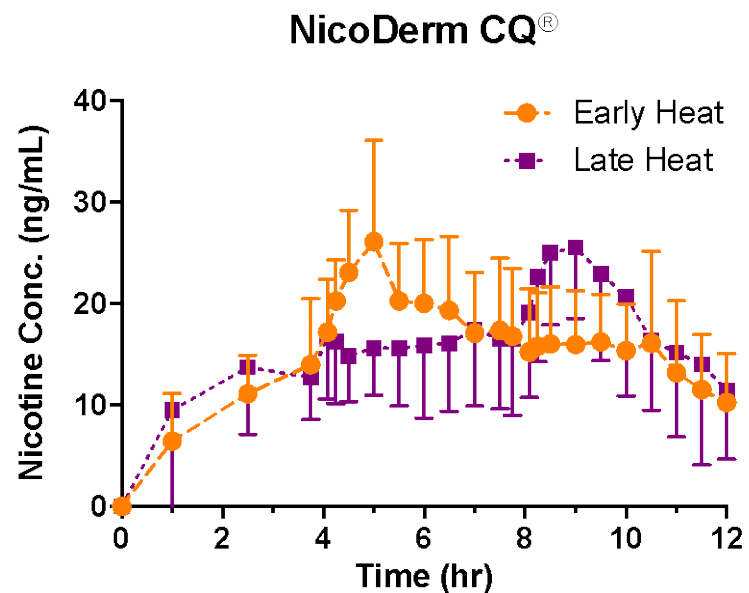


In Vivo Results



Mean ± SD
from 10 human subjects

Smokers 
Patch off 9h



Two-way ANOVA followed by Bonferroni's post-hoc multiple comparisons



Conclusions – Nicotine

- Early vs. Late Heat effect comparable both *in vitro* and *in vivo*
- Heat effect on two differently formulated TDS comparable both *in vitro* and *in vivo*
- *In vitro* and *in vivo* heat effect ratios were comparable
- Strong preliminary IVIVCs (IVIVRs) between IVPT and clinical human PK studies under the matched study designs



IVIVC

- Definition by the U.S. FDA
 - “a predictive mathematical model describing the relationship between an *in-vitro* property of a dosage form and an *in-vivo* response”
- Level A: a point-to-point correlation between *in vitro* and *in vivo* profiles
- Level B: comparison between *in vitro* dissolution time and *in vivo* residence time
- Level C: a single point correlation between *in vitro* and *in vivo* parameters (e.g. J_{\max} vs. C_{\max})

Level A is most informative and useful



Approach I Level A

Eq. 1 prediction while TDS was worn:

$$C_s = \frac{F \times R_{in} \times H_i}{CL_{IV}} \times (1 - e^{-k_1 t})$$

Eq. 2 prediction after TDS removal:

$$C_s = C_0 \times e^{-k_2 t}$$

Or may need 2 or 3 compartment model
Depending on drug and available data

C_s : Predicted in vivo serum concentration

F : Absolute bioavailability for TDS $F = \frac{AUC_{0-\infty, TDS} \times Dose_{IV}}{AUC_{0-\infty, IV} \times Dose_{TDS}}$

R_{in} : Rate of input (mean flux during steady-state in IVPT experiments)

H_i : In vitro heat effect coefficient (composite heat effect during and after heat exposure);
ratio of flux values with heat and without heat

CL : Total body clearance obtained from literature/product package information

k : Elimination rate constant obtained from literature/product package information

(k_1 : after IV dose; k_2 : after TDS dose) k_1 is a derived PK parameter from the two fundamental PK parameters (Cl and V). $k_1 = Cl/V$. k_1 is a re-parameterization of Cl and V
 $F \times R_{in}$ is used to mimic an IV dose and as a result Cl_{IV} is used. Therefore K_{IV} (Cl_{IV}/V)

t : Time after administration of TDS for Eq.1 and time after removal of TDS for Eq. 2

C_0 : Initial concentration after TDS removal



Approach II and III

1. Reconstruct baseline (without heat) profile by combining non-heat portion from two study designs



2. Deconvolute in vivo baseline conc. vs time profile using Phoenix[®]



3. Construct IVIVC model by plotting fraction permeated in vitro vs. fraction absorbed in vivo



4. Predict in vivo fraction absorbed using the IVIVC model and IVPT data



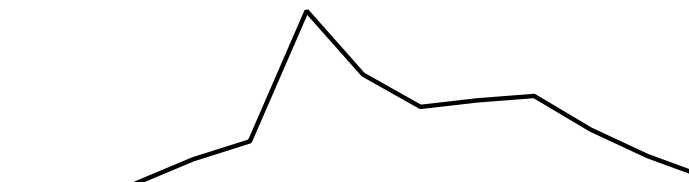
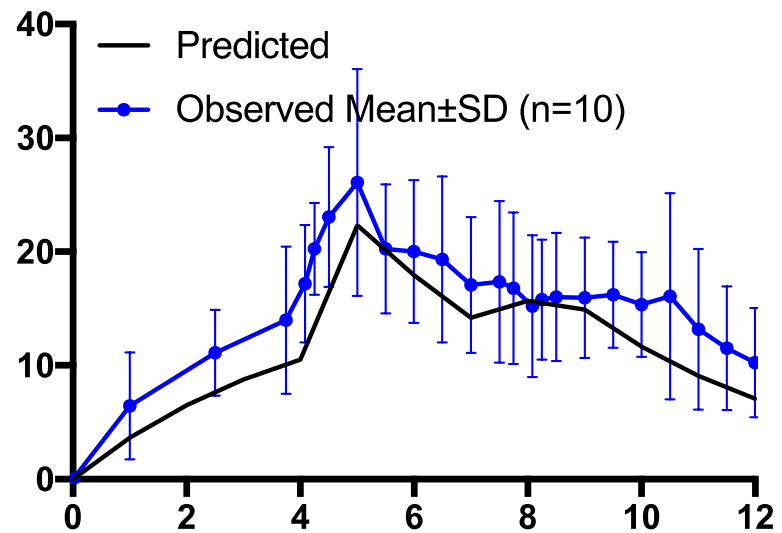
5. Convolute the predicted in vivo fraction absorbed data using Phoenix[®] to obtain conc. vs. time profile



6. Apply **in vitro** heat effect coefficient, H_i (**Approach II**) or **in vivo** heat effect coefficient, H_{ii} (**Approach III**) to the predicted in vivo profile



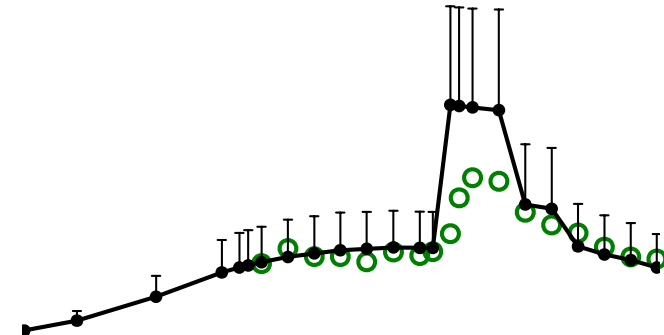
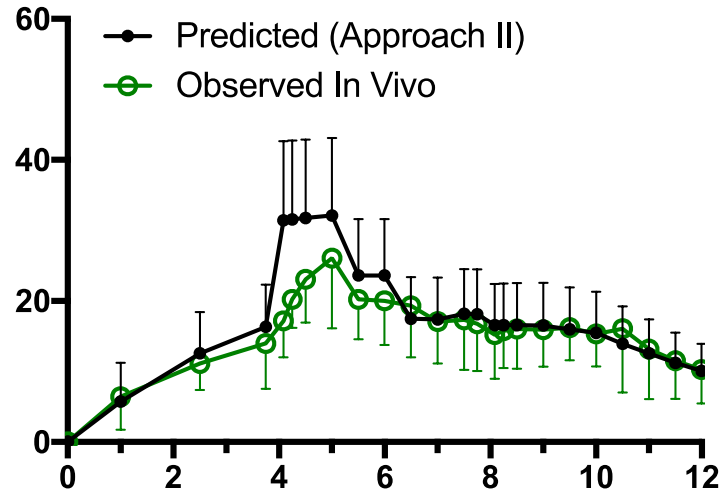
Approach I





Approach II

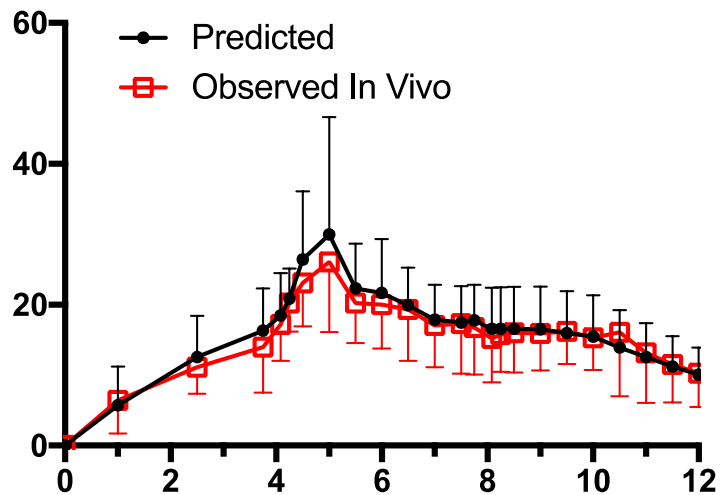
in vitro heat effect
coefficient, H_i





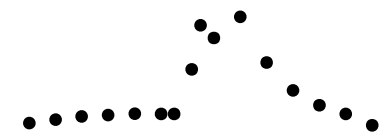
Approach III

in vivo heat effect
coefficient, H_{ii}



Aveva - Late Heat

Nicotine Conc. (ng/mL)



Time (h)



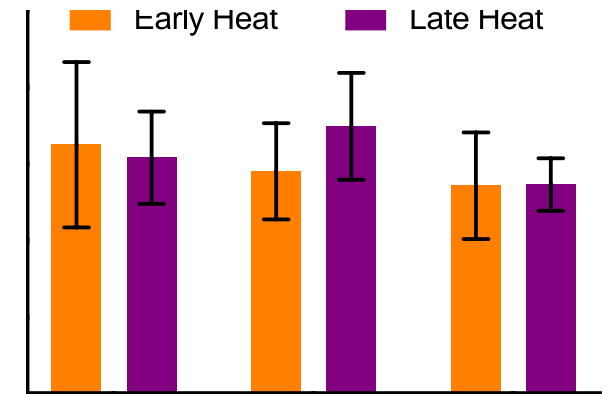
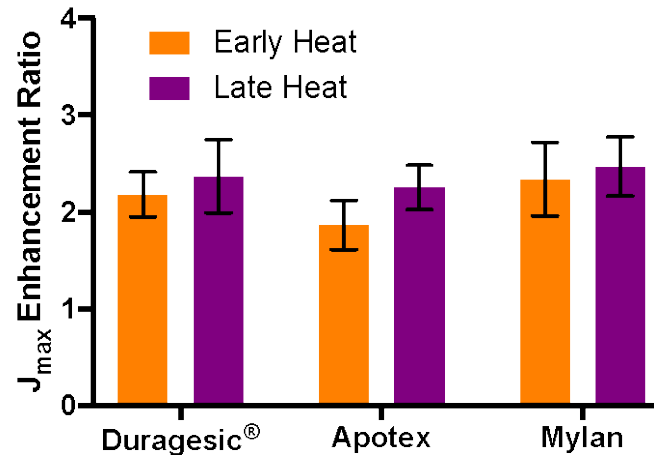
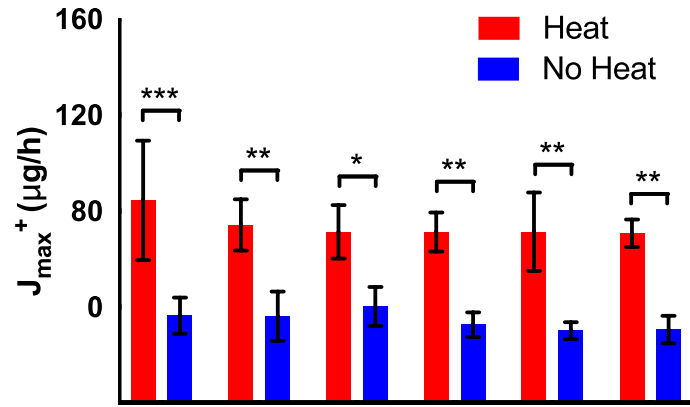
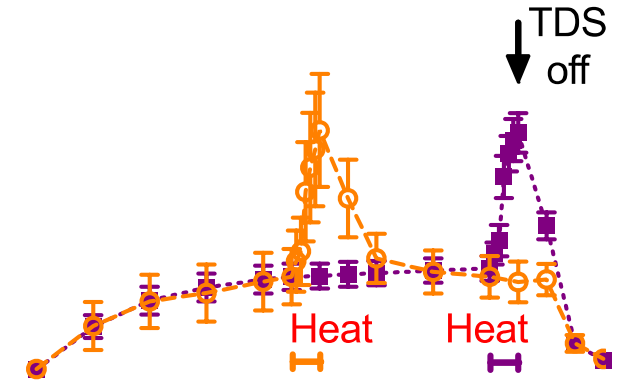
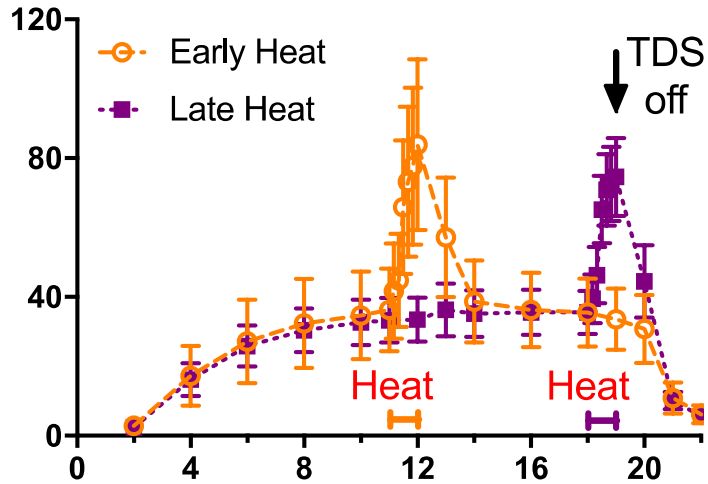
% Prediction Error

| Nicotine TDS | NicoDerm CQ [®] | | Aveva | |
|---------------------|--------------------------|-----------|------------|-----------|
| | Early Heat | Late Heat | Early Heat | Late Heat |
| Approach I | | | | |
| Total AUC | 20.3 | 12.9 | 7.5 | 5.0 |
| C _{max} | 14.4 | 16.6 | 9.8 | 13.5 |
| Approach II | | | | |
| Total AUC | 10.3 | 5.0 | 1.5 | 13.3 |
| C _{max} | 23.3 | 30.2 | 3.5 | 47.5 |
| Approach III | | | | |
| Total AUC | 5.1 | 1.2 | 1.1 | 4.5 |
| C _{max} | 15.0 | 5.8 | 8.9 | 17.7 |



Fentanyl IVPT Results

Mean \pm SEM from 4 donors
with n=4 per each donor



Flux+ = Flux value multiplied by TDS size to account for the whole TDS

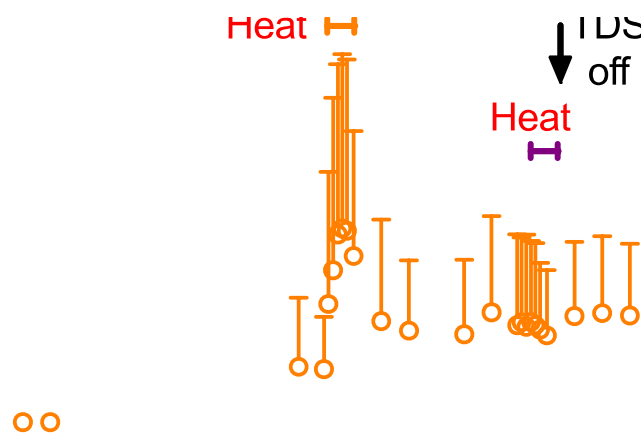
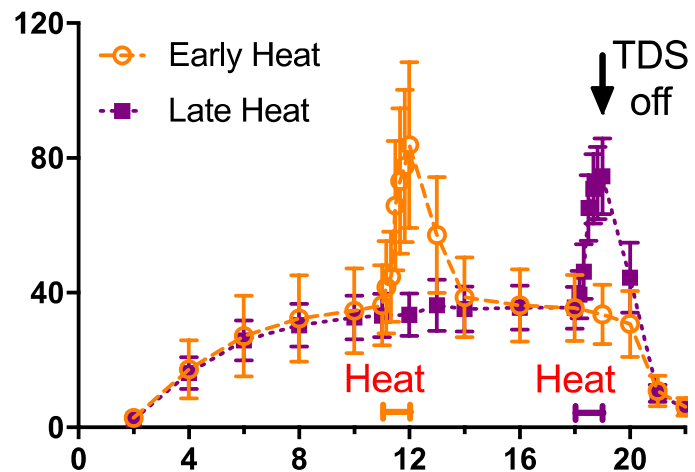
Two-way ANOVA followed by Bonferroni's post-hoc multiple comparisons



Mean \pm SEM from 4 donors
with n=4 per donor (Human Skin)

Fentanyl Results

Mean \pm SD from 10 Healthy Adults



Flux+ = Flux value multiplied by TDS size to account for the whole TDS



Clearance Value of Fentanyl

| Reference | Subject # | Condition | CL _{IV} (L/h) | # of comp for PK Analysis |
|--|-----------|-----------|------------------------|---------------------------|
| Ariano et al. J Clin Pharmacol 2001 | 18 | Healthy | 128 | 1 |
| Bower et al. Br J Anaesth 1982 | 7 | Healthy | 92 | 2 |
| Bentley et al. Anesth Analg 1982 | 5 | Surgical | 59 | 3 |
| McClain et al. Clin Pharmacol Ther 1980 | 5 | Healthy | 57 | 3 |
| Varvel et al. Anesthesiology 1989 ¹ | 8 | Surgical | 46 | 3 |
| Shibutani et al. Anesthesiology 2004 | 16 | Surgical | 43 | 3 |
| Haberer et al. Br J Anaesth 1982 | 13 | Surgical | 42 | 2 |
| Scott et al. J Pharmaol Exp Ther 1986 | 15 | Healthy | 34 | 2 |
| Hengstmann et al. Br J Anaesth 1980 | 5 | Surgical | 26 | 2 |
| Schleimer et al. Clin Pharmacol Ther 1978 | 6 | Surgical | 12 | 3 |
| Fung et al. J Clin Pharmacol 1980 | 9 | Healthy | 10 | 3 |
| Univ. of Maryland, Baltimore (ongoing) | 14 | Healthy | 11 | 2 |

Weighted Mean CL_{IV} from **Healthy subjects with PK value obtained from 2 or 3 compartmental analysis** = **33.6 L/h**

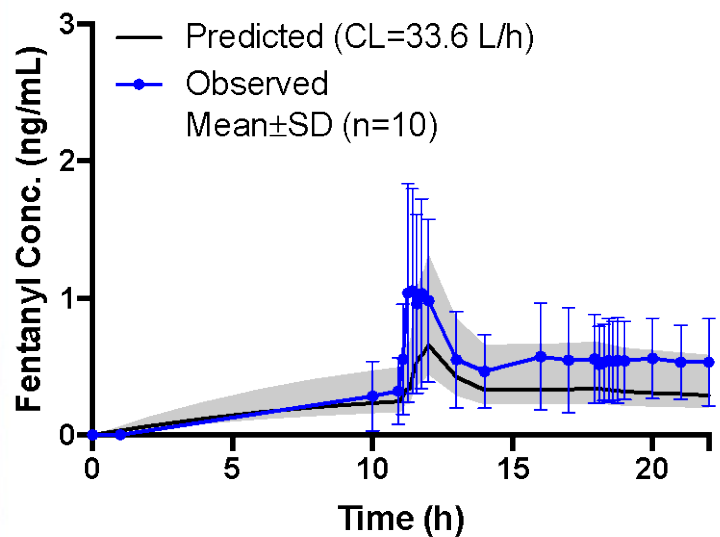
¹ Source of IV PK parameters reported in Duragesic® Package Insert



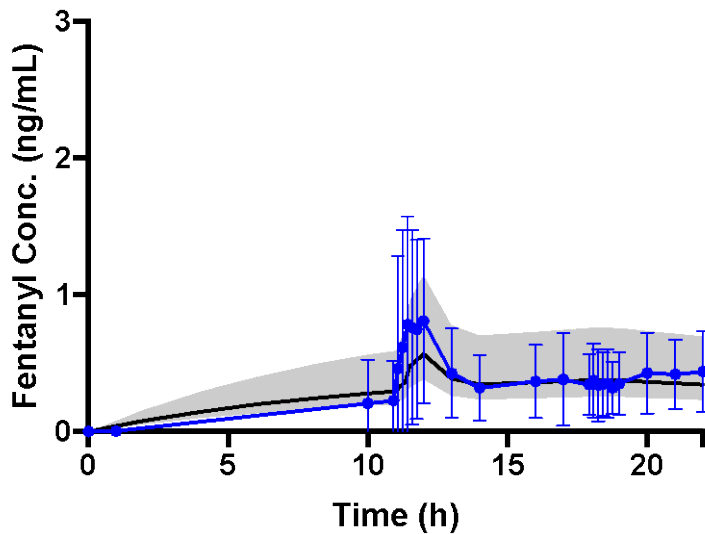
Approach I

Grey shade represents prediction range when inter-subject variability of CL = 50%

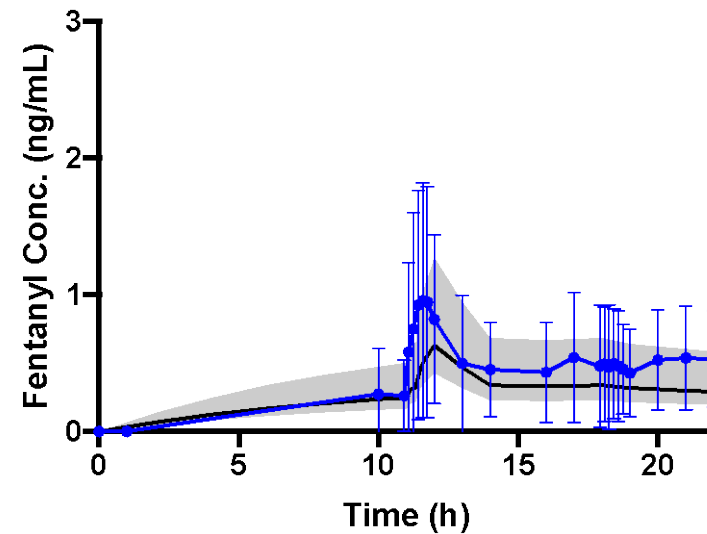
Duragesic® - Early Heat



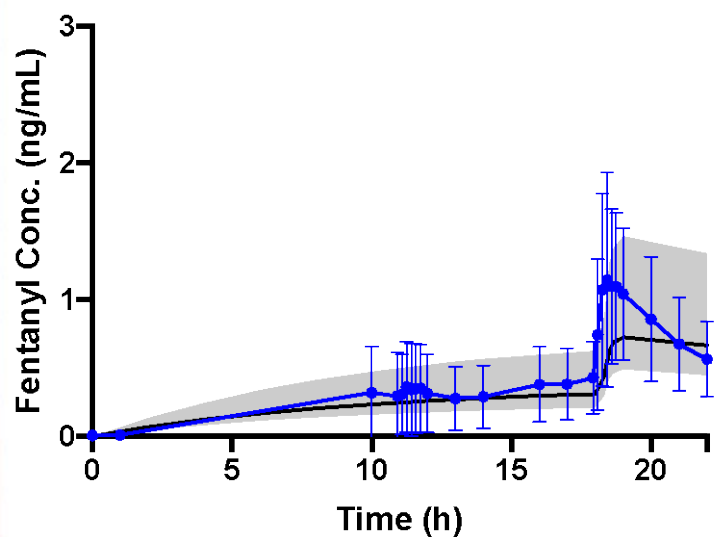
Apotex - Early Heat



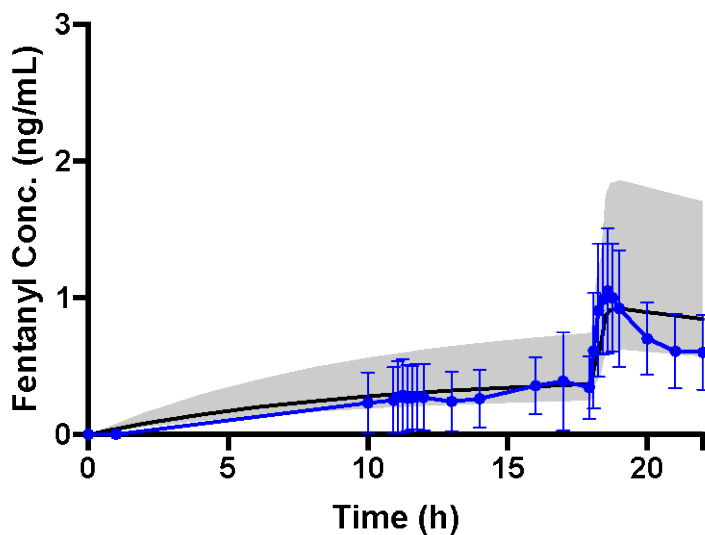
Mylan - Early Heat



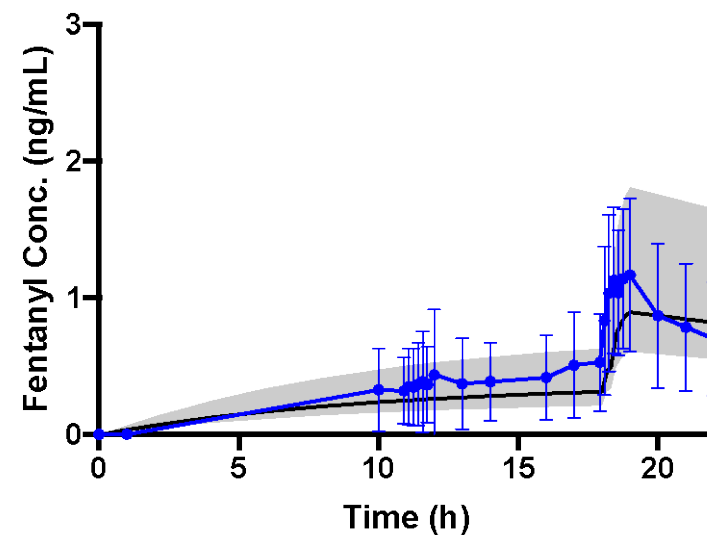
Duragesic® - Late Heat



Apotex - Late Heat



Mylan - Late Heat

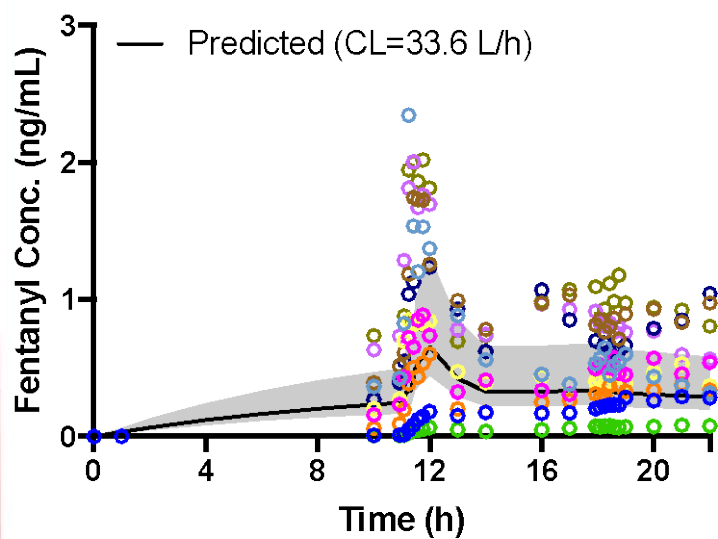




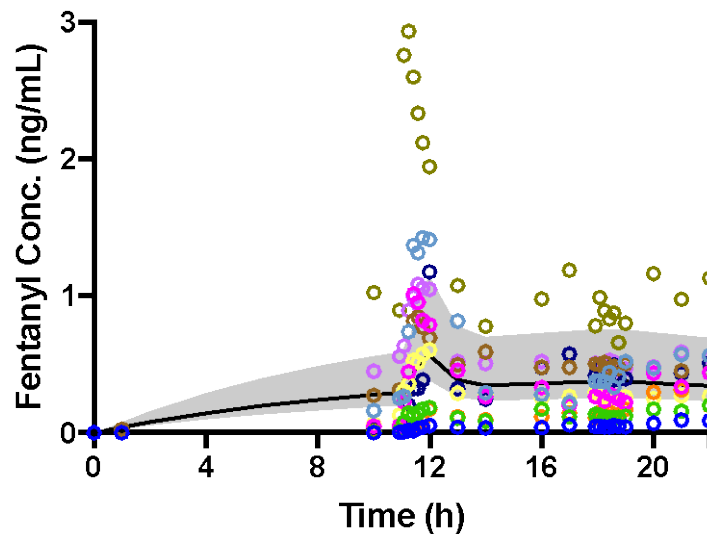
Approach I

Grey shade represents prediction range when inter-subject variability of CL = 50%

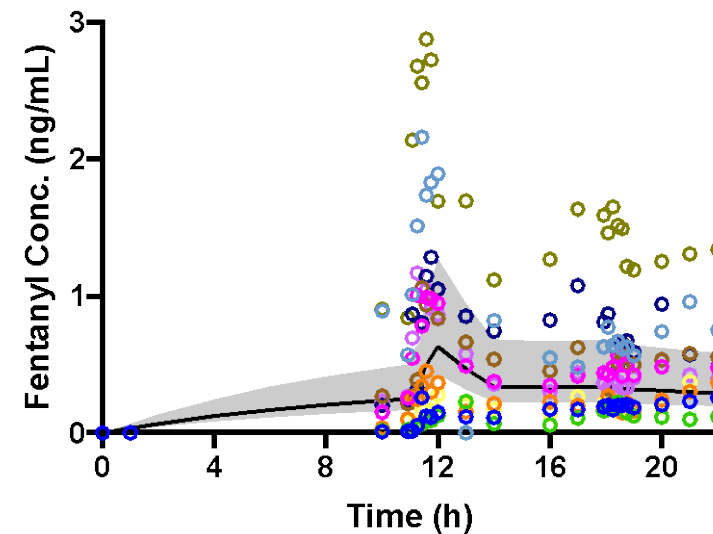
Duragesic® - Early Heat



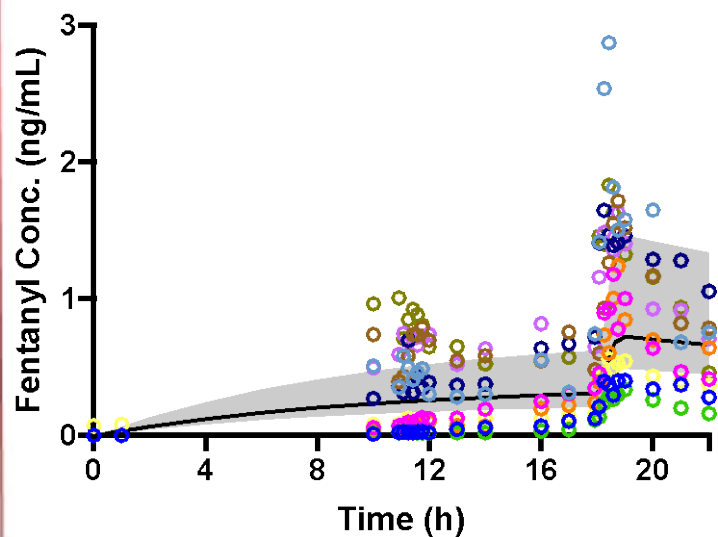
Apotex - Early Heat



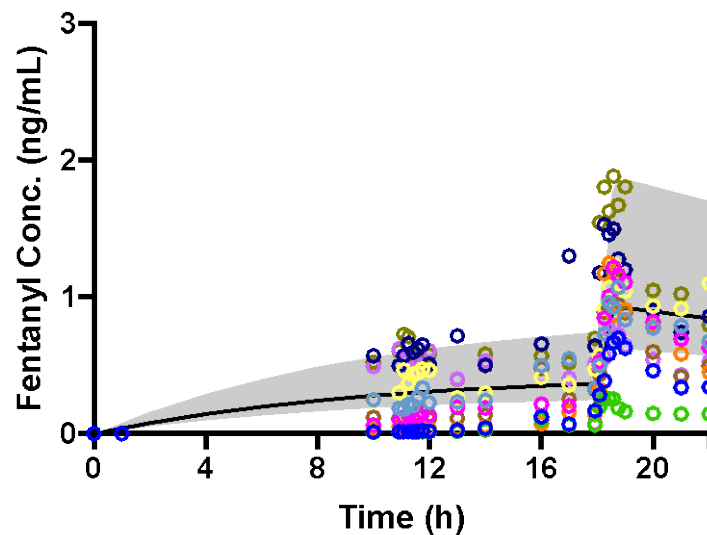
Mylan - Early Heat



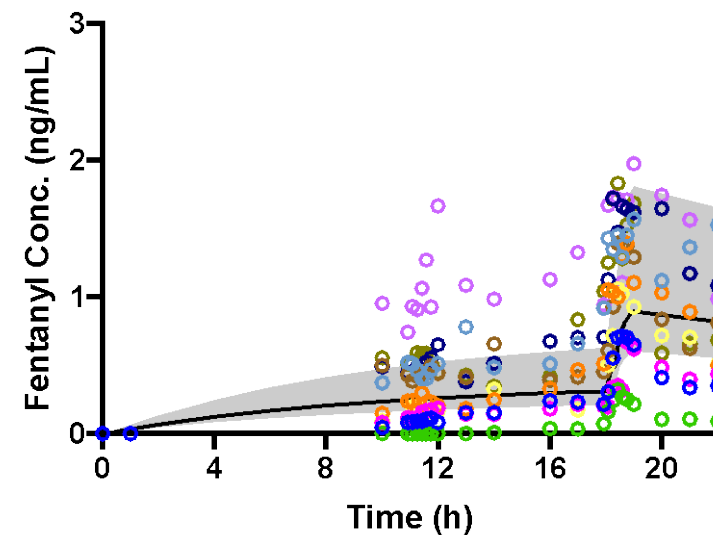
Duragesic® - Late Heat



Apotex - Late Heat



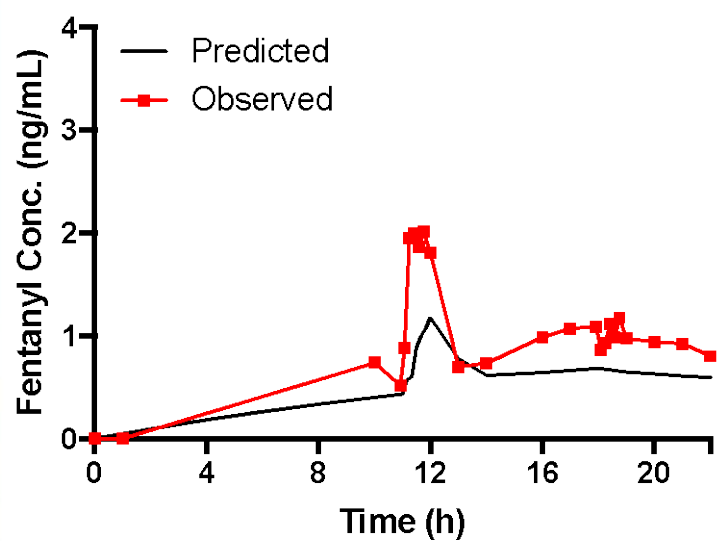
Mylan - Late Heat



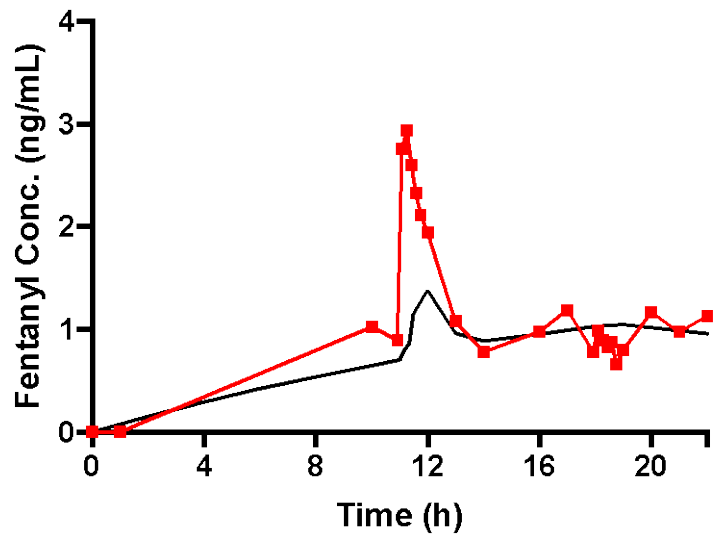


Approach I Subject TDF 024: Predicted using the subject's own F, CL_{IV} and k values

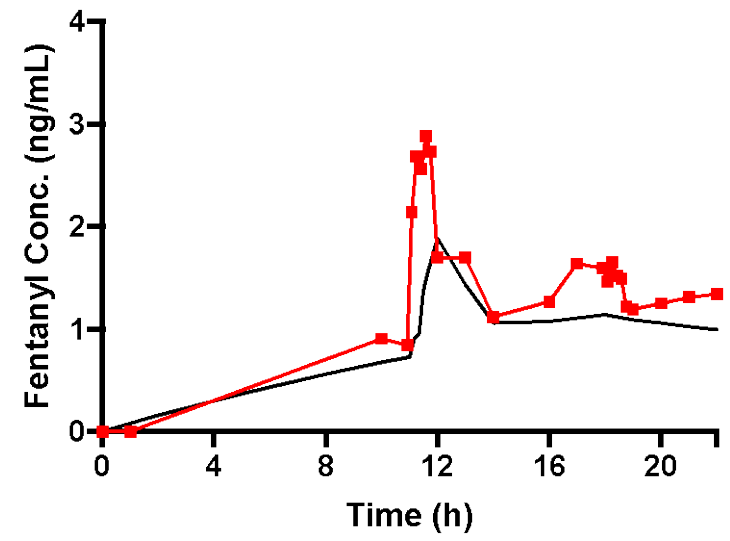
Duragesic® - Early Heat



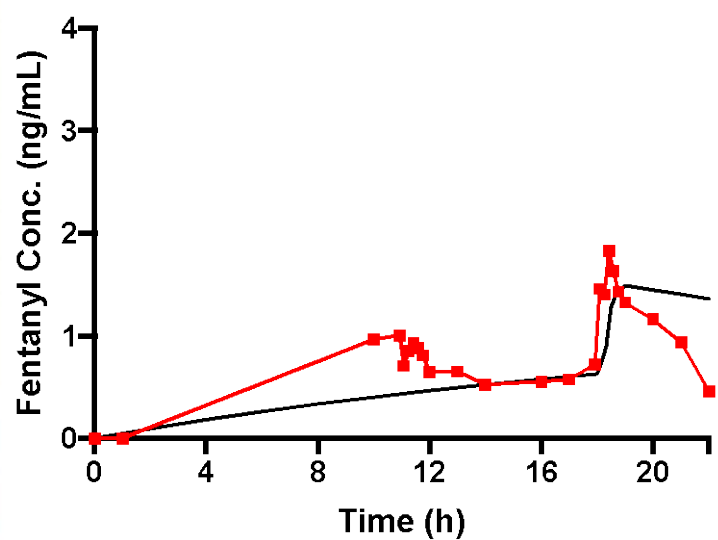
Apotex - Early Heat



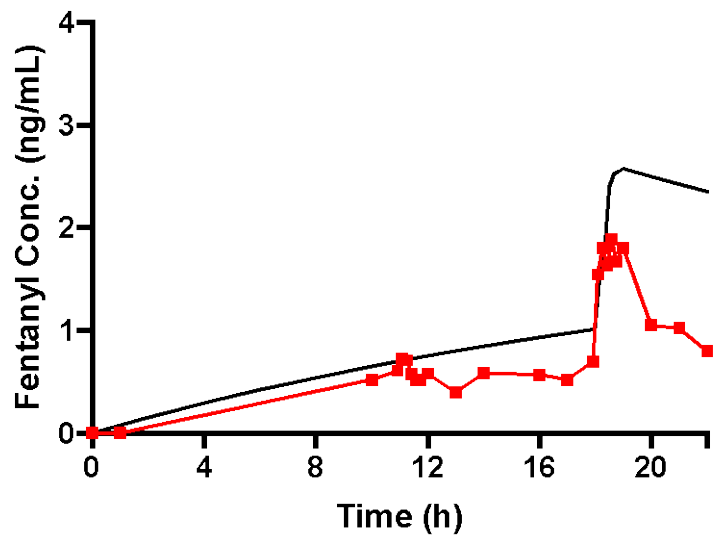
Mylan - Early Heat



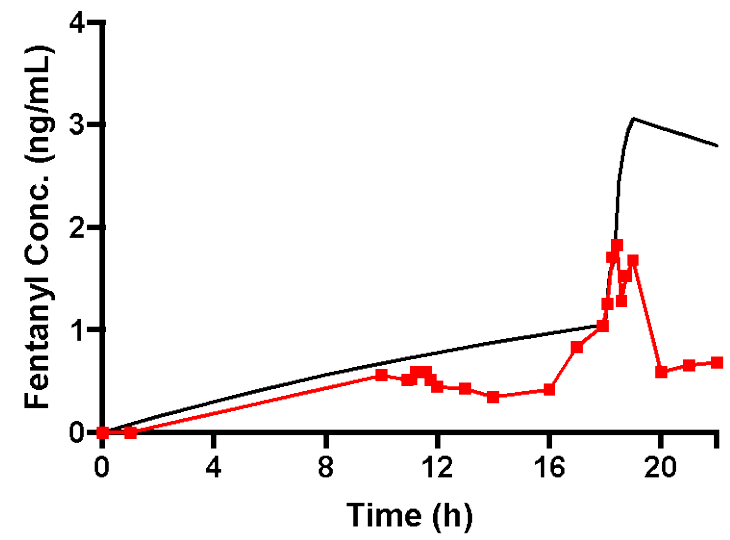
Duragesic® - Late Heat



Apotex - Late Heat



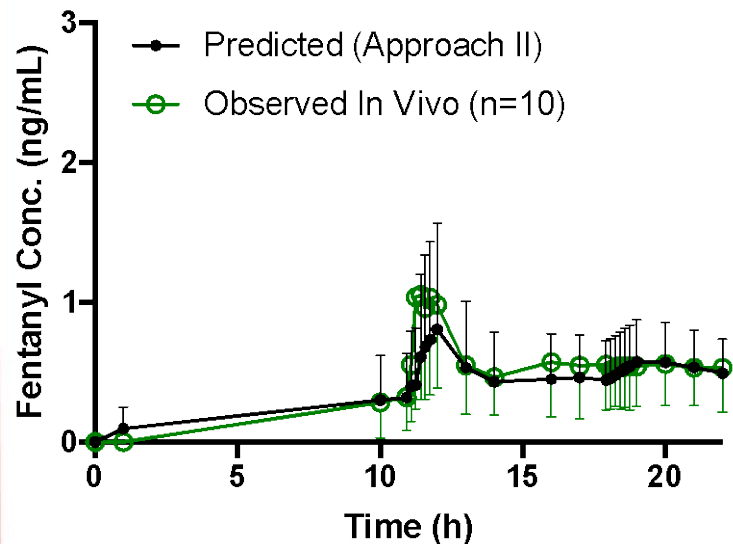
Mylan - Late Heat



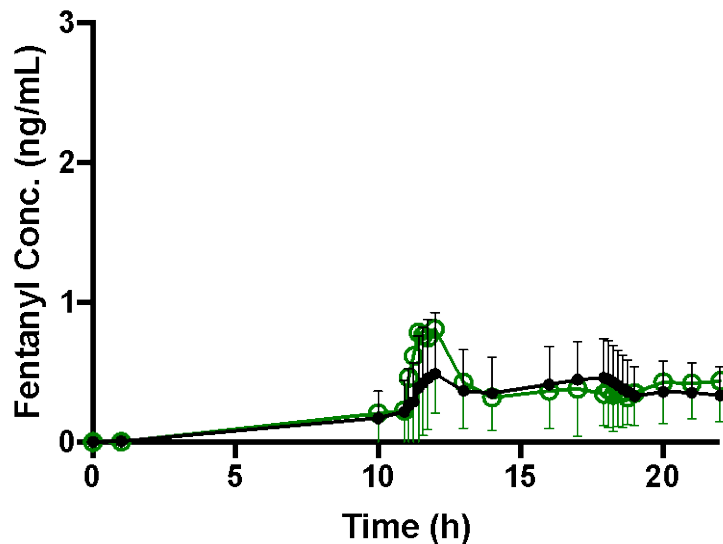


Approach II

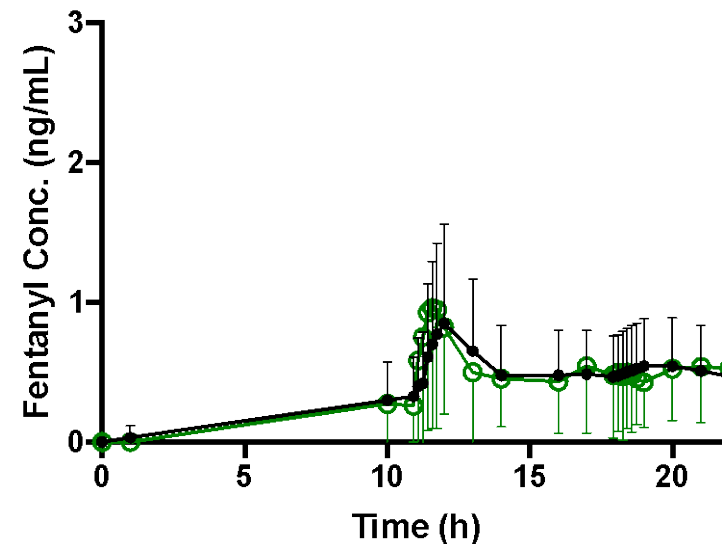
Duragesic® - Early Heat



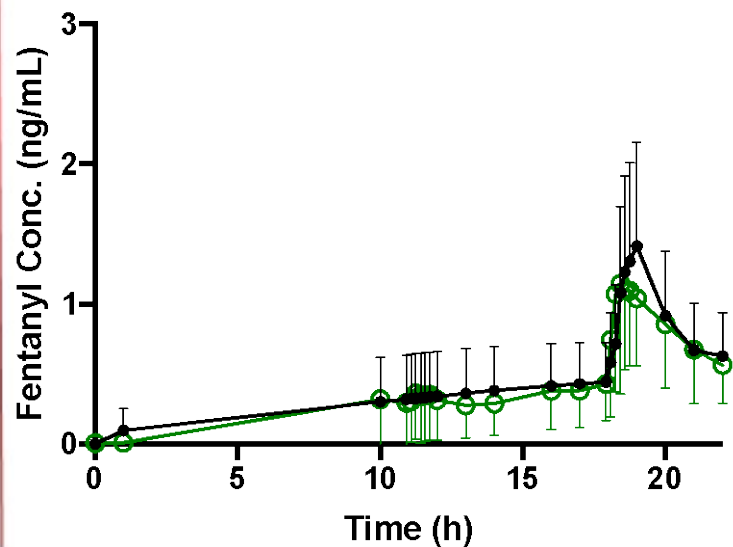
Apotex - Early Heat



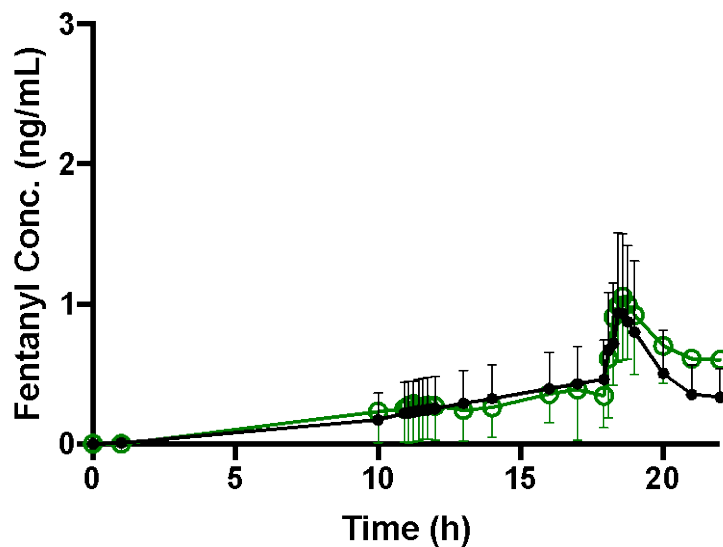
Mylan - Early Heat



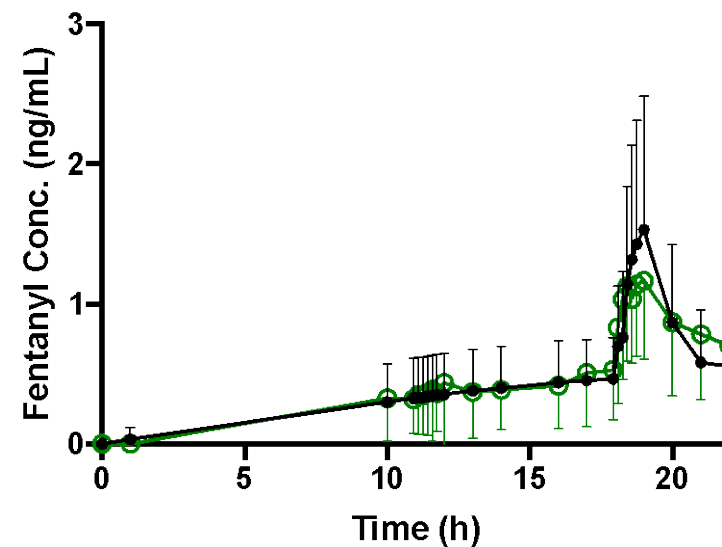
Duragesic® - Late Heat



Apotex - Late Heat



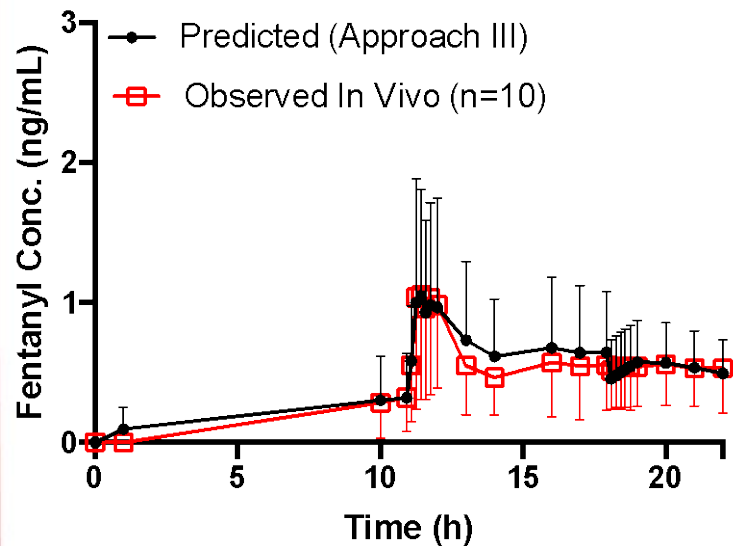
Mylan - Late Heat



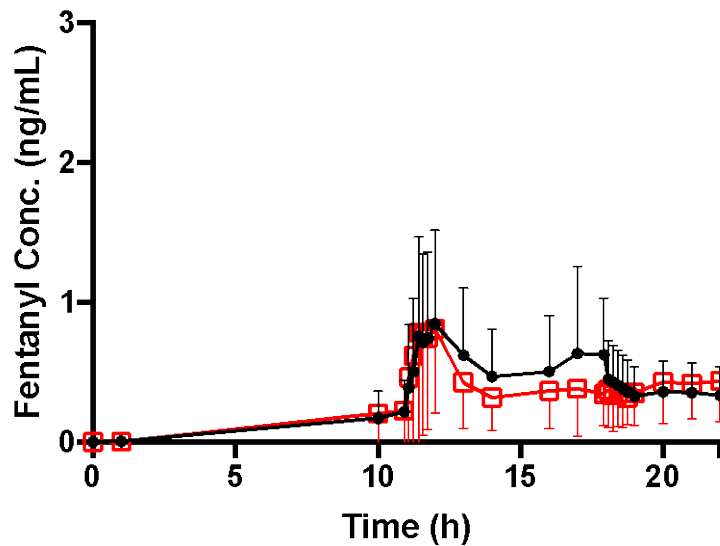


Approach III

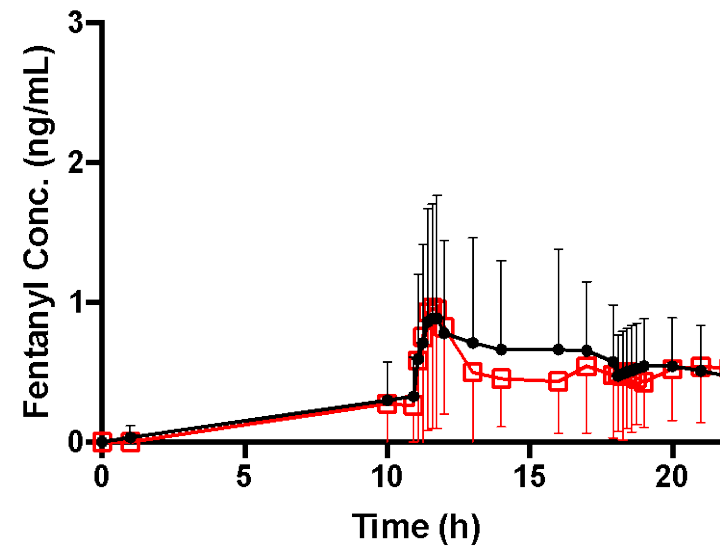
Duragesic® - Early Heat



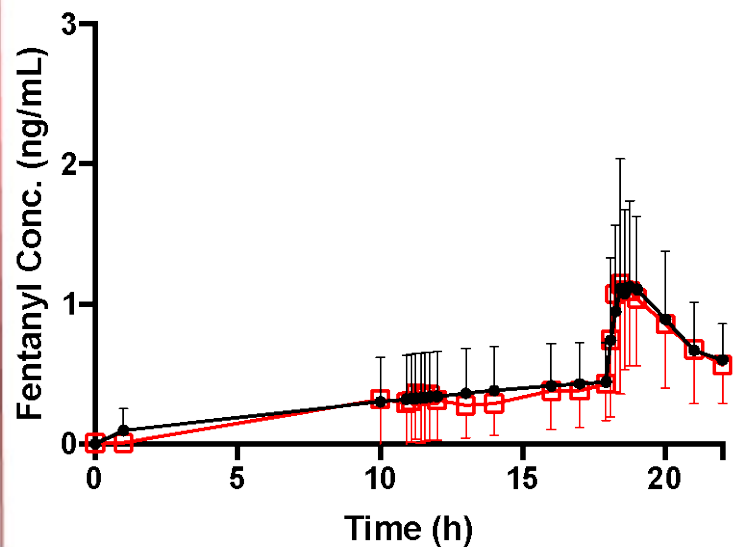
Apotex - Early Heat



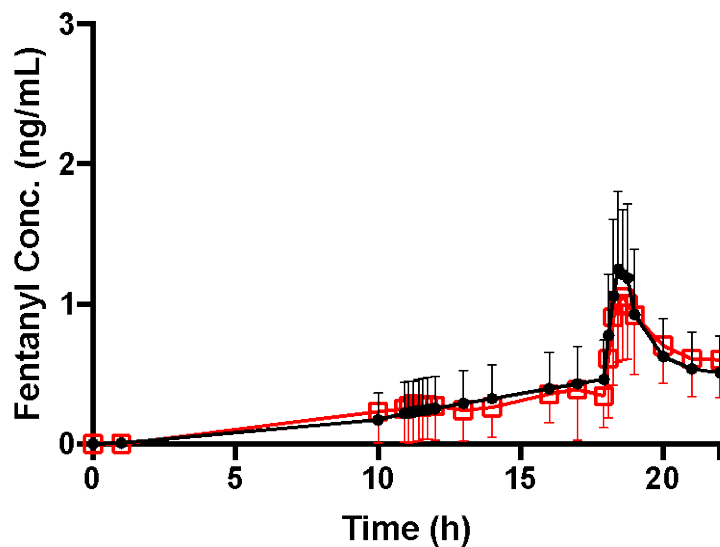
Mylan - Early Heat



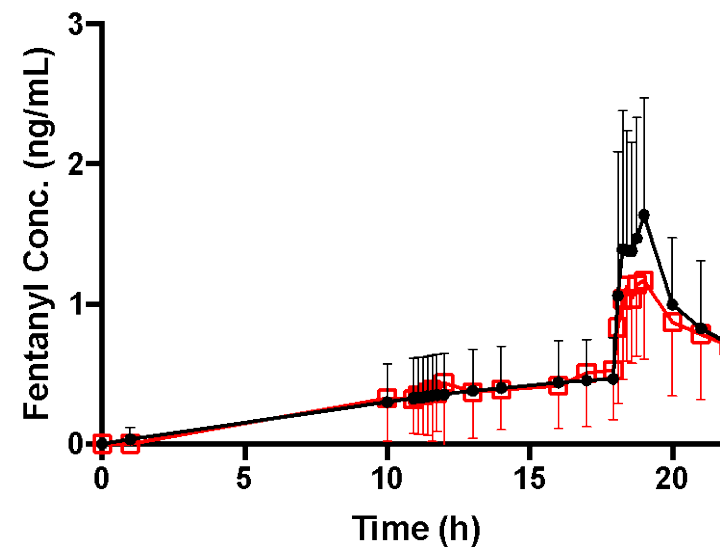
Duragesic® - Late Heat



Apotex - Late Heat



Mylan - Late Heat





% Prediction Error

| Fentanyl TDS | Duragesic® | | Apotex | | Mylan | |
|---------------------|------------|-----------|------------|-----------|------------|-----------|
| | Early Heat | Late Heat | Early Heat | Late Heat | Early Heat | Late Heat |
| Approach I | | | | | | |
| Total AUC | 31.7 | 17.5 | 4.0 | 19.3 | 24.3 | 18.4 |
| C _{max} | 37.7 | 36.8 | 29.8 | 12.4 | 34.1 | 23.2 |
| Approach II | | | | | | |
| Total AUC | 3.3 | 13.1 | 10.2 | 11.8 | 5.1 | 0.6 |
| C _{max} | 23.4 | 23.6 | 39.6 | 11.2 | 11.4 | 31.5 |
| Approach III | | | | | | |
| Total AUC | 15.2 | 10.1 | 11.9 | 0.8 | 18.1 | 8.3 |
| C _{max} | 0.5 | 2.3 | 4.4 | 18.7 | 7.7 | 40.5 |



Conclusions – Fentanyl

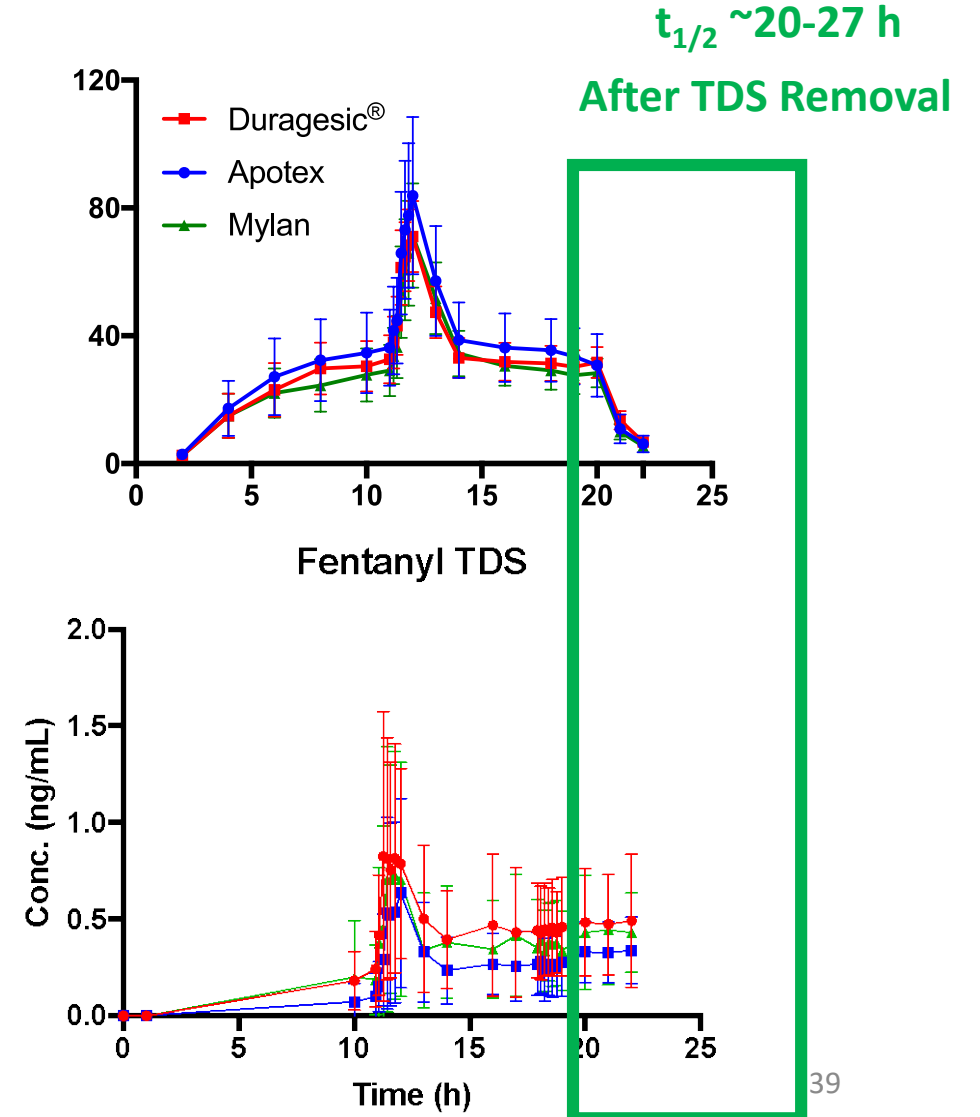
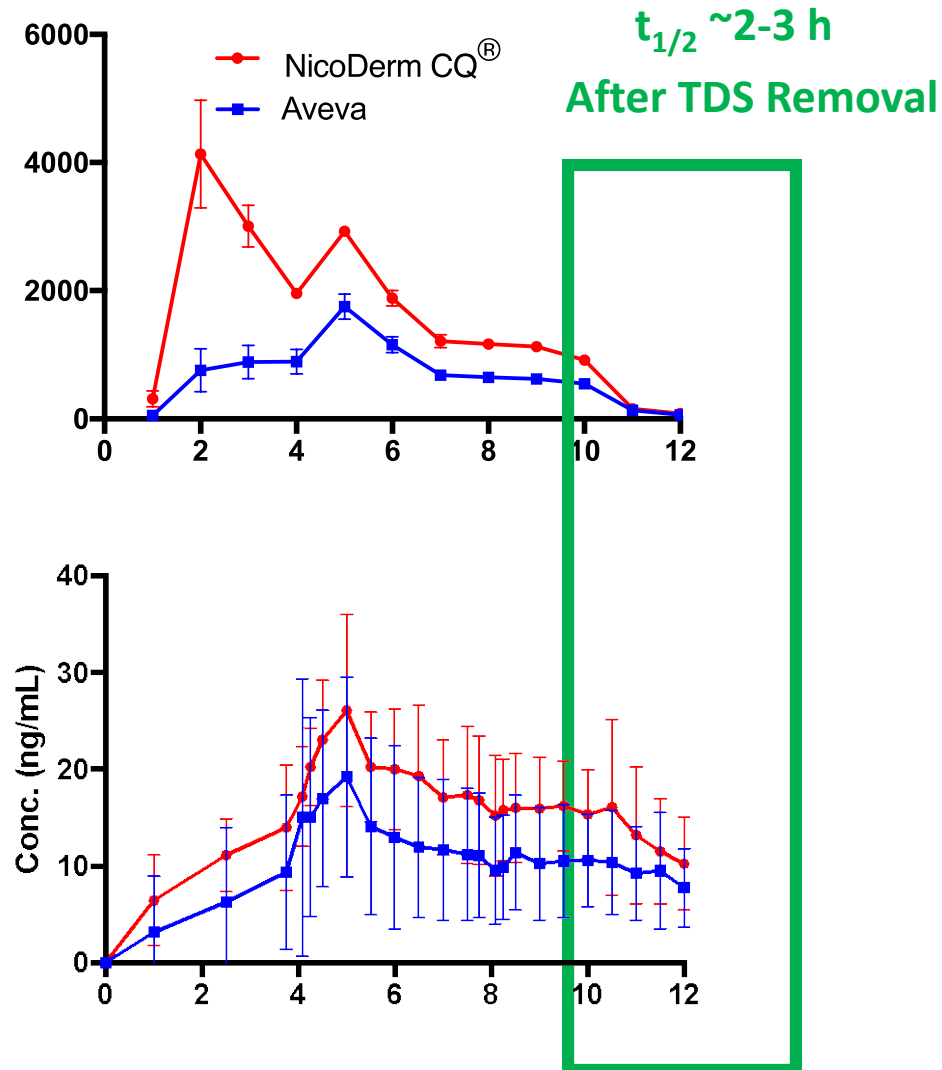
- Early vs. Late Heat effect comparable both *in vitro* and *in vivo*
- Heat effect on three differently formulated TDS comparable both *in vitro* and *in vivo*
- However, *in vivo* heat effect seemed to be higher compared to the *in vitro* heat effect
- Preliminary IVIVCs between IVPT and clinical human PK studies under the matched study designs
 - ⇒ Not as predictive compared to nicotine...



Why??



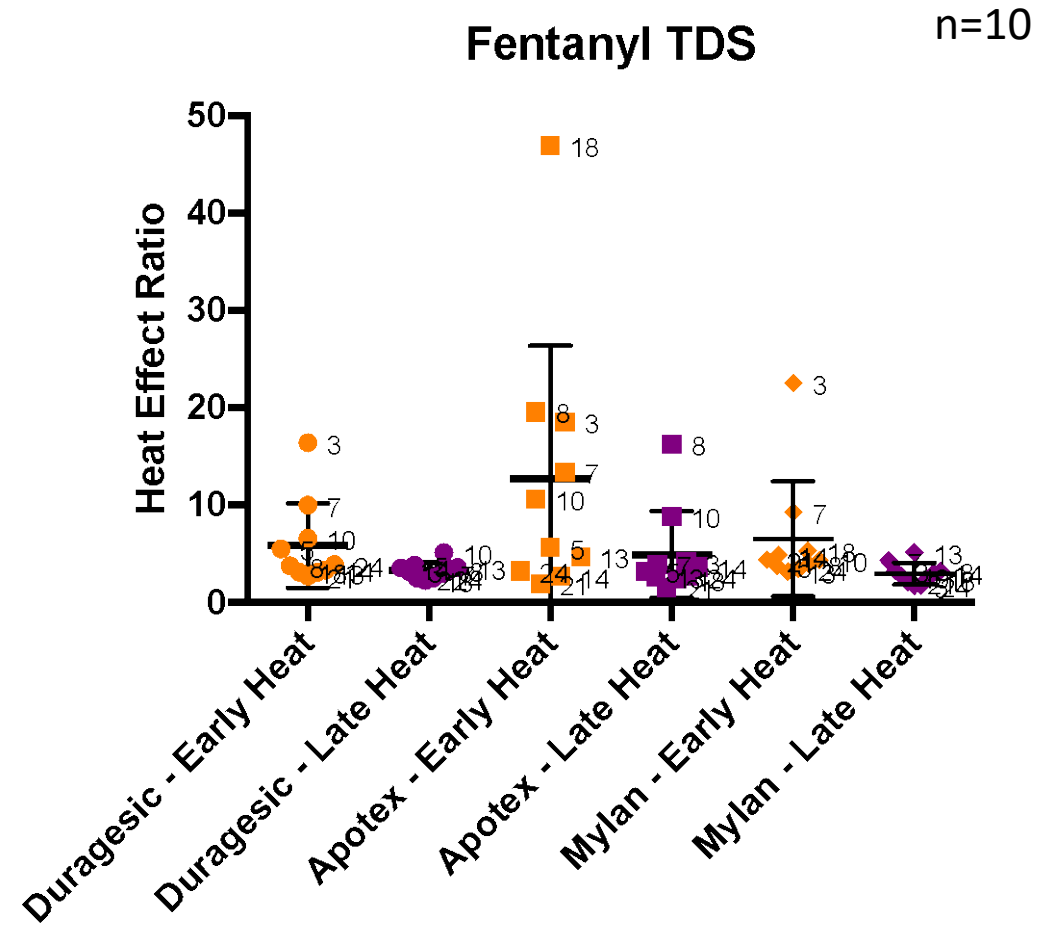
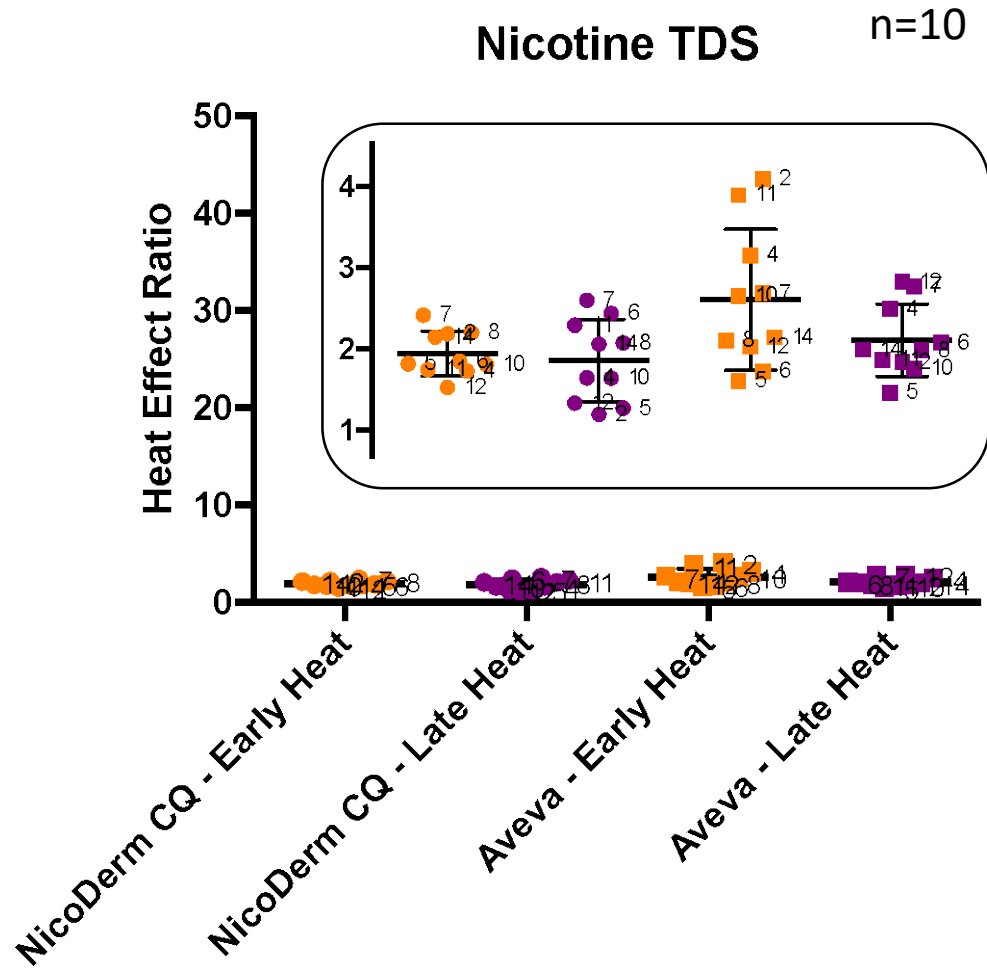
1. Lipophilicity of Fentanyl





2. High Inter-subject Variability of Fentanyl

Heat Effect Ratio was determined by the ratio of the C_{max} during the 3h window and the concentration immediately before heat application





Conclusions - IVIVC

- Three approaches were evaluated to demonstrate a preliminary Level A IVIVC (IVIVR) for TDS
- Good preliminary IVIVC demonstrated for nicotine TDS, including heat effect
- Weaker preliminary IVIVC found for fentanyl TDS
 - Limitation of mimicking drug reservoir in skin layers, microcirculation and subcutaneous tissue *in vitro*
 - High inter-subject variability for fentanyl (+ Lack of reliable PK parameters)

Take Home Messages

- An *in vitro* heat effect study may be able to predict the *in vivo* heat effect for some drugs, following an IVIVC validation
- For certain drugs, an *in vivo* heat factor may need to be determined
- Heat effects are drug molecule and formulation excipient dependent---Diclofenac formulation data not shown
- Patches are not the only topical products affected by heat



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Co-PIs

Dr. Hazem Hassan (UMB)



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Dr. Annette L. Bunge

Dr. Richard H. Guy

Dr. Tom Franz

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Dr. Wilbur Chen

Dr. Jeff Fink

Melissa Billington

UMB GCRC nurses

Clinical Study Participants

Past & Current Lab Members

Contributors to the work presented:

- Dr. Mingming Yu (LC/MS/MS)
- Soo Hyeon Shin (Fentanyl, nicotine, acyclovir, diclofenac)
- Sherin Thomas (Lidocaine, buprenorphine, diclofenac)
- Dana Hammell (Lab Manager and Document Control)
- Dani Fox (Clinical Coordinator)
- Sagar Shukla (Lidocaine)
- Paige Zambrana (Sunscreens & glucose monitoring)
- Qingzhao Zhang (Metronidazole & rivastigmine)
- Past: Juliana Quarterman
- Dr. Inas Abdallah

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- Dr. Priyanka Ghosh
- Dr. Caroline Strasinger



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Back-Up



Methods: Skin Preparation

- Fresh human skin samples obtained post abdominoplasty surgery
- Dermatomed to ~250 microns
- Frozen until the day of experiment

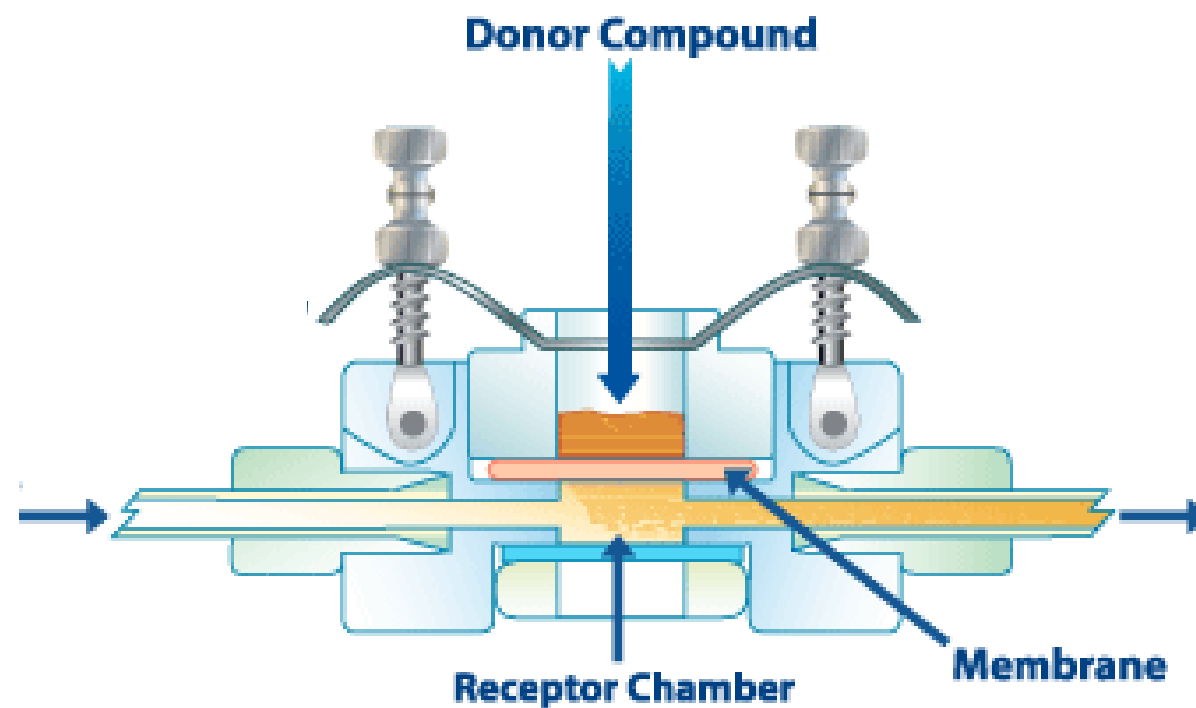
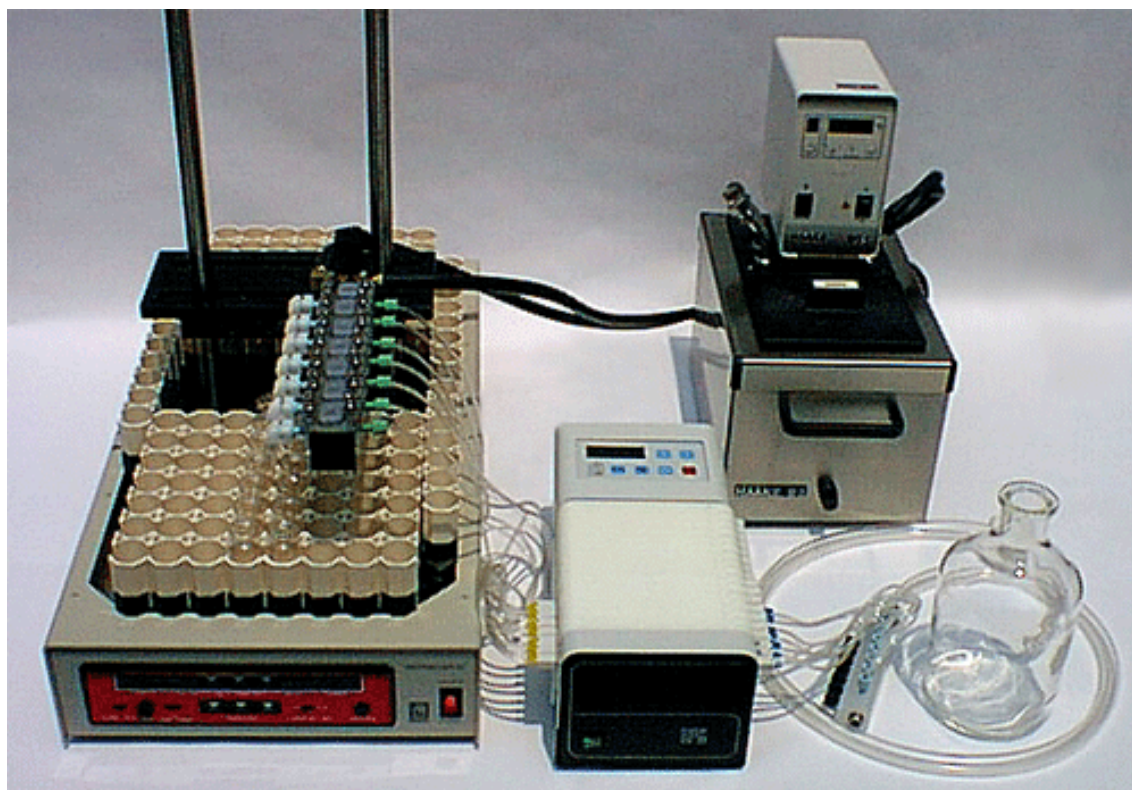


Image obtained from the Stinchcomb Lab's SOP



IVPT Setup

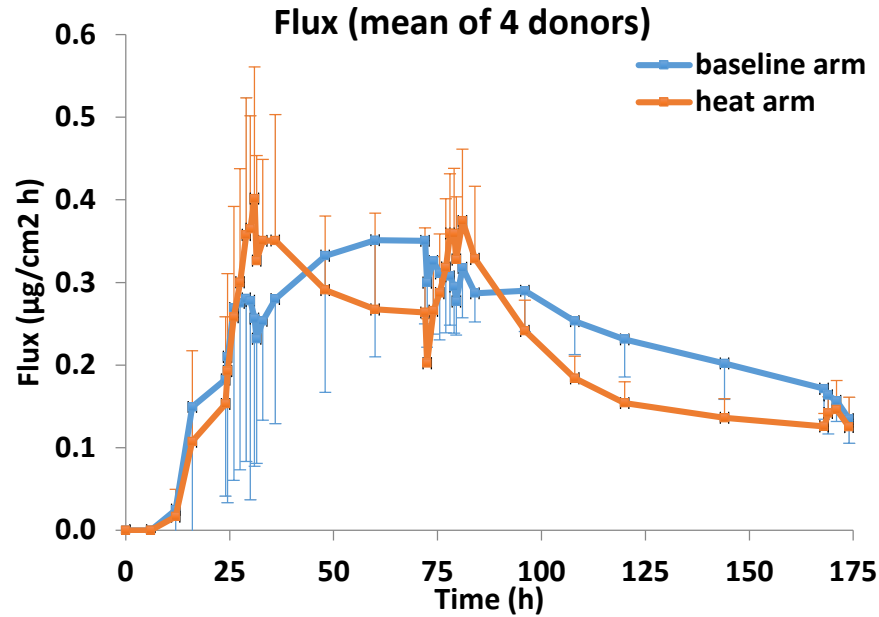
- In-line flow-through diffusion system
- Permeation area of 0.95 cm²





Buprenorphine Patch

Mean (\pm SD) *in vitro* flux n=4 donors



Human skin donor 4 (n=4 replicates per arm)

Heat Arm:

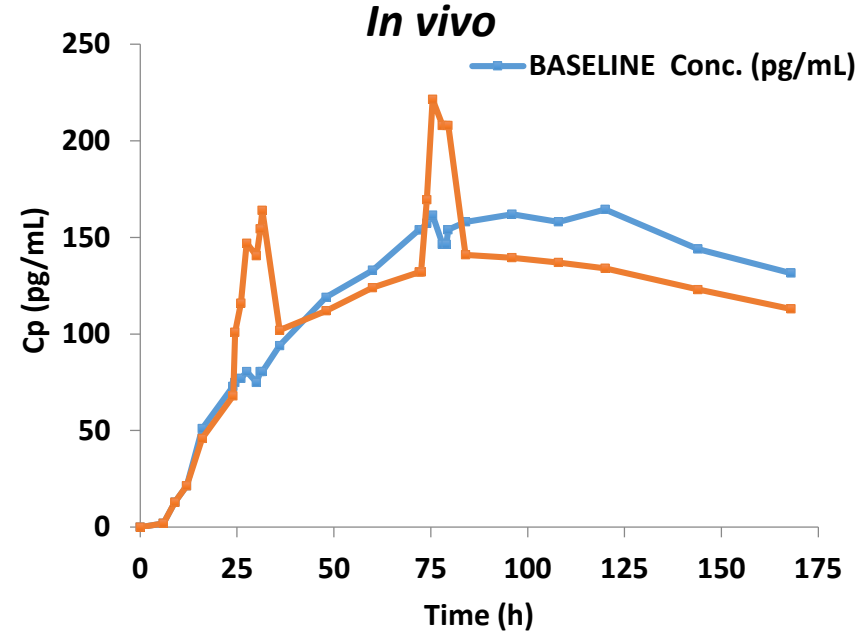
Early heat-heat applied from 24 to 31 h (every 2 h with 30 min gap)

Late heat- heat applied from 72 to 79 h (every 2 h with 30 min gap)

Patch off at 168 h

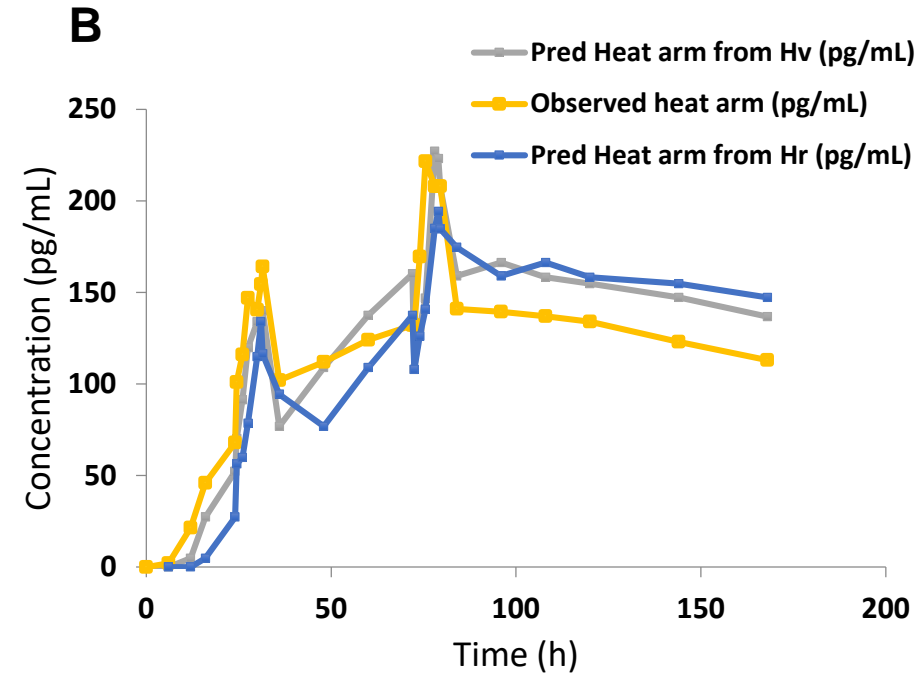
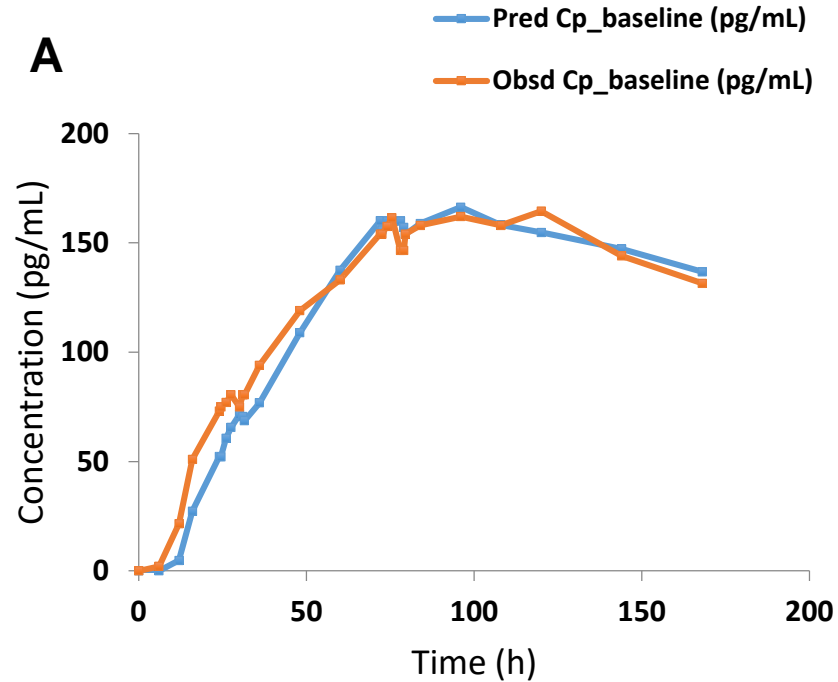
Mean *in vivo* concentration n=19/20 subjects

(values from graph grabbing software for graph taken from *Clinical Pharmacology and Biopharmaceutics Review* document for Butrans[®] available at Drugs@FDA.)





Buprenorphine Patch



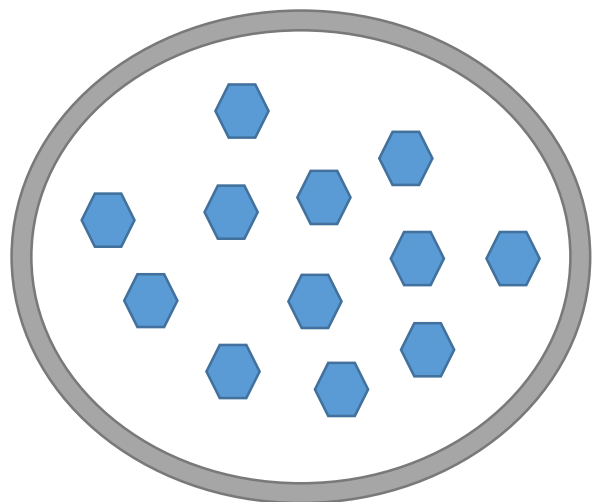
Plot for observed and predicted concentration versus time profiles for baseline arm (A) and heat arm (B)

Hv = *in vivo* heat factor **Hr = *in vitro* heat factor**

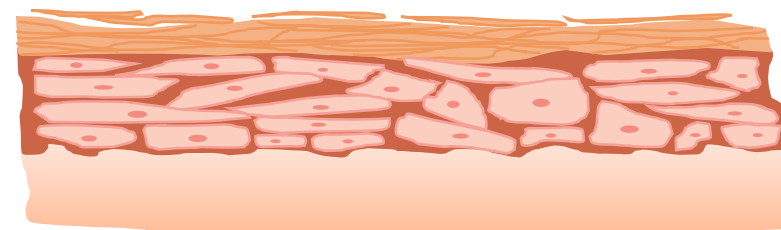
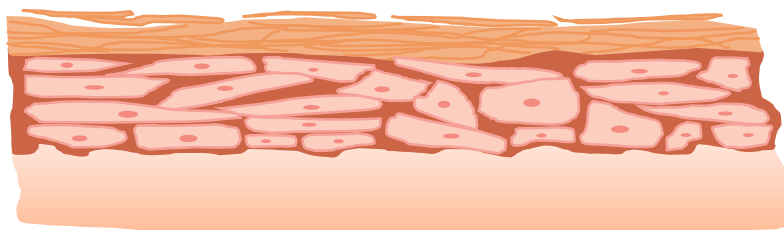
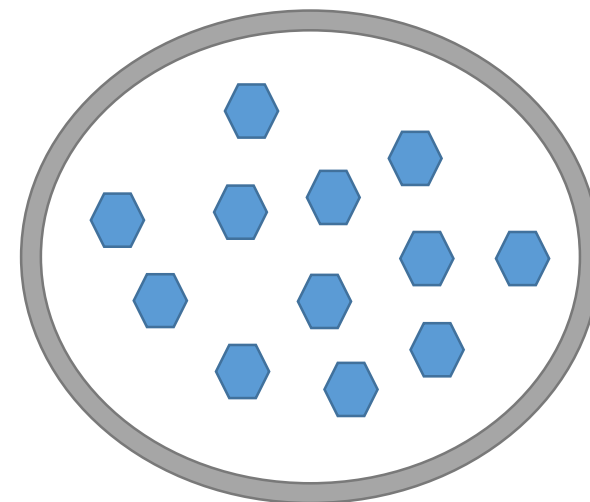


Influence of Heat on Percutaneous Absorption

1) ↑ Diffusivity of Drug from its Vehicle

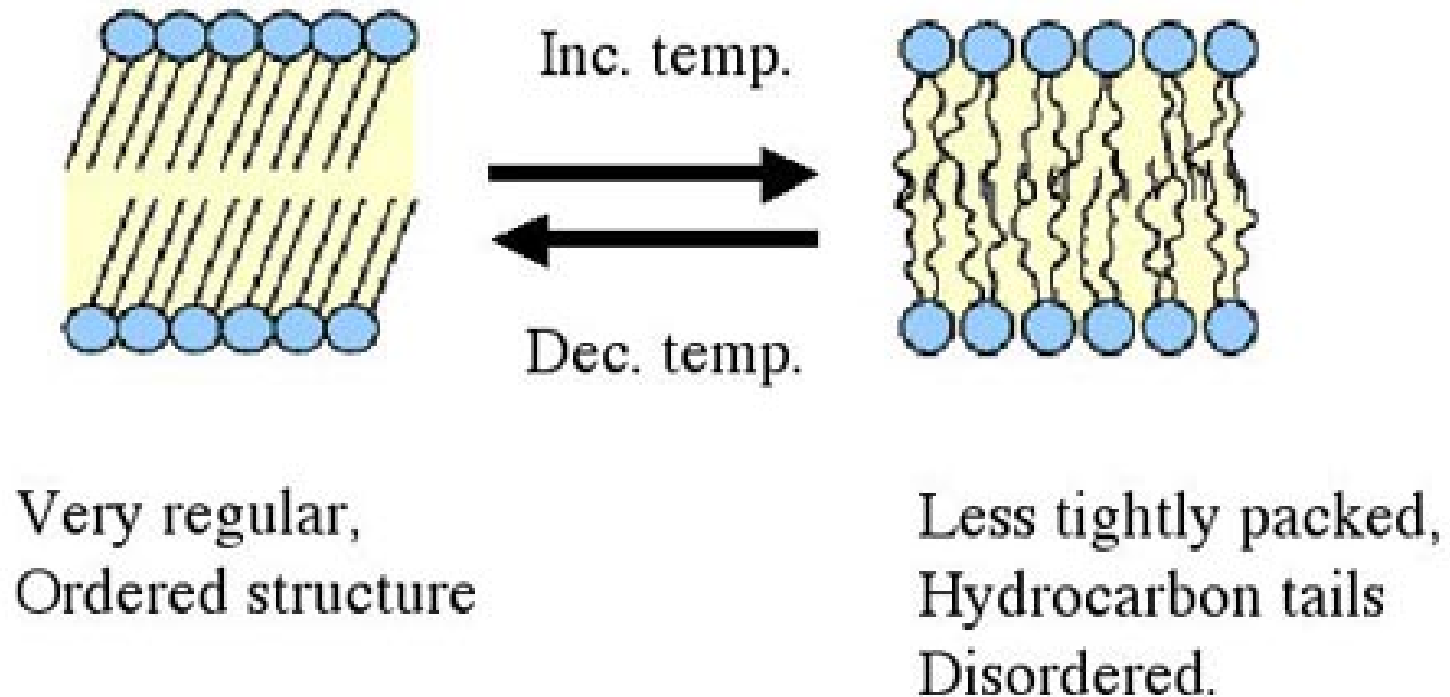


+ Heat →



Influence of Heat on Percutaneous Absorption

2) ↑ Fluidity of Stratum Corneum Lipids

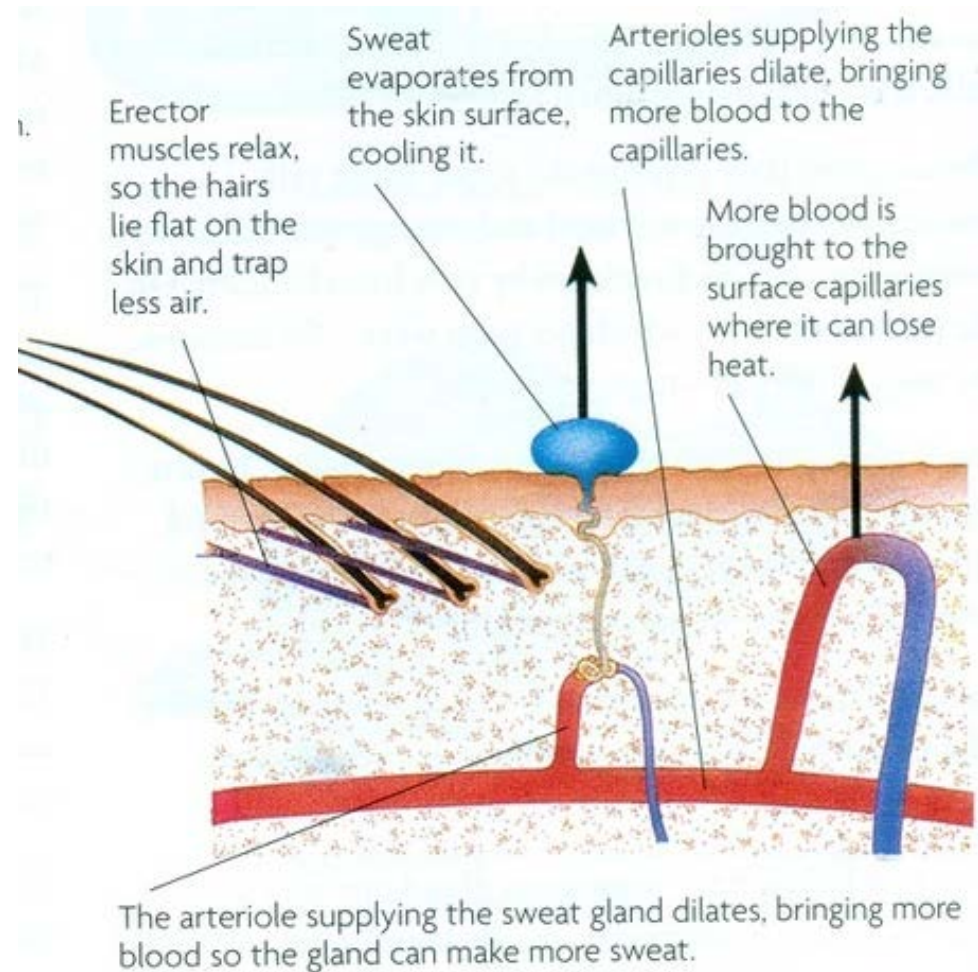


Influence of Heat on Percutaneous Absorption

3) ↑ Cutaneous Vasodilation

Body temperature regulation

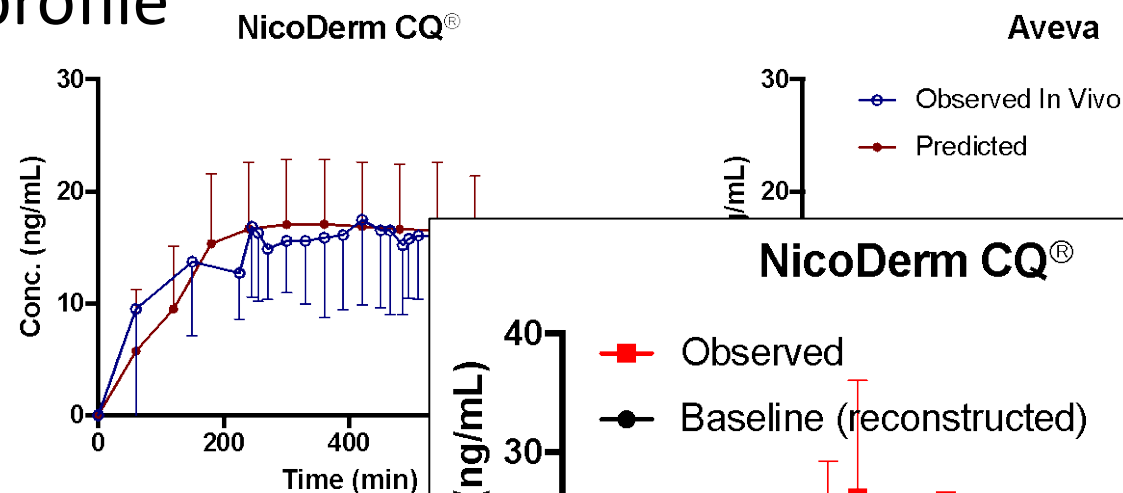
When the body is too hot





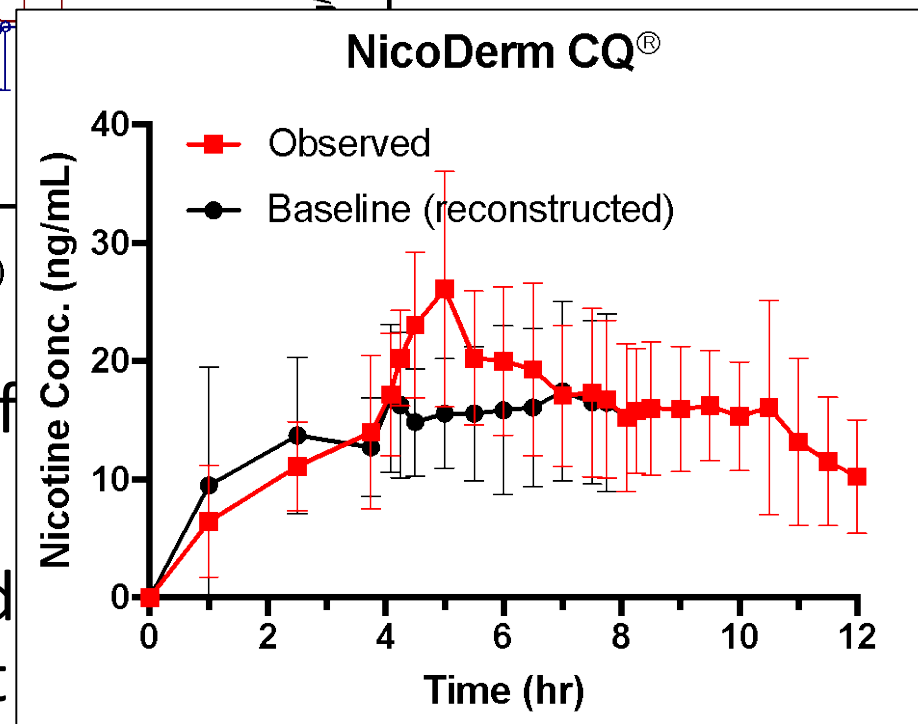
IVIVC: Level A (Approach II & III)

6) Convolute the predicted fraction of drug absorption vs time profile to obtain conc. vs time profile



7) Calculate *in vivo* heat factor (H_{ij}): ratio the reconstructed *in vivo* baseline prof exposure

8) Apply heat factor (H_i or H_{ij}) to the pred complete the prediction with the heat



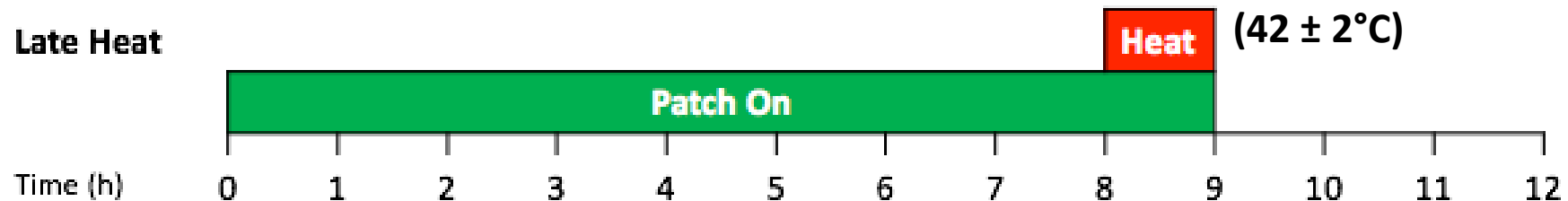
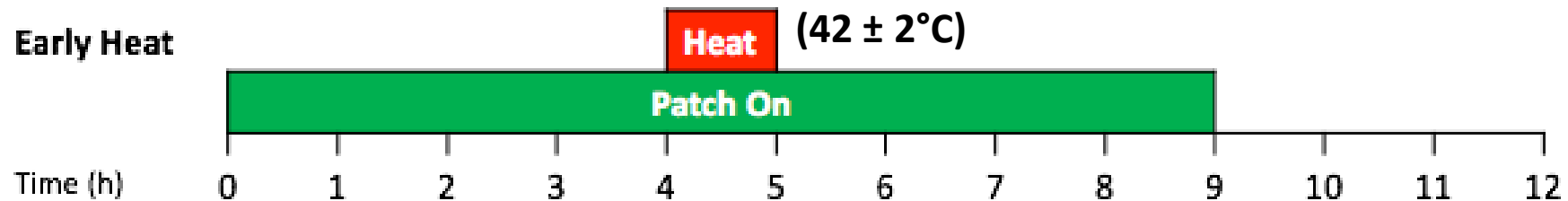


Nicotine TDS, 14 mg/24 h

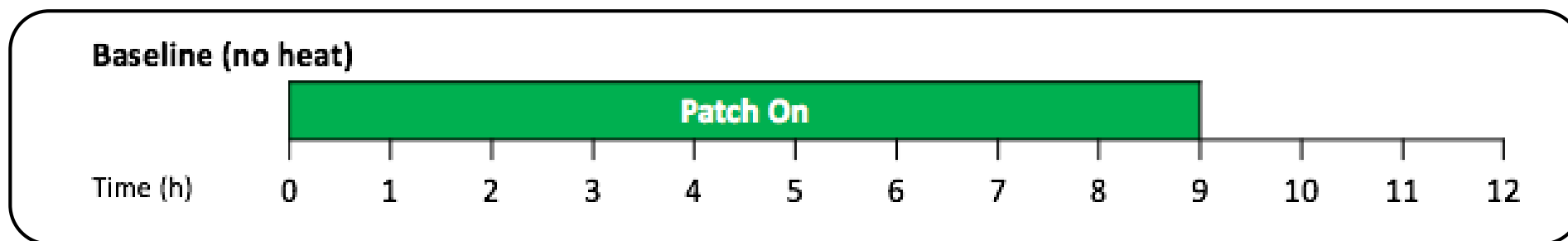
| | NicoDerm CQ[®] | Aveva |
|--|---|-------------------|
| TDS size (cm²) | 15.75 | 20.12 |
| Drug content (mg) | Not available | Not available |
| Rate/Area (µg/h/cm²) | 37 | 29 |
| Adhesive | Polyisobutylene | Acrylate/Silicone |
| Other Inactive ingredients | Ethylene vinyl acetate-copolymer, high density polyethylene between clear polyester backing | Polyester |



Study Designs


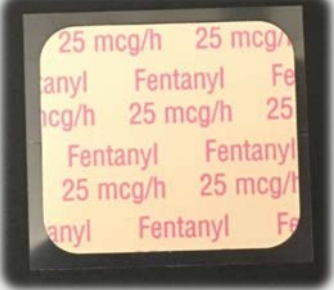



In Vitro Only



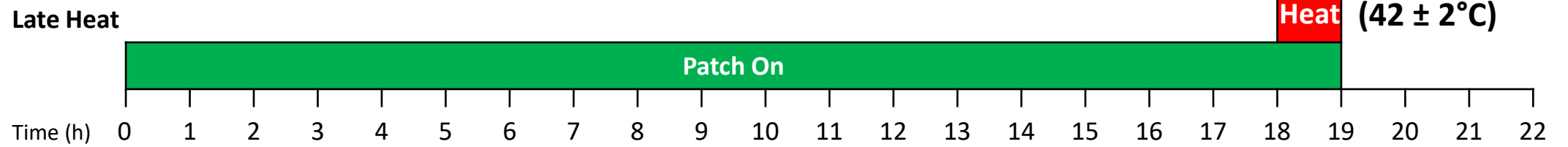
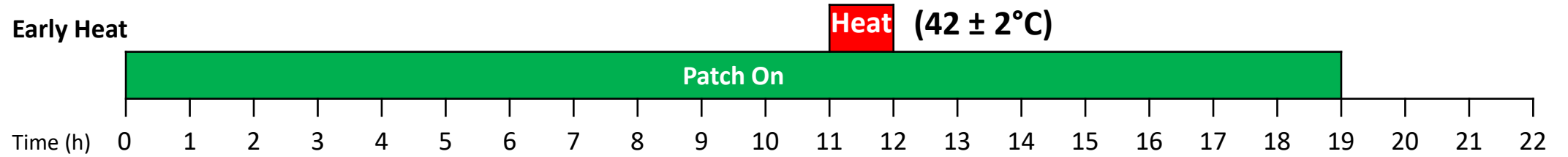


Fentanyl TDS, 25 $\mu\text{g}/\text{h}$

| | Duragesic® | Apotex | Mylan |
|-----------------------------|---|--|--|
| Drug Load (mg) | 4.20 | 2.76 | 2.55 |
| Size (cm ²) | 10.50 | 10.70 | 6.25 |
| Thickness (μm) | 110 | 200 | 190 |
| Adhesive | Polyacrylate | Polyisobutene | Silicone |
| Other Inactive Ingredients | Polyester/ ethyl vinyl acetate backing film, copovidone | Isopropyl myristate, octyldodecanol, polybutene, polyethylene/ aluminum/ polyester film backing | Dimethicone NF, polyolefin film backing |
| Appearance |  |  |  |

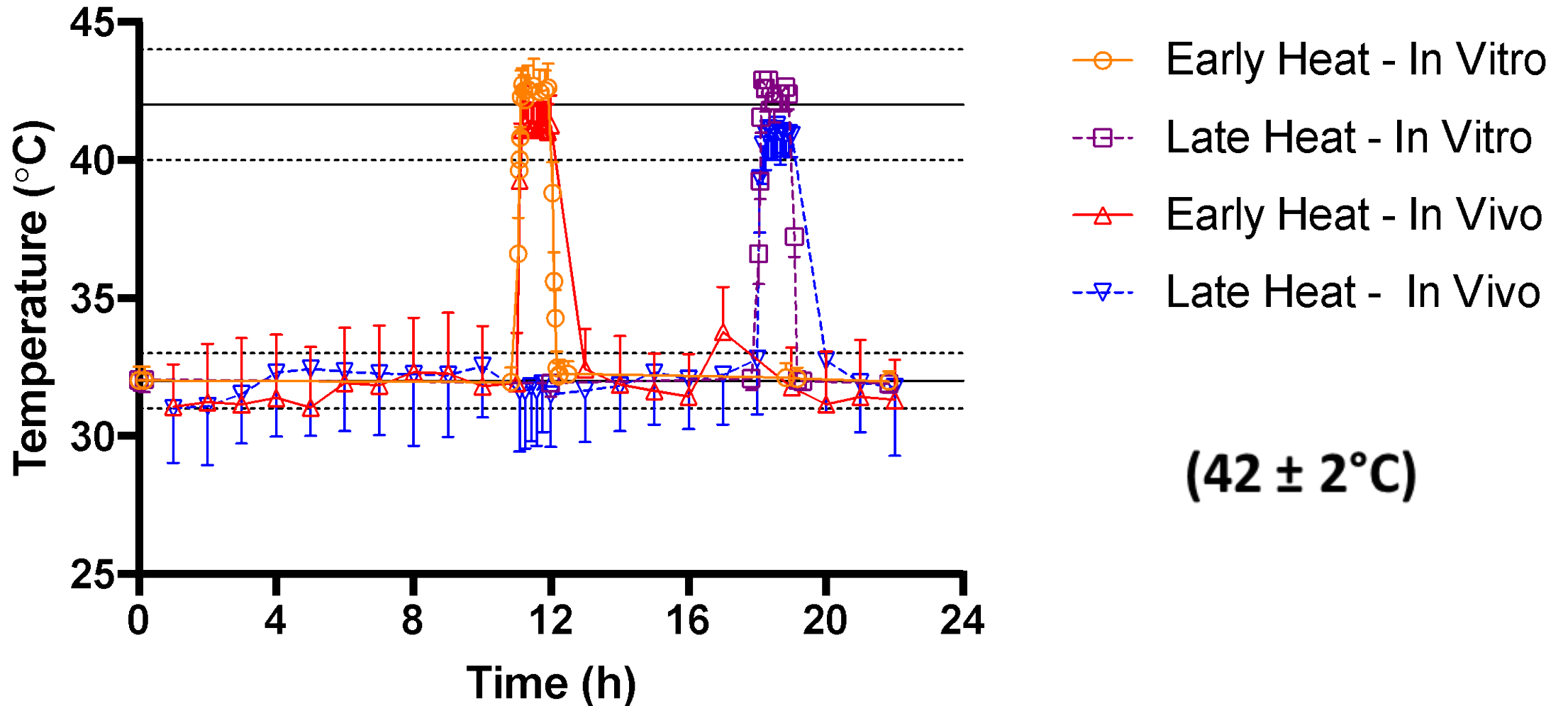


Study Designs



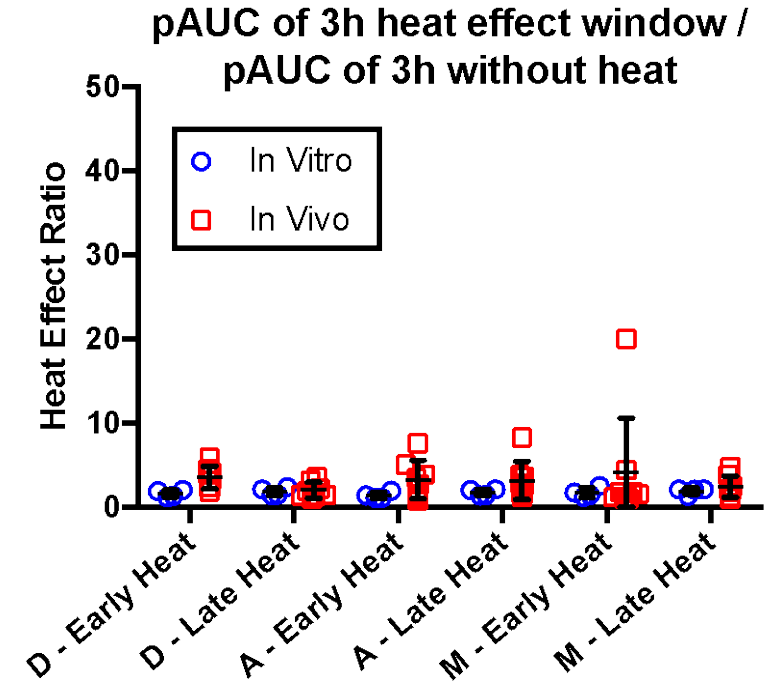
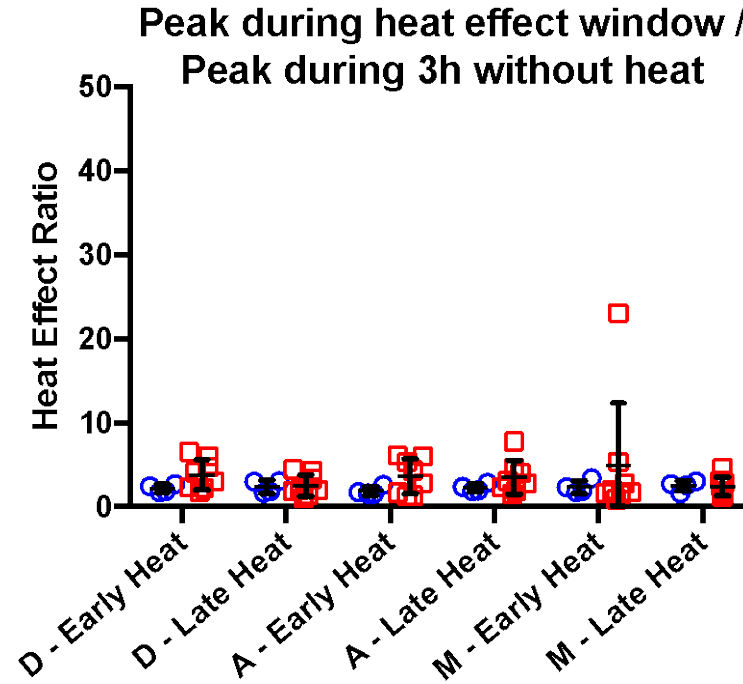
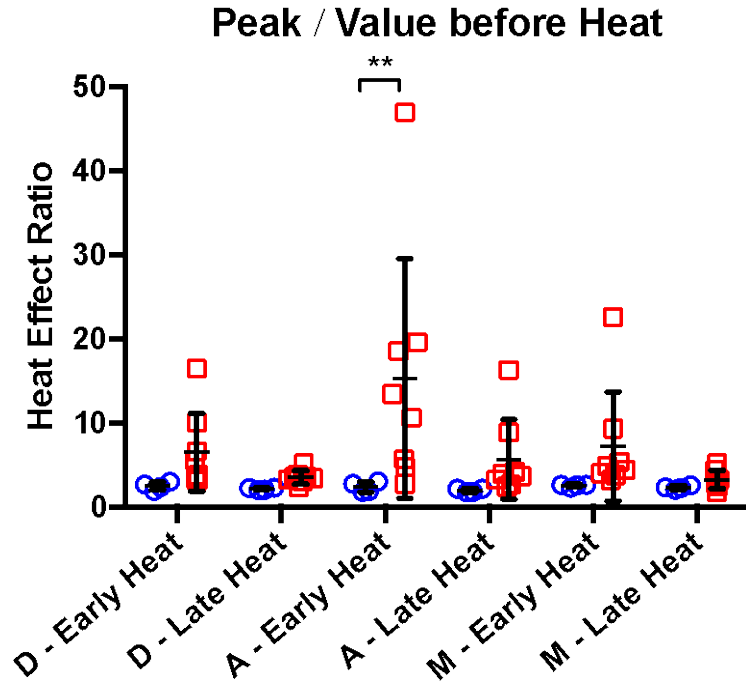


Temperature: *In Vitro* & *In Vivo*





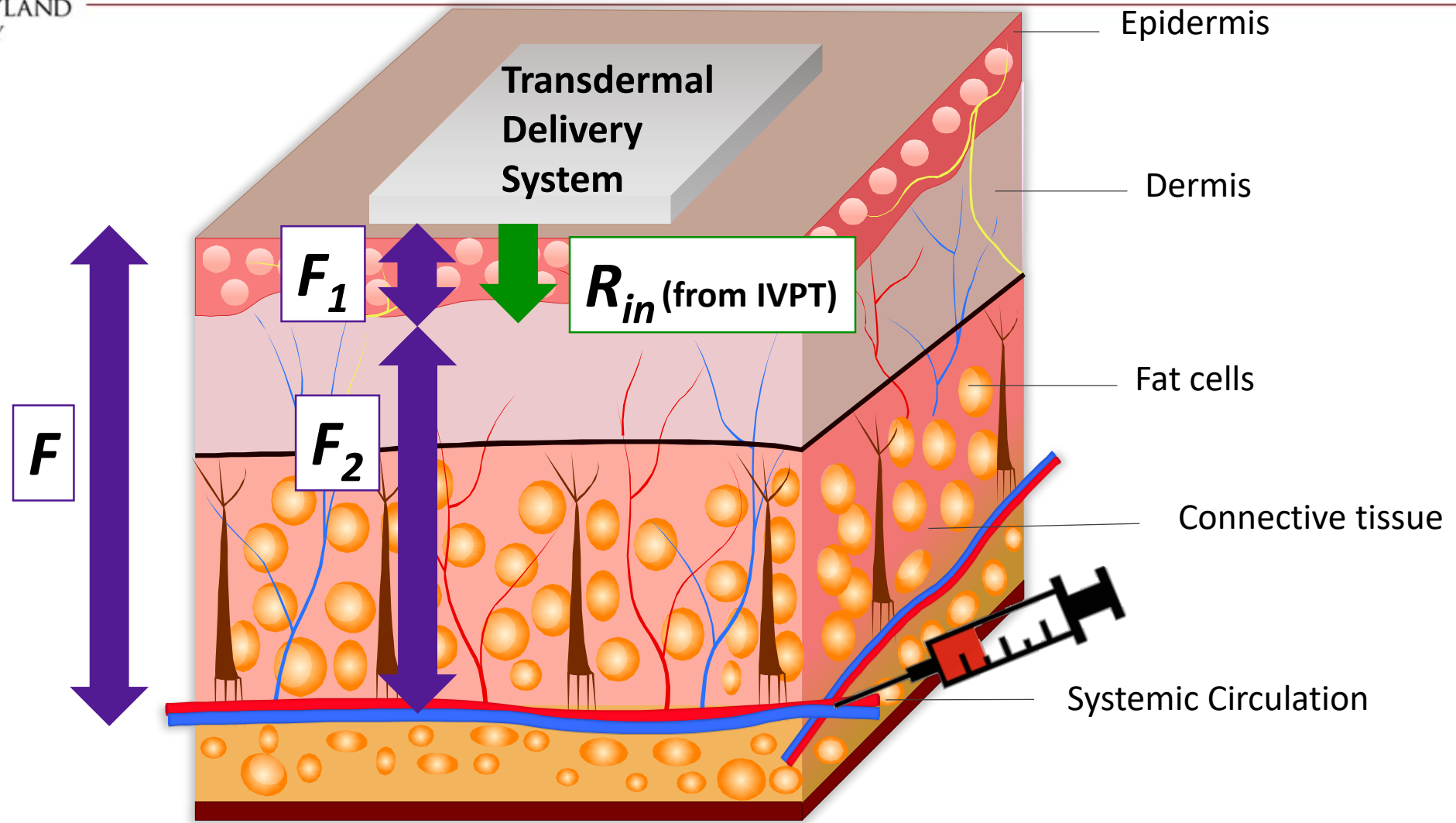
IVIVC: Heat Effects



D: Duragesic®
A: Apotex
M: Mylan

In vivo heat effect is greater than *in vitro*, with higher variability
(Two-way ANOVA followed by Bonferroni's post-hoc multiple pair comparisons)

- *In vitro* data from 4 donors with n=4 replicates per donor
- *In vivo* data from 10 subjects



R_{in} , obtained from IVPT, does not fully capture F , which accounts for skin metabolism and skin depot effect in skin layers. In addition, the small fraction of F accounted for in R_{in} (F_1) has limitations in capturing *in vivo* F since R_{in} is coming from the *in vitro* system. Thus using F in Equation 1 gives the best possible prediction of drug concentration in systemic circulation.



Diclofenac

| | Patch | Solution | 1% Gel | 3% Gel |
|-----------------------------------|--|--|--|--|
| Inactive ingredients | Adhesive in aqueous base containing sodium polyacrylate, sodium carboxymethylcellulose | DMSO, ethanol, purified water, propylene glycol, hydroxypropyl cellulose | Carbomer homopolymer Type C, cocoyl caprylcaprate, fragrance, isopropyl alcohol, mineral oil, polyoxyl 20 cetostearyl ether, propylene glycol, purified water, strong ammonia solution | Hyaluronate sodium, benzyl alcohol, polyethylene glycol monomethyl ether, purified water |
| Dose applied | - | 5 mg/cm ² | 10 mg/cm ² | 20 mg/cm ² |
| (Equivalent amount of diclofenac) | (878 mg/cm ²) | (approx. 100 µg/cm ²) | (approx. 100 µg/cm ²) | (approx. 300 µg/cm ²) |



Diclofenac

| Formulation | Heat Enhancement Ratio (Heat/No Heat) | | ## <i>p</i> value (Heat vs No Heat) | |
|-------------|---------------------------------------|-----------|-------------------------------------|-----------|
| | J_{\max} | Cum. Amt. | J_{\max} | Cum. Amt. |
| Patch | 2.3 | 5.0 | 0.034 | 0.104 |
| Solution | 4.0 | 5.0 | 0.006 | 0.002 |
| 1% Gel | 2.6 | 3.0 | 0.001 | <0.001 |
| 3% Gel | 1.0 | 0.87 | 0.961 | 0.883 |