

Simcyp Version 19 Release Webinar: Dermal Developments

Version 19 Release Webinars

Date	Webinar
Monday 21st October	Updating and Verification of Compounds
Tuesday 22 nd October	Qualification of Compounds
Thursday 24 th October	Dermal Developments
Tuesday 29 th October	Parent to Metabolite Inter-conversion
Thursday 31 st October	ADAM Developments Under FDA Grant
Monday 4 th November	Verification of Populations
Thursday 7 th November	Transporters IVIVE
Tuesday 12 th November	Enhancement/Expansion of PE and ASA

10th Annual Simcyp Virtual Seminar

The 10th Annual Simcyp Virtual Seminar on the Applications of Population-based IVIVE and PBPK provides the chance to join Consortium members, academic and regulatory Simcyp licence holders to discuss usage of the latest applications of the Simulator and recent advances in the field.

Wednesday 13th November (15:00 GMT)	Simcyp Virtual Seminar – Part I	
<i>Integration of PBPK modelling into protocol design to understand nuances in variability in drug exposure</i>		Flinders University, Australia
<i>The use of PBPK modelling across the pediatric age range using propofol as a case</i>		Freie Universitaet Berlin, Germany
<i>Bottom-up PBPK modelling using expression- and activity-based IVIVE to predict plasma and tissue xenobiotic concentrations</i>		Skin Research Institute of Singapore










Wednesday 20th November (15:00 GMT)	Simcyp Virtual Seminar – Part II	
<i>Striving for a neonatal PBPK model of morphine using patients' information: Knowledge gaps and next challenges</i>		Cincinnati Children's Hospital, OH, USA
<i>Application of PBPK modelling to predict the pharmacokinetics of drugs in the special populations</i>		NIPER, India
<i>PBPK modelling and in vitro biorelevant dissolution testing for prediction of oral budesonide pharmacokinetics in Crohn's disease</i>		University of Bath, UK

Registration is available via the Simcyp members' area: <https://members.simcyp.com>

Upcoming Events

Choose between the 5-day “all-in-one” workshop or various 2-day “focused” workshops covering specific topics

2020 Simcyp Workshops

-  **2d** **Washington DC, USA** • Sheraton Silver Spring Hotel
March 23-24: Best Practice, Biologics, FIH, Pediatrics, Absorption I,
March 25-26: Absorption II, Transporters, DDI, PE/PD, or Special Populations
-  **5d** **London, UK** • Holiday Inn London Kensington Forum
May 11-15: Full 1 week workshop
-  **2d** **Sheffield, UK** • Certara Simcyp Offices
June 23-24: Best Practice, Biologics, FIH, Pediatrics, Absorption I,
June 25-26: Absorption II, Transporters, DDI, PE/PD, or Special Populations
-  **5d** **Cambridge, MA, Boston** • Boston Marriott Cambridge
July 6-10: Full 1 week workshop
-  **5d** **Tokyo, Japan** • TKP Conference centre
August 17-21: Full 1 week workshop
-  **2d** **Sheffield, UK** • Mercure St Paul's Hotel & Spa
September 21-22: Best Practice, Biologics, Transporters, Pediatrics, Absorption I,
Absorption II, FIH, DDI, PE/PD, or Special Populations
-  **3d** **Sheffield, UK** • Mercure St Paul's Hotel & Spa
September 22-24: Annual Consortium Meeting
-  **5d** **Princeton, NJ, USA** • Nassau Inn
October 19-23: Full 1 week workshop
-  **5d** **Shanghai, China** • Radisson Blu Hotel Shanghai New World
November 16-20: Full 1 week workshop



CERTARA
Simcyp

REGISTRATION DEADLINE: September 24, 2019

Intensive Workshop
October 14-18, 2019

CERTARA
Simcyp

REGISTRATION DEADLINE: October 11, 2019

Intensive 2-day Workshops on Model-informed Drug Development
November 12-13 & 14-15, 2019 • Shanghai, China

Certara-Simcyp Focused Workshops supported by the Peking Union Medical College Hospital

Incorporating population variability into mechanistic prediction of PK and modeling PK/PD

Focused workshops offer an in-depth insight into specialist areas of modeling such as (arrow indicates availability for this event):

- ▶ Transporter-related ADMET
- Absorption and pharmacokinetic issues
- Pediatric drug development
- Parameter estimation (PE) and pharmacodynamics (PD)
- ▶ Complex drug-drug interactions (DDIs)
- Biologics
- Cardiac safety assessment using pre-clinical data
- Best practice in PBPK model building
- From discovery to first-in-human (FIH) using MVE-PBPK modeling

The model-informed approach to various aspects of drug development is rapidly being adopted by the leading pharmaceutical companies. The Simcyp workshops focus on the optimal use of compound-specific in vitro and in vivo data together with system-specific information related to humans to simulate and understand drug behavior in various target populations. This integrated approach informs decisions related to Investigational New Drugs and assists with the conduct and optimal design of clinical studies. The ultimate aim is to better understand drug PK/PD properties, reduce the cost and time of drug development, improve the quality of regulatory submissions, and eventually implement precision medicine.

Simcyp workshops are an ideal way to enhance the continuous education of scientists working in discovery, DMPK, clinical pharmacology, pharmaceuticals, and drug development. These events provide an excellent opportunity to develop skills, stay up to date with the latest scientific advances, and network with delegates from industry, academia, and regulatory agencies.

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CERTARA

Simcyp

ON-SITE EDUCATION

**Tailored, 1-2 day events
which can incorporate:**

- basic or advanced simulator training
- focused workshops covering key topics in model-based drug development
- guidance through hands-on exercises providing experience with practical applications of the simulator
- face-to-face support for internal Simcyp working groups

Simcyp education can cater for small groups of scientists looking for bespoke training and support through to larger workshop-style sessions. Prior to arranging the on-site education, two Simcyp Scientists, Dr Matthew Harwood & Dr Devendra Pade will work with clients to assess their needs and ensure the most appropriate format and material will be provided for the course.

For further information please contact Matt or Deven directly, or email: Simcyp.onsite@certara.com

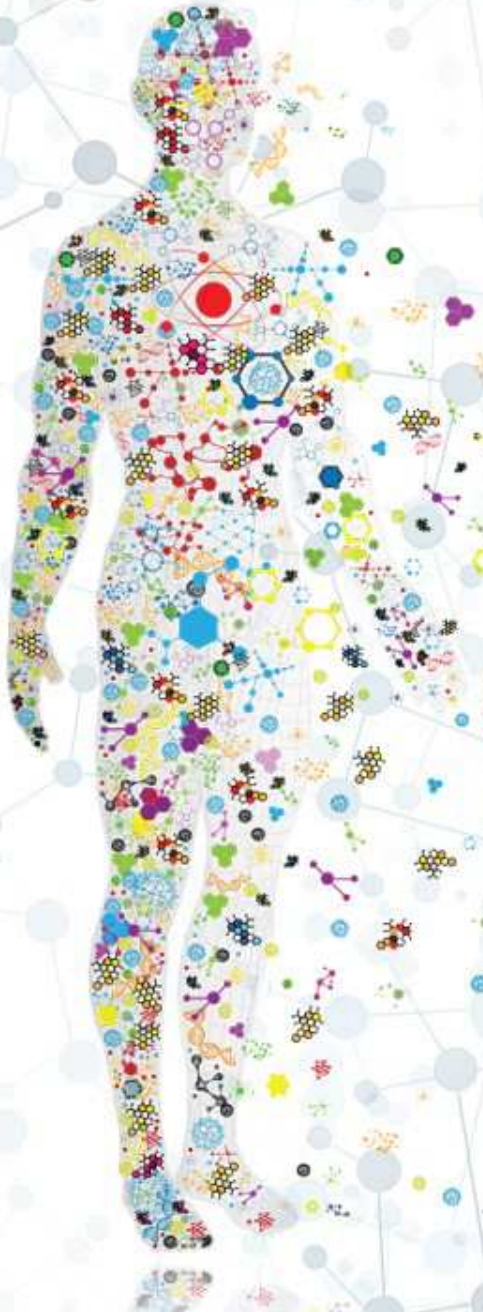


- Tailored to suit education needs at your site
- Co-ordinated by Scientists who will work with you to plan the education programme and team

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Formulation Drug Product Quality Attributes Additions in Dermal PBPK Model

The work is supported as a part of FDA
funded Grant 1U01FD006522-01



Outline of First Part of Presentation



Introduction and Background of the Current Work

Updates in Formulation Input Parameters in Dermal Module V19

- Evaporation Profile Input
- Supersaturation and Precipitation
- Particle Size Distribution
- Introduction of Formulation Toolbox
- Updates to Prediction Toolbox

Summary and Questions

Topical and Transdermal Products – Complex Products

Topical

- Solutions and Sprays
- Creams and Lotions
- Ointments and Oils
- Gels and Jellies
- Shampoos
- Aerosol Foams
- Patches, Tapes and Films

Transdermal

- Transdermal Delivery Systems – Patches
- Ointments
- Gels

Formulation options in Simcyp MPML MechDermA model

Solution

Emulsion

Suspension

Patches

Slide Adapted from Dr. Sam Raney Presentation at Complex Generic Drug Product Development Workshop held in September 25-25, 2019

Evaluation of BE for Topical Products

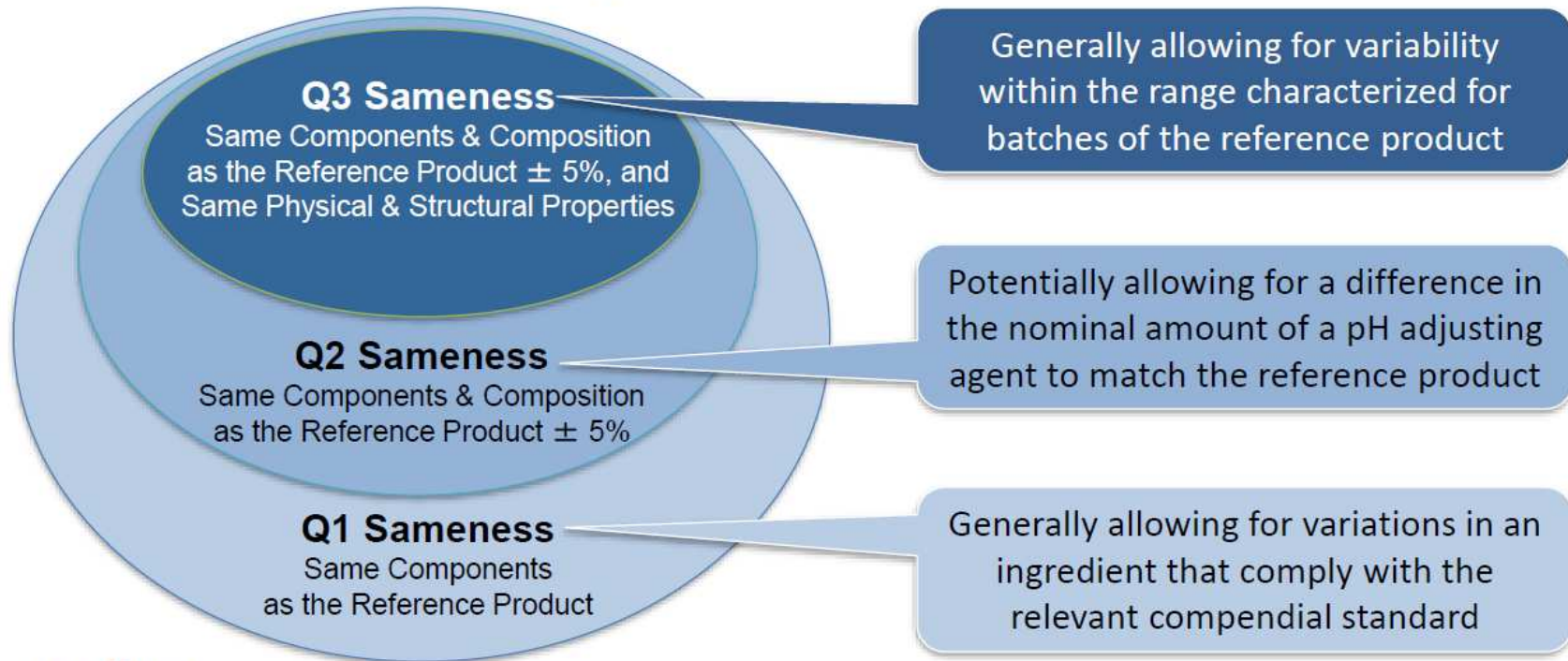


- A Modular Framework for In Vitro BE Evaluation
 - **Qualitative (Q1) and Quantitative (Q2) Sameness**
 - **Physical and Structural (Q3) Sameness**
 - **IVRT** (In Vitro Release Test)
 - **IVPT** (In Vitro Permeation Test)
- Multiple Approaches for BE Evaluation
 - **In Vivo Pharmacokinetic** Studies
 - **In Vivo Pharmacodynamic** (Vasoconstrictor) Studies
 - **In Vivo Comparative Clinical Endpoint BE** Studies
 - **In Silico** Quantitative Methods, Modeling and Simulation



Q3 Sameness for Topical Products

- An evolving concept for topical dermatological products



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Elef1

FDA - U

PSG – Product Specific Guidelines

Slide taken from Dr. Sam Raney Presentation at Complex Generic Drug Product Development Workshop held in September 25-25, 2019

Major Questions -

1. How can we relate these Q3 properties of the test formulation to its clinical performance?
2. Can we identify critical Q3 properties which are likely to exert significant effects on the bioavailability of the API from the formulation?
3. Can we come up with a safe space where we can widen the range of these critical Q3 properties ?

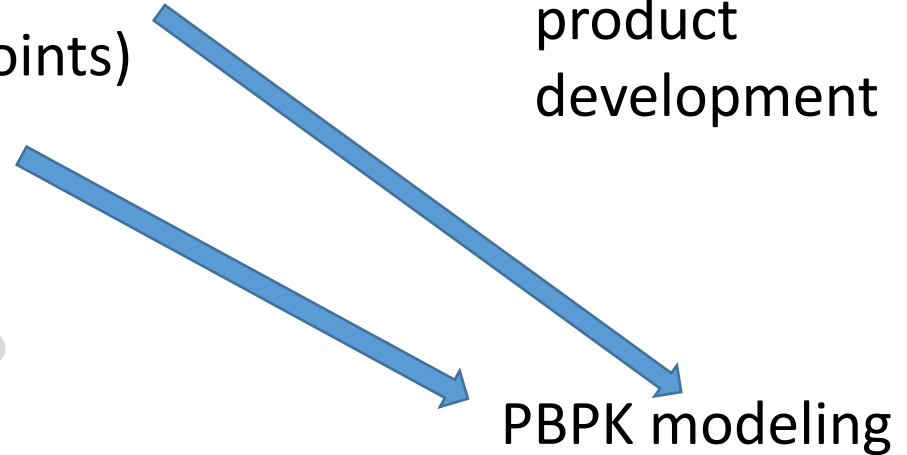
PBPK Modelling can play a very important role in answering all these questions

PSGs for Complex Topical/Transdermal Products

Option 1: R and T are Q1/2

- *In vitro* characterization (Q3 similarity+ IVRT + IVPT)
- (+ *In vivo* BE study with PK endpoints)
- (+ *In vivo* comparative clinical end point BE study)

—————→ PBPK modeling to support drug product development



Option 2: R and T are not Q1/2

- *In vivo* comparative clinical end point BE study

-----→ PBPK modeling to support alternative BE approaches

R: Reference, T: Test
IVRT – In vitro Release Testing
IVPT – In vitro Permeation Testing

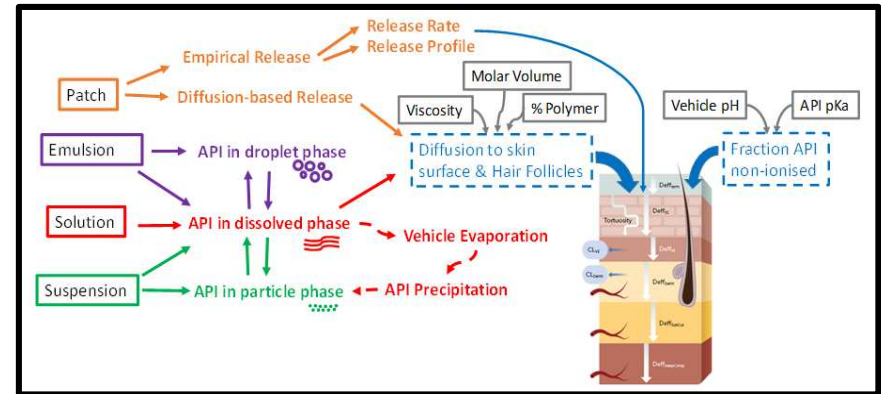
Q3 properties for single/multiple phase systems

- Appearance
- Microscopy
- Particle Size
- Polymorphic form
- Drying rate (weight loss)
- Specific gravity
- Rheology
- pH
- Water activity
- Globule Size
- Solubility of drug in different phases
- Supersaturation and Precipitation potential
- Impact of container closure system

Formulation Q3 inputs in MPML-MechDermA model

Q3 properties for single/multiple phase systems

- Appearance
- Microscopy
- **Particle Size**
- Polymorphic form
- **Drying rate (weight loss)**
- **Specific gravity**
- **Rheology**
- **pH**
- Water activity
- **Globule Size**
- **Solubility of drug in different phases**
- **Supersaturation and Precipitation potential**
- Impact of container closure system



Formulation Inputs in MPML
MechDermA Model

Major Screen Change in V19 (Permeability)

Split of Permeability and Formulation Sections

SV-Alfentanil simuCY

Depth resolved Dermis model

Permeability | Formulation

	Whole SC	Top 25% SC Layers	Upper Middle 25% SC Layers	Lower Middle 25% SC Layers	Bottom 25% SC Layers
Stratum Corneum (SC) Permeability Scalar	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>

Drug Partition Coefficients

Stratum Corneum Lipid : Water K_p	<input type="text" value="36.963571854"/>	Dermis : Viable Epidermis K_p	<input type="text" value="1"/>
Stratum Corneum Lipid : Vehicle K_p	<input type="text" value="36.963571854"/>	Dermis : Blood K_p	<input type="text" value="1.9307982865"/>
Stratum Corneum Lipid : Viable Epidermis K_p	<input type="text" value="11.535329989"/>	Dermis : Sebum K_p	<input type="text" value="0.9088896119"/>
Sebum : Water K_p	<input type="text" value="40.668934234"/>	Muscle : Subcutis K_p	<input type="text" value="1"/>
Sebum : Vehicle K_p	<input type="text" value="40.668934234"/>	Blood : Muscle K_p	<input type="text" value="1"/>
Subcutis : Dermis K_p	<input type="text" value="1E-06"/>	Blood : Subcutis K_p	<input type="text" value="1"/>

Drug Diffusion Coefficients (cm²/h)

SC Lipid (D _{scLip})	<input type="text" value="1.0870009313"/>	Dermis (D _d)	<input type="text" value="0.0009327245"/>
Viable Epidermis (D _{ve})	<input type="text" value="0.0009327245"/>	Sebum (D _{se})	<input type="text" value="0.0004769545"/>
Subcutis (D _{subcutis})	<input type="text" value="1E-05"/>	Muscle (D _{muscle})	<input type="text" value="1E-05"/>

Keratin Binding Kinetics

Steady State

frac Fraction Unbound in SC K_D keratin

Dynamic Adsorption/Desorption Kinetics

Corneocyte membrane permeability (cm/h)

Fraction of drug non-ionised in corneocyte ($f_{ni,com}$) CV (%)

Dermis Parameters

Major Screen Change in V19 (Formulation)

Split of Permeability and Formulation Sections

SV-Alfentanil sim⁴CYT

Depth resolved Dermis model

Permeability | **Formulation**

Duration of Application (h) CV (%)

Formulation Options and Parameters

Formulation pH is skin surface pH Formulation pH CV (%)

Fraction non-ionised at skin surface $f_{n,skin\ surface}$

Formulation drug liberation lag time (h) CV (%) Apply lag time to vehicle evaporation

Consider Vehicle Evaporation

Temperature of skin (°C)	<input type="text" value="32"/>	Vapour pressure of vehicle at skin temperature (mm Hg)	<input type="text" value="43"/>	Mean	CV (%)
MW of vehicle (g/mol)	<input type="text" value="18"/>	Air velocity (m/sec)	<input type="text" value="0.5"/>	<input type="text" value="30"/>	<input type="text" value="30"/>
Density of vehicle (g/ml)	<input type="text" value="1"/>	Maximum % (v/v) vehicle evaporated	<input type="text" value="50"/>	<input type="text" value="30"/>	<input type="text" value="30"/>

(Zero Order) Evaporation rate (ml/h)

First Order Evaporation Rate Constant K_{ER} (1/h)

Vehicle Evaporation Profile

Custom Dermal - Drug/Formulation Parameter(s)

Allow drug to precipitate

Solution

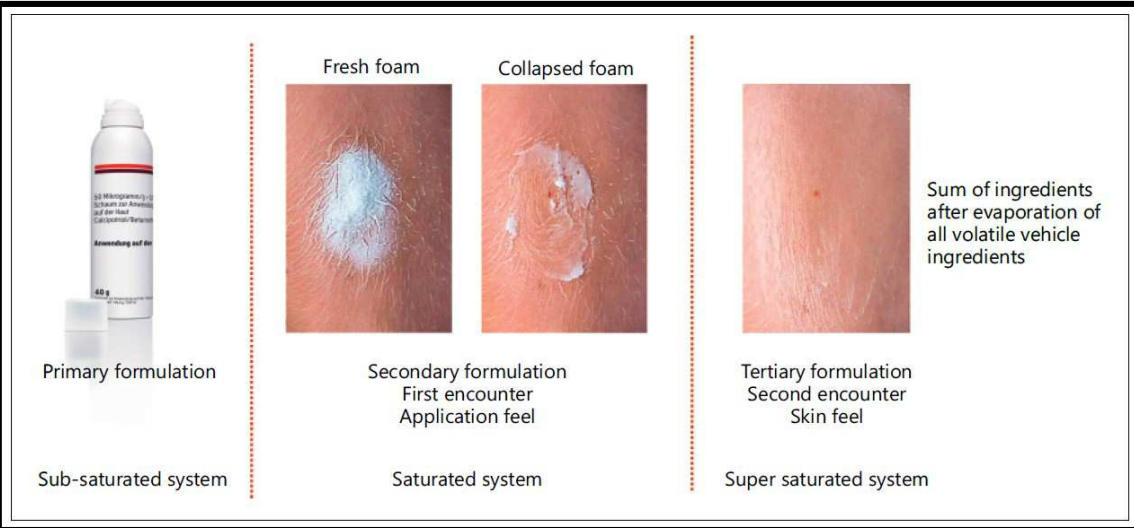
Diffusion Coeff (cm ² /h) <input type="text" value="0.0150523982"/> <input type="checkbox"/>	Vehicle molar volume (mL/mol) <input type="text" value="18"/> <input type="checkbox"/>
Drug Solubility in Continuous Phase (mg/mL) <input type="text" value="1"/>	Viscosity (centipose) <input type="text" value="1"/>
Particle Count for Precipitation <input type="text" value="68436.625529"/> <input type="checkbox"/>	

Emulsion

Suspension / Paste

Dermal Patch

Metamorphosis of Topical Formulations

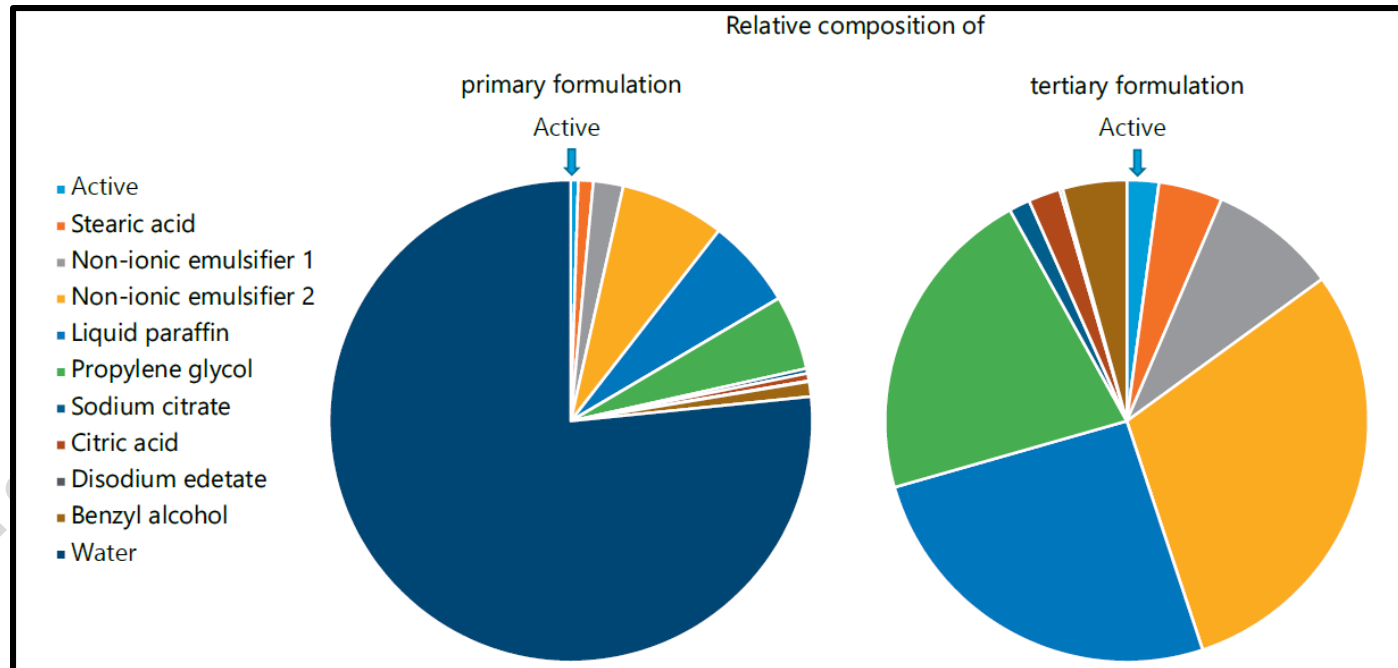


Metamorphosis of the vehicle: primary, secondary and tertiary formulation before, during and after the application procedure of a pressurized product.

Image adapted from Lind et al, Dermatol Ther (Heidelb) 2016; 6: 413-425.

Metamorphosis of the vehicle: relative composition of a commercial topical formulation before (as present in a tube) and after application onto the skin (as present on the skin).

Image adapted from Suber et al, Curr Probl Dermatol. Basel, Karger, 2018, vol 54, pp 152-165



Vehicle Evaporation in MPML MechDerma Model

Options currently available in V18

Formulation Options and Parameters

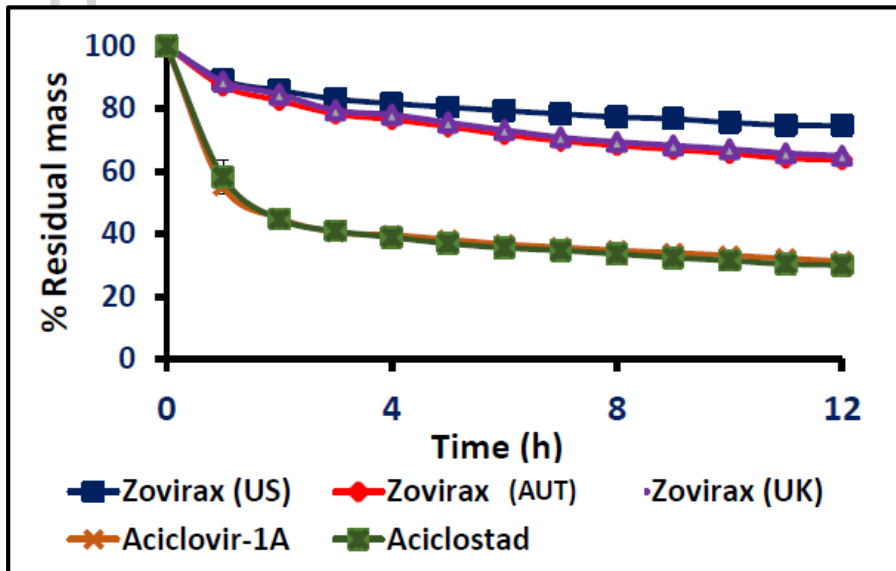
Formulation pH is skin surface pH Formulation pH: 7.74 CV (%): 0
 Fraction non-ionised at skin surface $f_{skin\ surface}$: 0.9700201
 Formulation drug liberation lag time (h): 0 CV (%): 30
 Consider Vehicle Evaporation Apply lag time to vehicle evaporation

Temperature of skin (°C): 32 Vapour pressure of vehicle at skin temperature (mm Hg): 43
 MW of vehicle (g/mol): 18 Air velocity (m/sec): 0.5
 Density of vehicle (g/ml): 1 Maximum % (v/v) vehicle evaporated: 23

(Zero Order) Evaporation rate (ml/h): 1.280496
 First Order Evaporation Rate Constant K_{ev} (1/h): 0.63

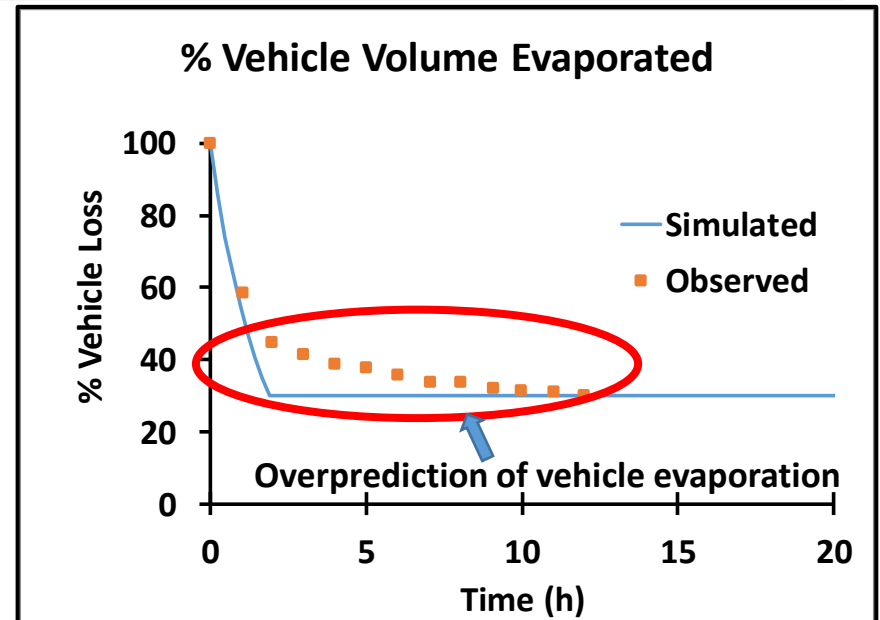
Allow drug to precipitate (only suspension and emulsion with particles)

Estimated First Order Evaporation Rate



Vehicle Evaporation Profile of Acyclovir Commercial Products

Data obtained from Prof. Narasimha Murthy (University of Mississippi) Presentation



Observed and Simulated Vehicle Evaporation Profile of Aciclostad

Vehicle Evaporation – User Input Profile Input Option In V19

Acyclovir

GI Tract Lung Skin Vaginal Tract

MechDerMA Model

Multi-phase multi-layer (MPML) MechDerMA Model

Single Layer Dermis Model

Depth Resolved Dermis Model

Permeability Formulation

Duration of Application (h) CV (%)

Formulation Options and Parameters

Formulation pH is skin surface pH Formulation pH

Consider Vehicle Evaporation

Temperature of skin (°C)

MW of vehicle (g/mol)

Density of vehicle (g/ml)

(Zero Order) Evaporation rate (ml/h)

First Order Evaporation Rate Constant K_{ER}

Vehicle Evaporation Profile

Custom Dermal - Drug/Formulation Parameter(s)

Allow drug to precipitate

Mechanistic Growth Model (only suspensions and emulsions with particles)

Empirical Model (only solutions and emulsions without particles)

Vehicle Evaporation Profile

Note - Vehicle Evaporation Profile should be defined as volatile vehicle loss (%v/v) vs Time.

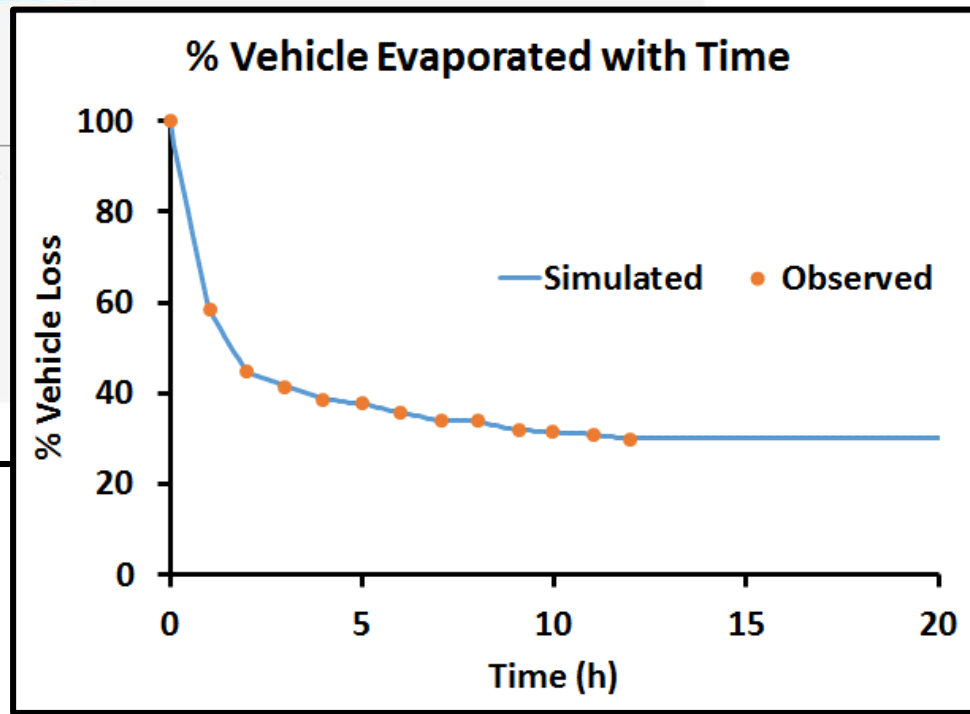
Discrete

Time (h)	0	1	2	3	4	5	6	7	8
Vehicle loss (%v/v)	0	41.62	55.22	58.53	61.36	62.34	64.25	66.16	66.2
CV (%)	0	0	0	0	0	0	0	0	0

Weibull Function

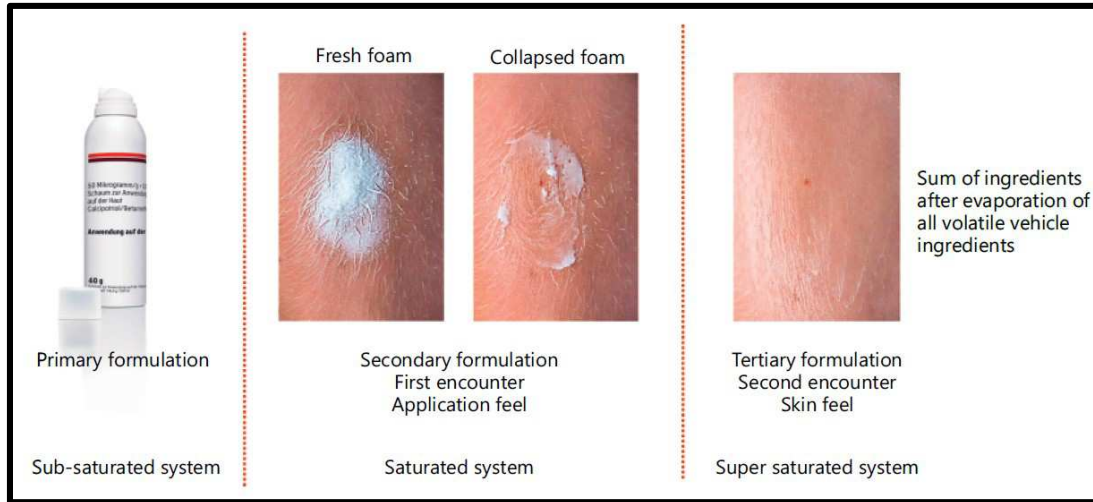
	Mean	CV(%)
Fmax (%)	<input type="text" value="100"/>	<input type="text" value="10"/>
alpha	<input type="text" value="2"/>	<input type="text" value="10"/>
beta	<input type="text" value="1"/>	<input type="text" value="10"/>

Interpolation Method Linear Piecewise Cubic Polynomial



Observed and Simulated Vehicle Evaporation Profile of Aciclostad

Supersaturation and Precipitation Topical Formulations

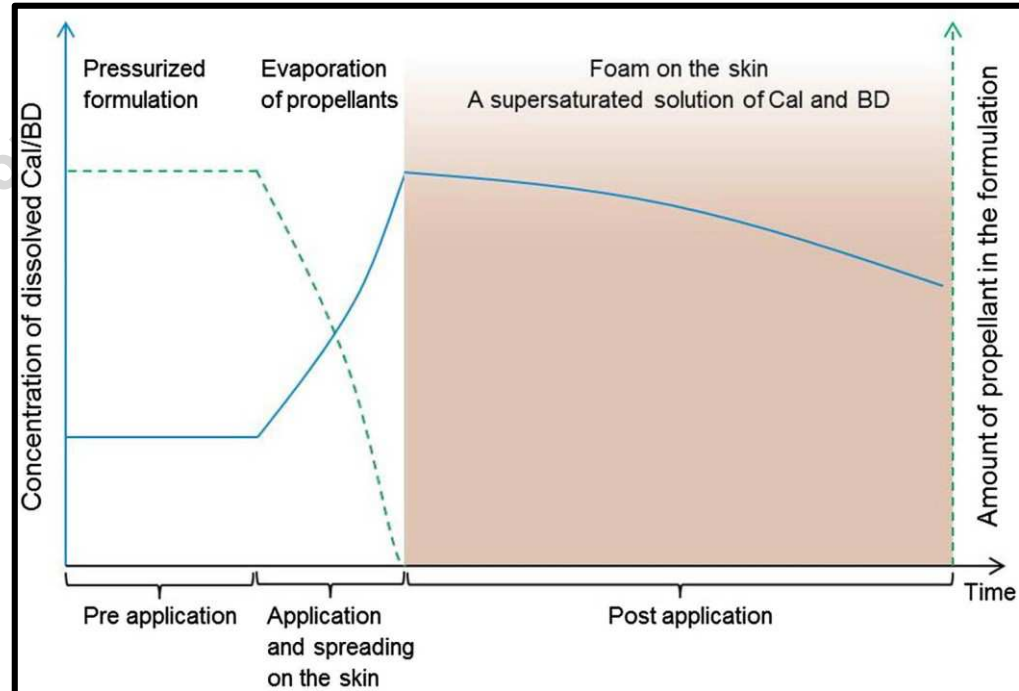


Metamorphosis of the vehicle: primary, secondary and tertiary formulation before, during and after the application procedure of a pressurized product.

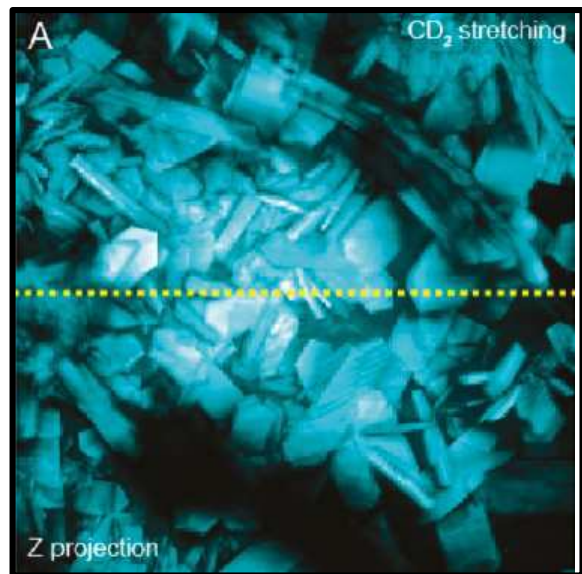
Image adapted from Lind et al, Dermatol Ther (Heidelnb) 2016; 6: 413–425.

Change in concentration of active ingredients dissolved in the aerosol form formulation over application time. BD Betamethasone dipropionate, Cal calcipotriene.

Image adapted from Lind et al, Dermatol Ther (Heidelnb) 2016; 6: 413–425.



Supersaturation and Precipitation Topical Formulations



Crystal formation on the skin surface 25 min post-topical application of a solution of ibuprofen in PG

Image taken from Saar et al, Mol. Pharmaceutics 2011, 8, 969–975

Supersaturation and Precipitation Model – Screen Design (Already in V18)

Formulation Options and Parameters

Formulation pH is skin surface pH Formulation pH: CV (%):
 Fraction non-ionised at skin surface $f_{\text{skin surface}}$:
 Formulation drug liberation lag time (h): CV (%): Apply lag time to vehicle evaporation

Consider Vehicle Evaporation

Temperature of skin (°C): <input type="text" value="32"/>	Vapour pressure of vehicle at skin temperature (mm Hg): <input type="text" value="43"/>	Mean: <input type="text" value="43"/>	CV (%): <input type="text" value="30"/>
MW of vehicle (g/mol): <input type="text" value="18"/>	Air velocity (m/sec): <input type="text" value="0.5"/>	Mean: <input type="text" value="0.5"/>	CV (%): <input type="text" value="30"/>
Density of vehicle (g/ml): <input type="text" value="1"/>	Maximum % (v/v) vehicle evaporated: <input type="text" value="50"/>	Mean: <input type="text" value="50"/>	CV (%): <input type="text" value="30"/>

(Zero Order) Evaporation rate (ml/h): CV (%):
 First Order Evaporation Rate Constant K_{ER} (1/h): CV (%):

Custom Dermal - Drug/Formulation Parameter(s)

Allow drug to precipitate (only suspension and emulsion with particles)

Particle Growth 'Reverse DLM' – replaces FO rate constant approach

where $C_{\text{bulk}} > S_{\text{formulation}}$

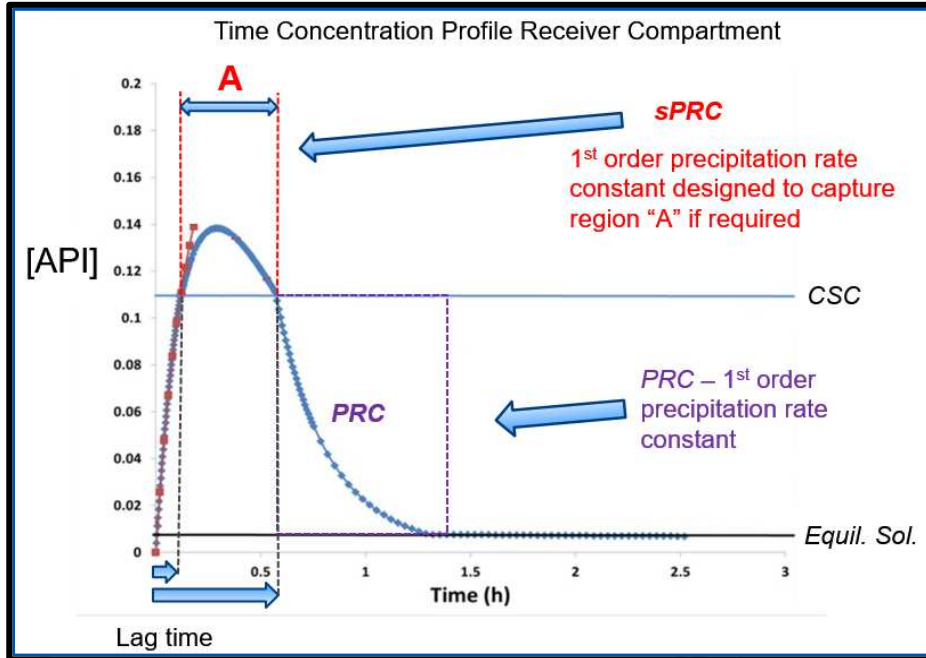
$$PR(t) = -N \frac{D_{\text{eff}}(t)}{h_{\text{eff}}(t)} 4\pi a(t) (a(t) + h_{\text{eff}}(t)) (S_{\text{surface}}(t) - C_{\text{bulk}}(t))$$

- Solution
- Emulsion
- Suspension / Paste

Diffusion Coeff (cm²/h): Vehicle viscosity (centipose):
 Drug solubility in vehicle (mg/mL):
 Volume fraction of solid particle (%): Vehicle molar volume (mL/mol):
 Particle diameter (µm):

Supersaturation and Precipitation Empirical Model

- Formulation - Solution and Emulsion without particle



Empirical Model

IF supersaturated conditions encountered THEN:

Dissolution stops

Precipitation can only begin when

CSC is reached

CSC is a critical conc. at which precipitation starts

[Drug] may continue to rise due to slow permeation of drug from skin

Supersaturated conc. may exceed CSC (CSR x Eq.Sol)

CSC – Critical Supersaturation Concentration

CSR – Critical Supersaturation Ratio

PRC – Precipitation Rate Constant (1/h)

sPRC – Secondary Precipitation Rate Constant (1/h)

Allow drug to precipitate

Mechanistic Growth Model (only suspensions and emulsions with particles)

Empirical Model (only solutions and emulsions without particles)

Critical Supersaturation Ratio

Precipitation Rate Const. (1/h)

Apply Secondary PRC Secondary PRC (1/h)

Reference Concentration Total Concentration in continuous phase (unionized + ionized)

Unionized Concentration in continuous phase

Supersaturation and Precipitation Simcyp Results

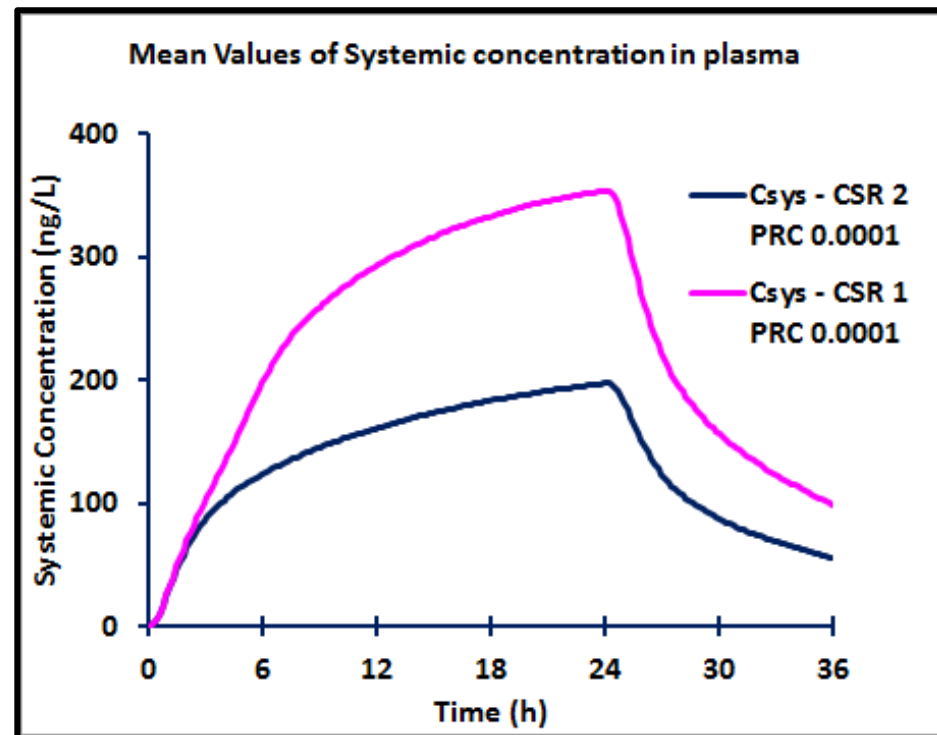
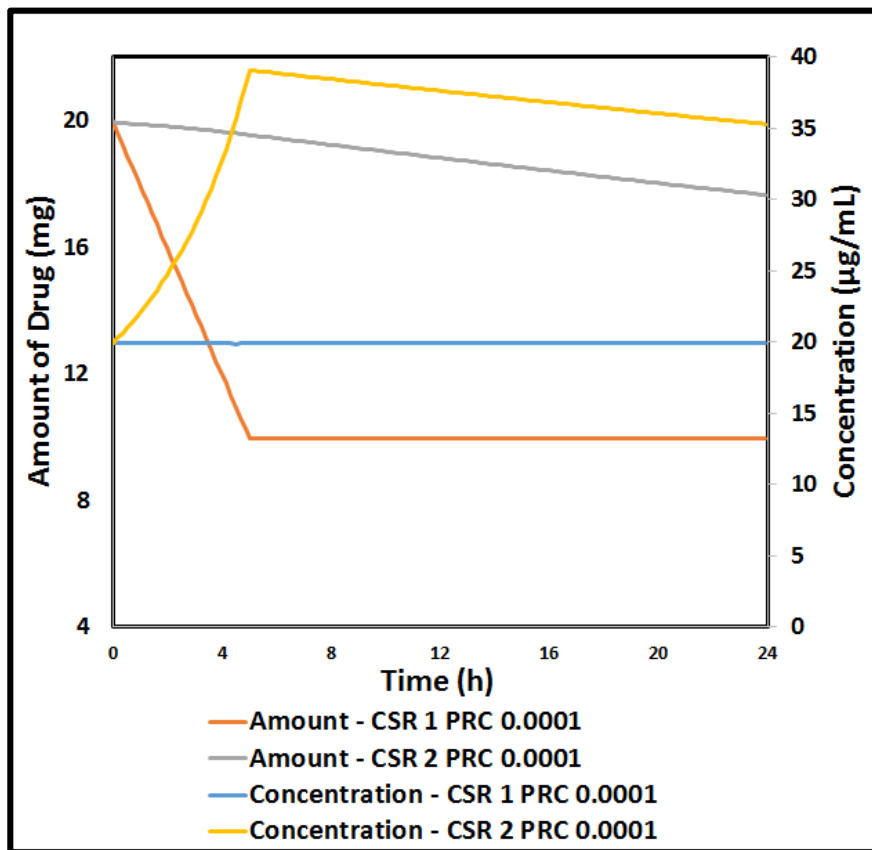
Formulation Solution

Dose 20 mg

Solubility 20 mg/mL

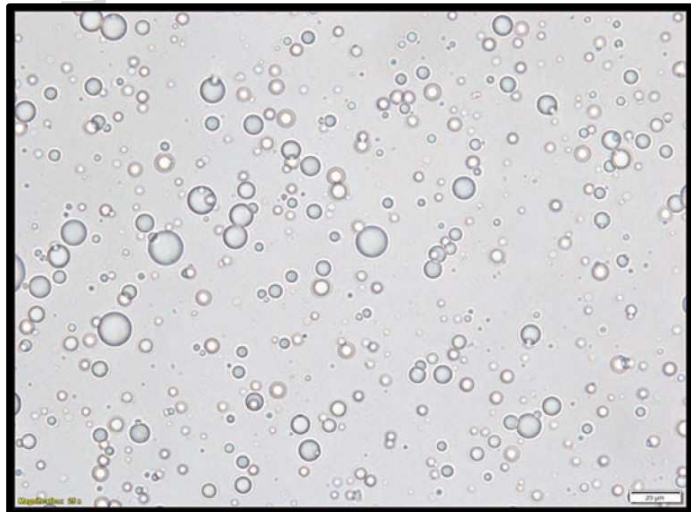
Volume of Formulation 1 mL

Vehicle Evaporation = 0.1 mL/h



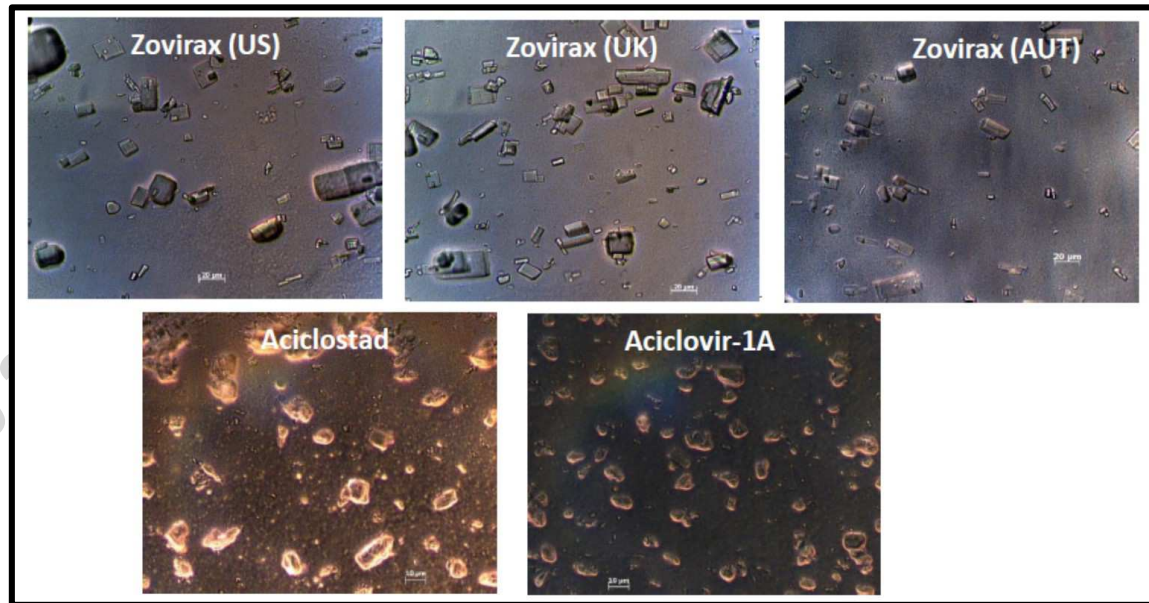
Particle Size Distribution (PSD) – Droplet and Solid Particles

- Topical Emulsion semisolid formulation containing suspended API are most complex pharmaceutical products to develop for application to the skin
- Ratio of dissolved active to suspended active can influence skin permeation
- API PSD or globule PSD can have significant effect on the bioavailability of API



Microscopic picture (Magnification, 500×) of the carbomer “Emulgel” containing dissolved API and dispersed oil globules in the gel matrix.

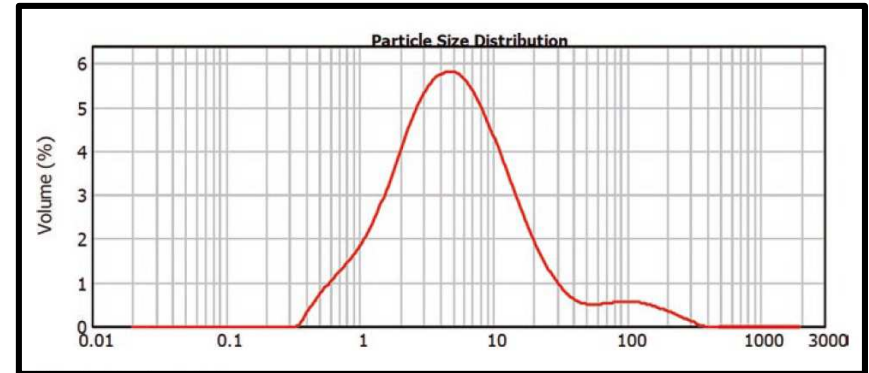
Taken from book The Role of Excipients in the Microstructure of Topical Semisolid Drug Products



Acyclovir API characterization in five commercial products.
Slide obtained from Prof. Narasimha Murthy (University of Mississippi) Presentation

Particle Size Distribution (PSD) – Droplet and Solid Particles

In V19, users can define the particle size distribution (PSD) of the droplets and solid particles as either monodisperse or polydisperse particle size distribution in emulsion and suspension formulation.



Custom Dermal - Drug/Formulation Parameter(s)

Allow drug to precipitate

Solution

Emulsion

Diffusion Coeff (cm ² /h)	<input type="text" value="1.37474885E-4"/>	Vehicle molar volume (mL/mol)	<input type="text" value="18"/>
	<input type="checkbox"/>	Viscosity (centipose)	<input type="text" value="129062"/>
Volume fraction of dispersed phase (%)	<input type="text" value="2.4968"/>	Drug solubility ratio dispersed/continuous phase	<input type="text" value="13.5"/>
Radius of dispersed phase droplets (μm)	<input type="radio"/> Monodispersed <input checked="" type="radio"/> Polydispersed	Droplet permeability (cm/h)	<input type="text" value="1E-05"/>
Number of droplets per cm ³ (N/mL)	<input type="text" value="2059920944"/>		

Particles in continuous phase?

Volume fraction of solid particle (%)	<input type="text" value="0.5235987755"/>	Drug solubility in continuous phase (mg/mL)	<input type="text" value="1"/>
Radius of particles (μm)	<input checked="" type="radio"/> Monodispersed <input type="radio"/> Polydispersed	Particle Count for Precipitation	<input type="text" value="177776071.43"/>
Number of particles per cm ³ (N/mL)	<input type="text" value="10000000"/>		
Density of solid particle (g/mL)	<input type="text" value="1.2"/>		

Suspension / Paste

Dermal Patch

Particle Size Distribution (PSD) – Droplet and Solid Particles

Particle Size Distribution of Dispersed Phase Droplets (Volume Fraction %)

Distribution Function

Normal
 Log Normal
 Weibull

Radius Mean (μm)
 Radius Mean (μm)
 Alpha

CV (%)
 CV (%)
 Beta

Radius Mean (μm)

Radius(μm) Minimum
 Maximum

Discrete

Simulation Parameters

Number of Particle Size Bins (Simulation)

Dosage Form PSD

Radius(μm)
 Minimum
 Maximum

Volume %

Particle Radius (μm)

Cum Volume %

OK Cancel

Particle Size Distribution of Dispersed Phase Droplets (Volume Fraction %)

Distribution Function

Discrete + -

Volume Fraction (%)

Particle Radius (μm)	1	25	50	100
Volume Fraction (%)	25	25	25	25

IMPORT Number of Particle Size Bins Entered (Imported)

Simulation Parameters

Number of Particle Size Bins (Simulation)

Dosage Form PSD

Radius(μm)
 Minimum
 Maximum

Volume %

Particle Radius (μm)

Cum Volume %

OK Cancel

Introduction of Formulation Toolbox

- Allows selection of components of dermal formulations
- Prediction of molar volume of the formulation vehicle
- Correction stratum corneum lipid: water partition coefficient with vehicle: water solubility ratio

Testosterone sim

Dephased Dermal Model

Permeability Formulation

Duration of Application (h) CV (%)

Formulation Options and Parameters

Formulation pH is skin surface pH Formulation pH

Fraction non-ionised at skin surface $f_{n,skin\ surf}$

Consider Vehicle Evaporation Formulation drug liberation lag time

Temperature of skin (°C)

MW of vehicle (g/mol)

Density of vehicle (g/ml)

(Zero Order) Evaporation rate (ml/h)

First Order Evaporation Rate Constant K_{ER} (1/h)


Vehicle Evaporation Profile

Custom Dermal - Drug/Formulation Parameter(s)

Allow drug to precipitate

Solution

Diffusion Coeff (cm²/h)

Vehicle molar volume (mL/mol) 

Drug Solubility in Continuous Phase (mg/mL)

Viscosity (centipose)

Particle Count for Precipitation

Dermal Formulation Popup

Formulation Constituents*

Vehicle Component	Component	% (v/v)	‡ Density ρ	Molar Mass	Molar Volume	Viscosity	Intrinsic Solubility (mg/mL)
Vehicle Component 1	Water	50	1	18.02	18.11		10
<input checked="" type="checkbox"/> Vehicle Component 2	Propylene Glycol	30	1.0276	76.09	74.05		
<input checked="" type="checkbox"/> Vehicle Component 3	Propylene Glycol	20	0.7736	60.1	76.5		
Total Vehicle	Polyethylene Glycol 2-Propanol Oleic Acid	100	1		18.11	3000	5.994320

Formulation Diffusion Coeff (cm²/h)

Diffusion Coeff in Vehicle

Method

Solubility

Formulation f_{ni}^{**}

Solubility at formulation pH*

Vehicle: Water Solubility Ratio

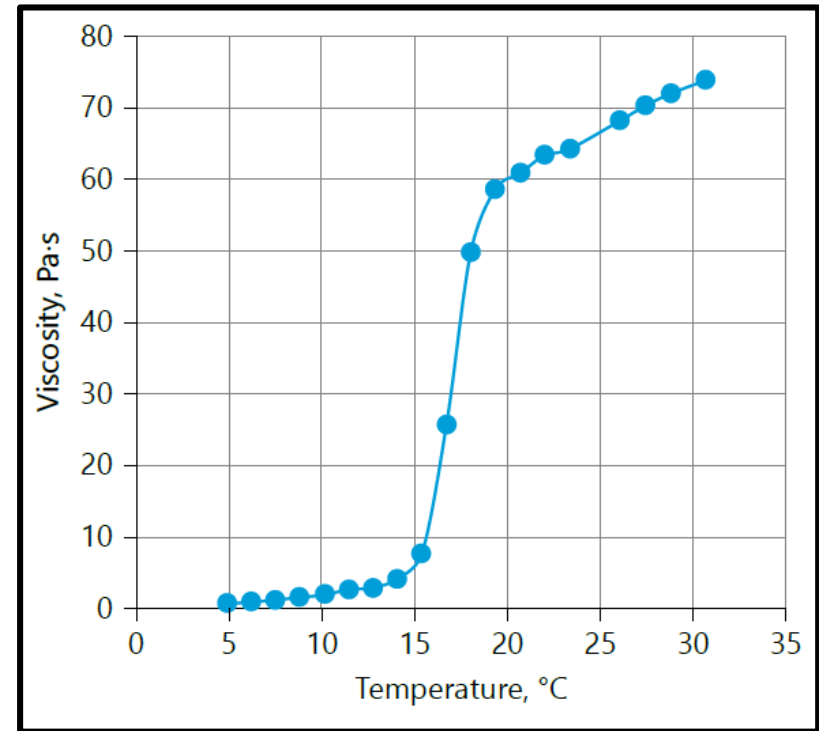
* Constituents of the continuous phase of the selected formulations.
 ** f_{ni} is calculated from the formulation pH assuming Henderson Hasselback, Caution should be used when simulation non-aqueous vehicles as HH may not apply.
 ‡ Density of the entire vehicle must be defined in order to calculate Molar Volume.
 † Applied as a scalar to K_{scv} and $K_{sebum,v}$

Eig FDA

Rheological Considerations

Product	Viscosity, Pa·s			Yield Stress, Pa
	@shear rate: 20 s ⁻¹	@ shear rate 3300s ⁻¹	@ shear rate: 0.0025 s ⁻¹	
Zovirax-USA	17	0.28	8360	50
Zovirax-UK	N/A	N/A	31000	300
Zovirax-AUT	N/A	N/A	30100	300
Aciclostad	3.2	0.06	29300	100
Aciclovir- 1A	2.6	0.06	28100	100
	Dictates the behavior during the initial application	Dictates the behavior during spreading the sample on skin	Dictates at rest condition, i.e., diffusion of drug through thin film	

Viscosity Differences of Commercial Acyclovir Products at Different Shear Rates



Some thermo-sensitive gels show increased product viscosity (Pa·s) at body temperature (°C). As a consequence, a prolonged residence of actives on skin is observed due to increased viscosity of vehicle (e.g., Linoseptic[®] [octenidin dihydrochlorid, phenoxyethanol as disinfectants]).

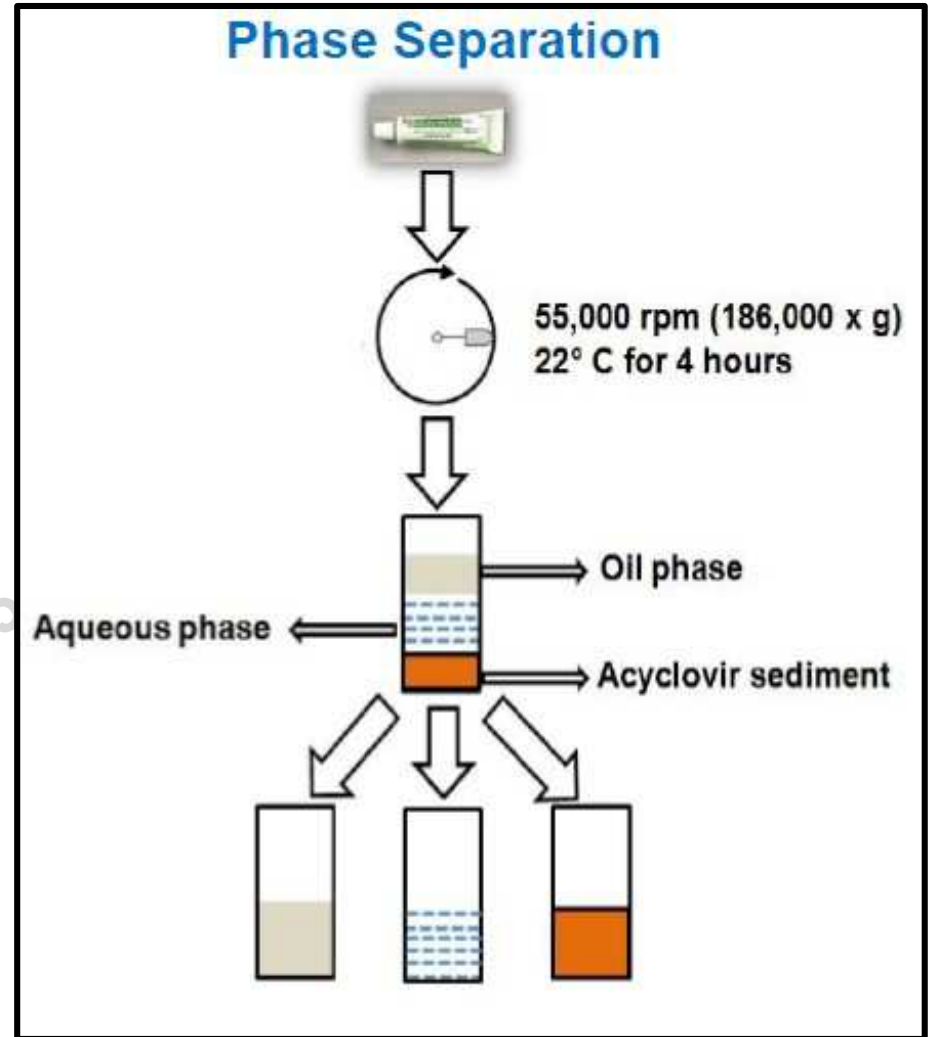
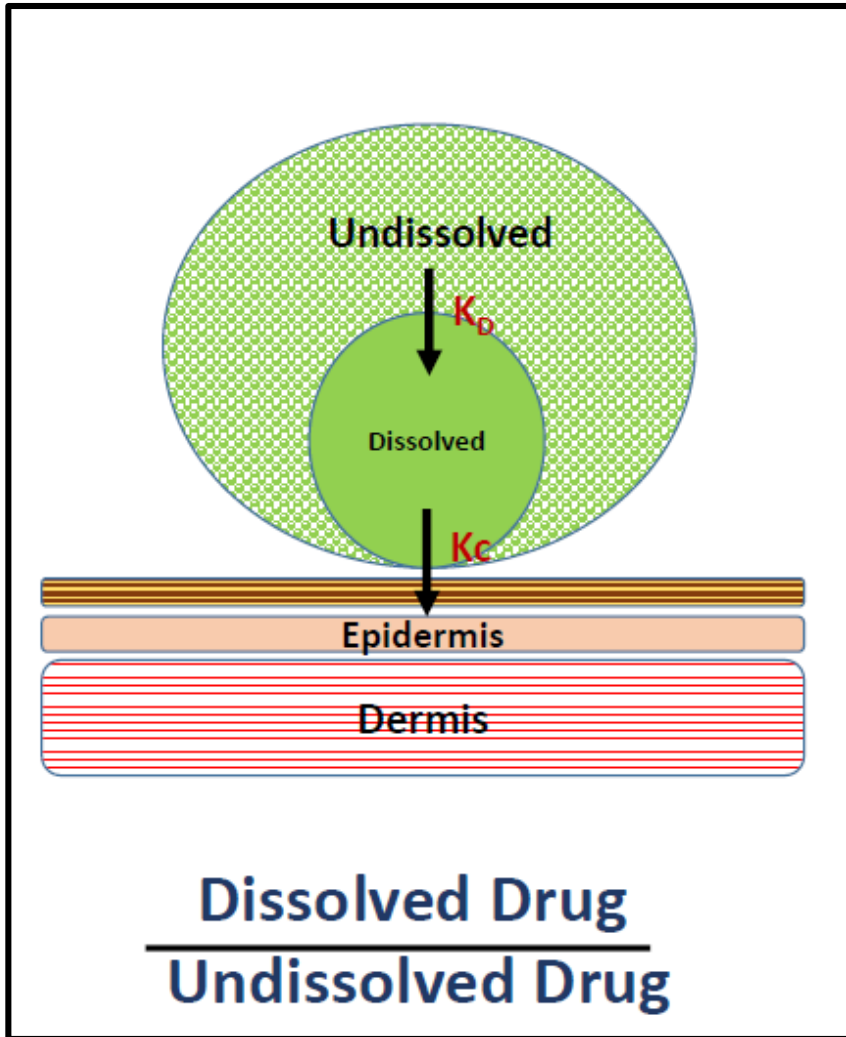
Rheological Considerations – Addition of LUA Functions

The image displays the MPML Skin software interface. On the left, the 'Formulation Options and Parameters' panel is visible, with 'Custom Dermal - Drug/Formulation Parameter(s)' selected. The main window shows a menu path: 'Functions' > 'Formulation Parameters' > 'setVehicleViscosityV'. A second window shows the Lua code for this function:

```
function setVehicleViscosity_V(t, V)
  if (t < 0.25) then
    V = 17
  else
    V = 8360
  end
  return V
end
```

Eleftheria Ts
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Distribution of Drug in Topical Multi-Phase Systems



Slide obtained from Prof. Narasimha Murthy (University of Mississippi) Presentation

Variation of Drug Solubility Ratio – LUA Script

Emulsion

Diffusion Coeff (cm ² /h)	<input type="text" value="1.3747489258"/>	Vehicle molar volume (mL/mol)	<input type="text" value="18"/>	
		Viscosity (centinpoise)	<input type="text" value="129062"/>	
Volume fraction of dispersed phase (%)	<input type="text" value="2.4968958222"/>	Drug solubility ratio dispersed/continuous phase	<input type="text" value="13.5"/>	
Radius of dispersed phase droplets (µm)	<input checked="" type="radio"/> Monodispersed <input type="text" value="1.425"/>	<input type="radio"/> Polydispersed	Droplet permeability (cm/h)	<input type="text" value="1E-05"/>
Number of droplets per cm ³ (N/mL)	<input type="text" value="2060000000"/>			
<input checked="" type="checkbox"/> Particles in continuous phase?				
Volume fraction of solid particle (%)	<input type="text" value="0.5235987755"/>	Drug solubility in continuous phase (mg/mL)	<input type="text" value="1"/>	
Radius of particles (µm)	<input checked="" type="radio"/> Monodispersed <input type="text" value="5"/>	<input type="radio"/> Polydispersed	Particle Count for Precipitation	<input type="text" value="15915494.309"/>
Number of particles per cm ³ (N/mL)	<input type="text" value="10000000"/>			
Density of solid particle (g/mL)	<input type="text" value="1.2"/>			

Permeability | Formulation

Duration of Application (h) 24 CV (%)

Formulation Options and Parameters

- Formulation pH is skin surface pH Formulation pH
- Fraction non-ionised at skin surface
- Formulation drug library
- Consider Vehicle Evaporation
 - Temperature of skin (°C)
 - MW of vehicle (g/mol)
 - Density of vehicle (g/ml)
 - (Zero Order) Evaporation rate (ml/h)
 - First Order Evaporation Rate Constant K_{ER} (1/h)
 - Vehicle Evaporation Profile
- Custom Dermal - Drug/Formulation Parameter(s)
- Allow drug to precipitate

Solution

MPML Skin

File Edit Options Tools Functions **MPML-MechDerMA**

- Stratum Corneum Permeability Scalars
- Drug Partition Coefficients
- Drug Diffusion Coefficients
- Skin Physiology
- Formulation Parameters

setVehicleViscosityV

setDrugSolRatioDC

lag time to vehicle evaporation


CV (%)

<input type="text"/>	<input type="text" value="30"/>
<input type="text"/>	<input type="text" value="30"/>
<input type="text"/>	<input type="text" value="30"/>

Eleftheria Tsakalozou

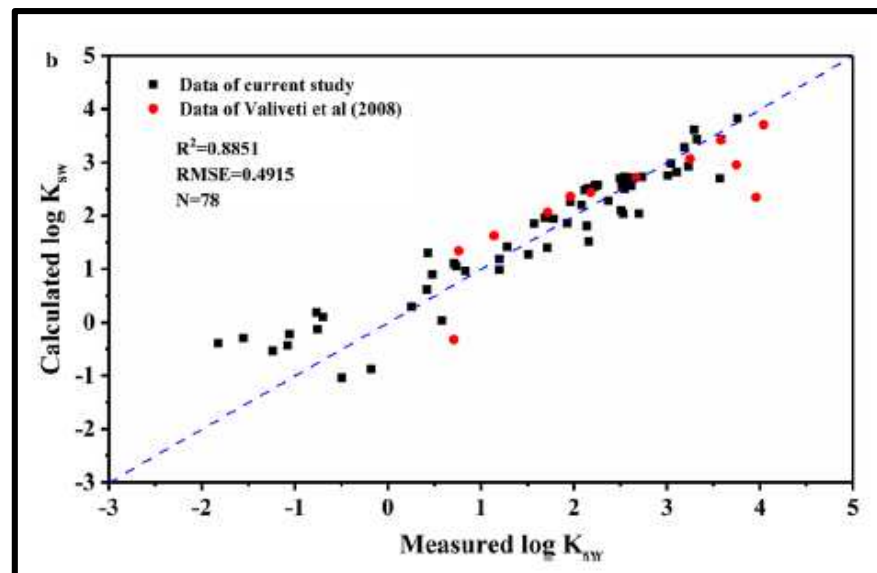
RESEARCH PAPER

Determining the Effect of pH on the Partitioning of Neutral, Cationic and Anionic Chemicals to Artificial Sebum: New Physicochemical Insight and QSPR Model

Senpei Yang¹ • Lingyi Li¹ • Tao Chen² • Lujia Han¹  • Guoping Lian^{2,3}

$K_{sb:w,Yang}$

$$= \left(f_{ni \text{ skin surface}} * \left(\frac{1 + 0.71 * 10^{pH-6.95}}{1 + 10^{pH-6.95}} \right) \right)$$



Summary of Dermal Quality Attributes V19 Updates

- Input of User-Defined Evaporation File
- Empirical model to define supersaturation and precipitation from solution and emulsion without particle formulations
- Input of polydisperse particle size distribution for globules and solid particles
- Addition of formulation toolbox
- LUA functionality to alter vehicle viscosity and drug distribution ratio between phases
- Addition of new QSAR for Sebum:Water Partition Coefficient

Skin Team Members



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Sumit.Arora@certara.com



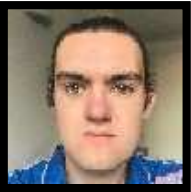
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Questions?



CERTARA[®]

Simcyp

Release Webinar:

**Physiology Related
Enhancements**



Dermal Enhancements in V19

- Depth Resolved Dermis Model

Developed under FDA grant:

Characterization of key system parameters of mechanistic dermal PBPK models in various skin diseases and performance verification of the model using observed local and systemic concentrations

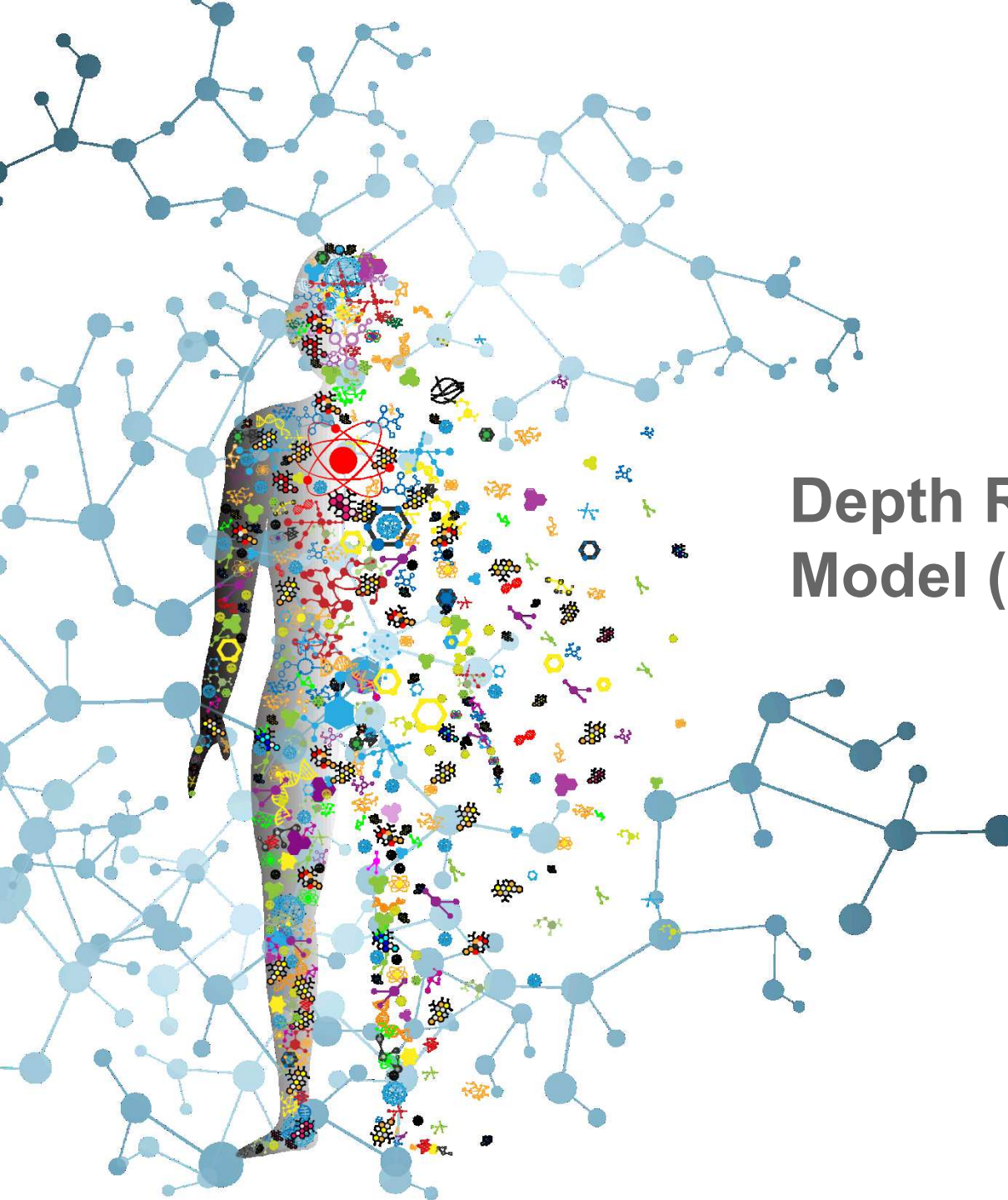
(Grant Award #1 U01FD006521-01)

- Comedone Model (Acne)
- Updates to Psoriasis population file

CERTARA[®]

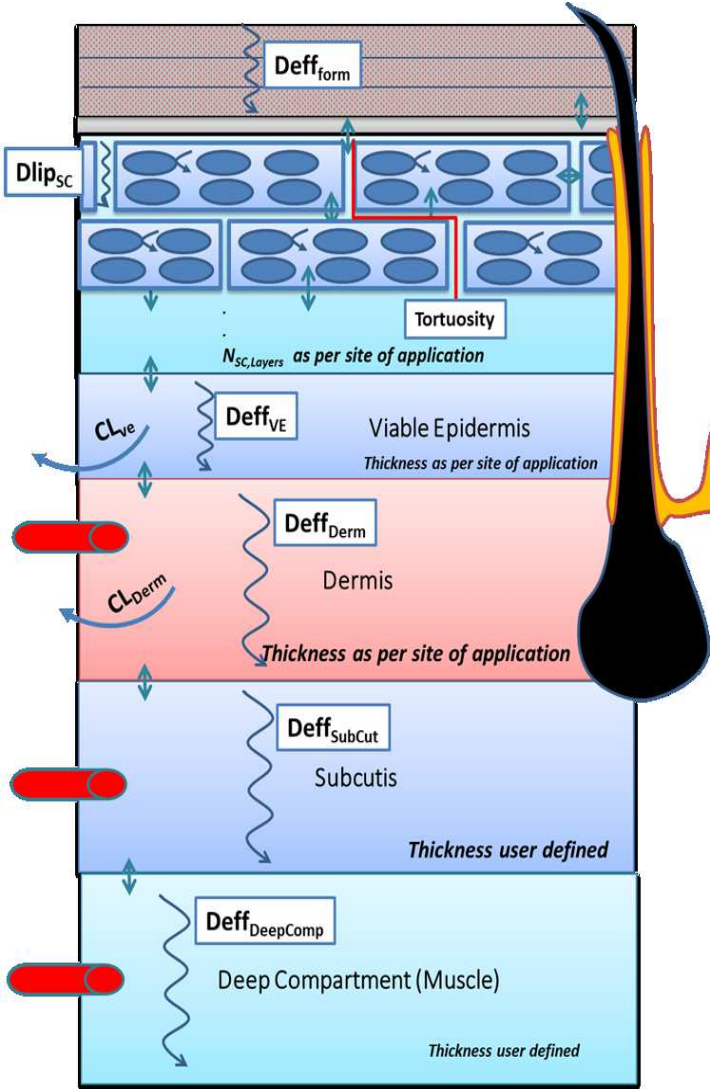
Simcyp

Depth Resolved Dermis Model (DRDM)



Background

Eleftheria Tsakalozou



Formulation (Gel, cream, lotions, paste, patch, ointments, etc.)

- Stratum Corneum (SC)**
- Define cell shape and size
 - Cell membrane permeability
 - Keratin bonding kinetics
 - Tortuosity and diffusivity
 - Hair follicle density and size

- Viable Epidermis (VE)**
- Thickness, diffusivity
 - Metabolism

- Dermis**
- Thickness, diffusivity
 - Metabolism, blood flow

- Subcutis**
- Thickness, diffusivity
 - Blood flow

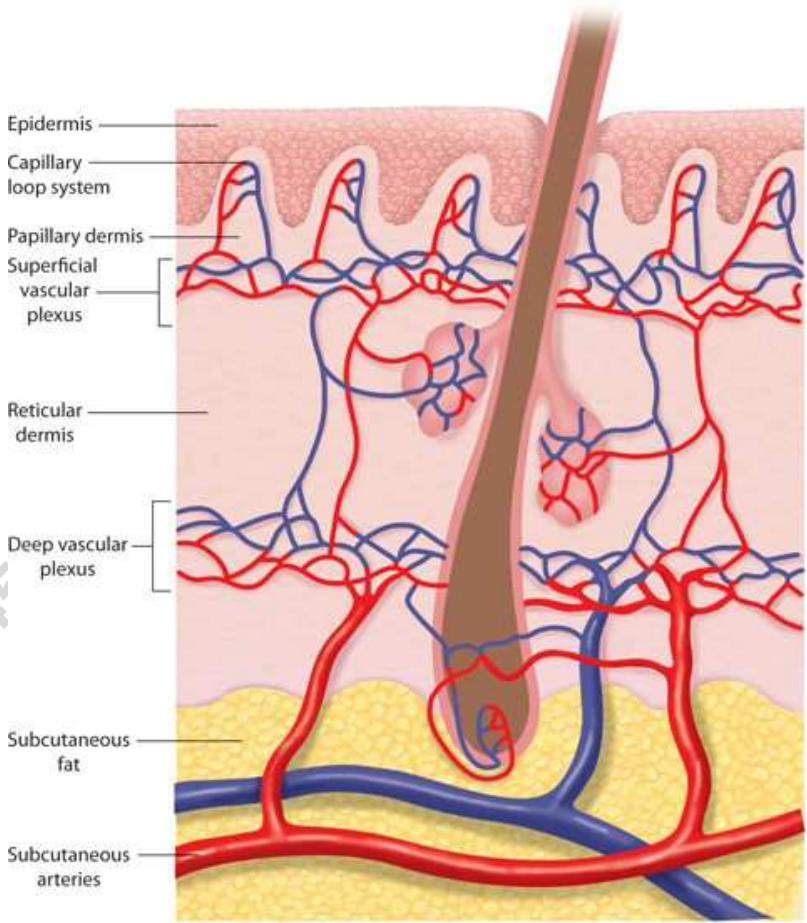
- Deep Tissue**
- Thickness, diffusivity
 - Blood flow

FDA

Dermis Physiology

Two distinct layers

- Papillary ~ 200 μm
- Reticular ~ 1000 μm



Source: Rose L. Hamm: *Text and Atlas of Wound Diagnosis and Treatment*:
www.accessphysiotherapy.com
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Eleftheria Tsakalozou

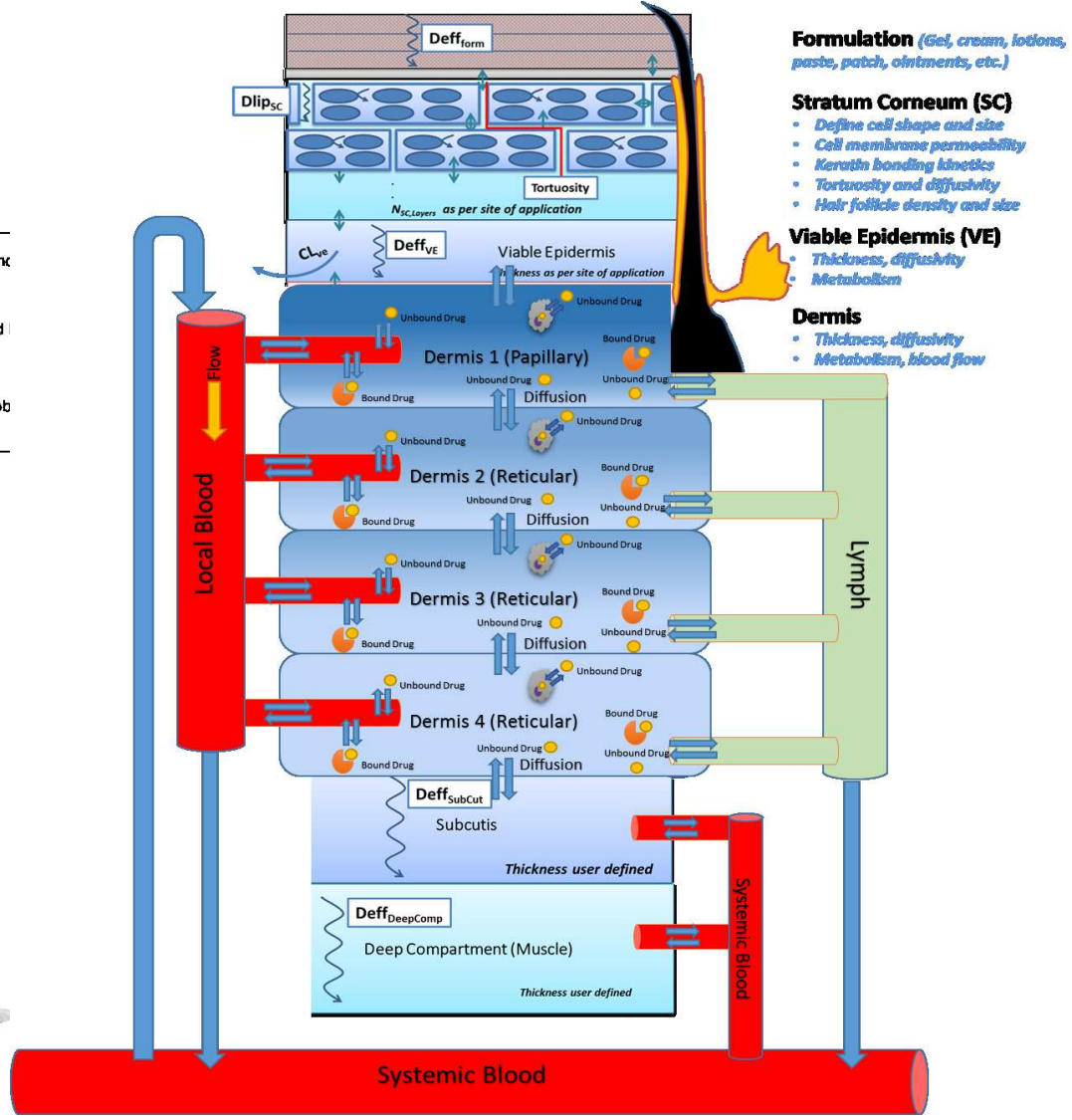
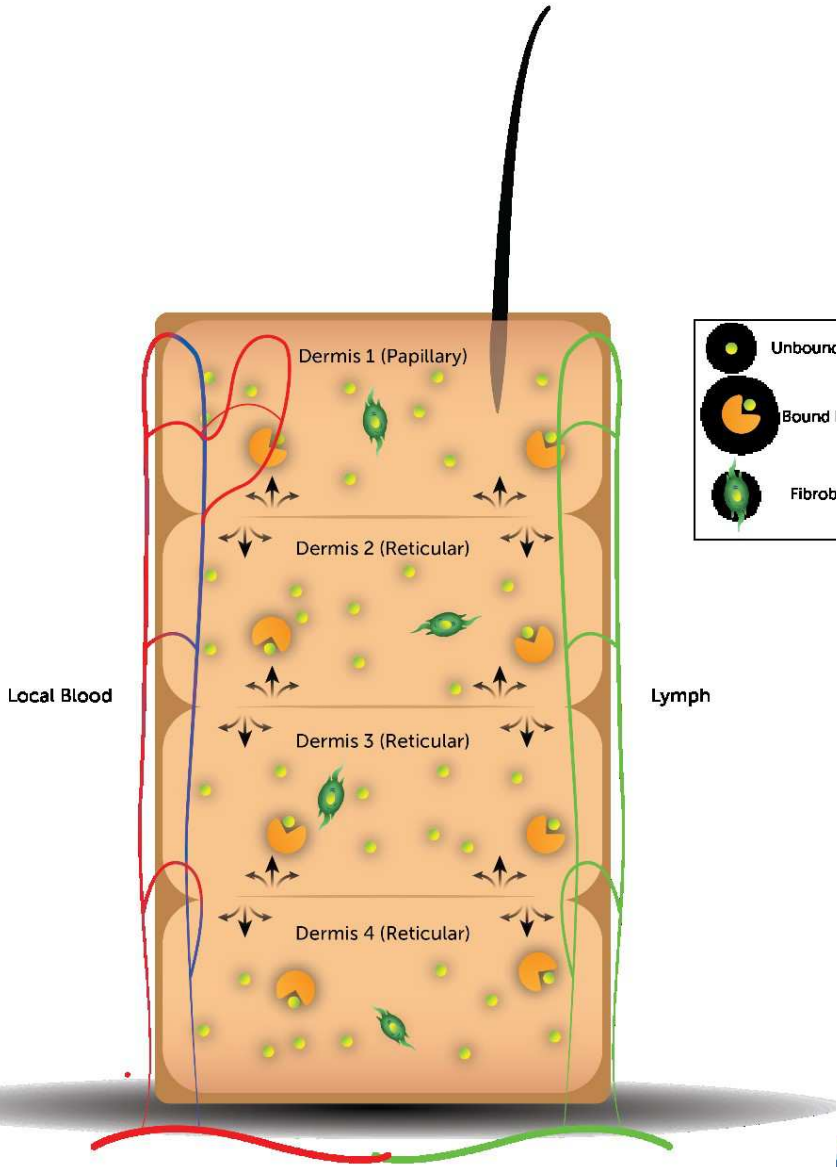
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Depth Resolved Dermis Model (DRDM)

Model Development:

- Collect physiology data
- Develop a mechanistic framework to represent this physiology
- Collect measured data for diffusion in the dermis, new mechanistic QSAR developed in house
- Verify with measured concentration-depth data

North Resolved Dermis Model



Formulation (Gel, cream, lotions, paste, patch, ointments, etc.)

Stratum Corneum (SC)

- Define cell shape and size
- Cell membrane permeability
- Keratin bonding kinetics
- Tortuosity and diffusivity
- Hair follicle density and size

Viable Epidermis (VE)

- Thickness, diffusivity
- Metabolism

Dermis

- Thickness, diffusivity
- Metabolism, blood flow

Compound Screen

SV-Alfentanil simCYT

GI Tract | Lung | **Skin** | Vaginal Tract

MechDermA Model

Multi-phase multi-layer (MPML) MechDermA Model

Single Layer Dermis Model

Depth Resolved Dermis Model

Permeability | Formulation

	Whole SC	Top 25% SC Layers	Upper Middle 25% SC Layers	Lower Middle 25% SC Layers	Bottom 25% SC Layers
Stratum Corneum (SC) Permeability Scalar	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>

Eleftheria

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Physiology

Dermis

Total Thickness (µm)

Regional dermis blood flow scalar*

DRDM Parameters

	Vessel Radii (µm)				Volume Fraction (cm ³ /cm ³) †						
	Layer Thickness (%)*	Capillary Wall Thickness (µm)	Capillary Lumen	Capillary External	Capillary †		Lymph †		Fibroblast	Excluded	Interstitial Fluid
					Mean	CV (%)	Mean	CV (%)	Mean	Mean	
Dermis 1	<input type="text" value="12.24"/>	<input type="text" value="3.5"/>	<input type="text" value="1.75"/>	<input type="text" value="5.25"/>	<input type="text" value="0.0199"/>	<input type="text" value="21.85"/>	<input type="text" value="0.0144"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.2567"/>	<input type="text" value="0.706"/>
Dermis 2	<input type="text" value="29.25"/>	<input type="text" value="4.4"/>	<input type="text" value="7.66"/>	<input type="text" value="12.06"/>	<input type="text" value="0.014"/>	<input type="text" value="37.54"/>	<input type="text" value="0.0341"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.24298"/>	<input type="text" value="0.706"/>
Dermis 3	<input type="text" value="29.25"/>	<input type="text" value="4.4"/>	<input type="text" value="7.66"/>	<input type="text" value="12.06"/>	<input type="text" value="0.014"/>	<input type="text" value="37.54"/>	<input type="text" value="0.0341"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.27298"/>	<input type="text" value="0.706"/>
Dermis 4	<input type="text" value="29.25"/>	<input type="text" value="4.4"/>	<input type="text" value="7.66"/>	<input type="text" value="12.06"/>	<input type="text" value="0.014"/>	<input type="text" value="37.54"/>	<input type="text" value="0.0341"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.24298"/>	<input type="text" value="0.706"/>

Lymph Flow Scalar

Fraction of ISF accessible by albumin

Albumin concentration in accessible spce (g/L)

* % of the total dermis thickness. † Capillary and lymph volume fractions are that occupied by lumen + endothelium. ‡ Volume of substituent per cm³ of tissue

Elefth

FDA - 091

Skin Thickness Changes in Normal Aging Skin

M.C. Branchet^a, S. Boisnic^b, C. Frances^c, A.M. Robert^a

^aLaboratoire de Biochimie du Tissu Conjonctif, UA CNRS 1174, Faculté de Médecine, Université Paris XII, Créteil; ^bService d'Anatomie et de Cytologie Pathologique, Groupe Hospitalier Pitié-Salpêtrière, ^cService de Médecine Interne, Groupe Hospitalier Pitié-Salpêtrière, Paris, France

Table 4. Evolution of the thickness (μm) of the superficial dermis with age

Age, years	Women	Men	Total
20–30	212 (72.3)	256.7 (38)	234.4 (58.5)
30–40	241.9 (44.8)	167.3 (5.2)	204.6 (50.3)
40–50	169.2 (9.5)	185.5 (20.2)	174.6 (11.5)
50–60	241.8 (28.3)	180.9 (21.8)	211.3 (43)
60–70	235.5 (32.4)	175 (39.7)	200.9 (46.7)
70–80	251.3 (39.5)	265.7 (76.1)	257.8 (56.2)

Values are mean with the SD given in parentheses.

Table 6. Decrease of the thickness (μm) of the dermis with age

Age, years	Women	Men	Total
20–30	1,802.9 (335)	2,284.9 (475.8)	1,995.5 (420)
30–40	1,494.4 (500)	1,606.4 (520.2)	1,550.4 (476.6)
40–50	1,869.3 (76.9)	1,230.4 (99.6)	1,602.5 (330.8)
50–60	1,519.1 (255.7)	1,460.8 (675.5)	1,495.8 (384.5)
60–70	1,381.4 (657.5)	1,305 (496)	1,330.7 (513.1)
70–80	1,204.6 (482.2)	1,347.5 (33.9)	1,245.4 (400)

Values are means with the SD given in parentheses.

Dermis Volume Fractions

The exclusion of human serum albumin by human dermal collagenous fibres and within human dermis

Joel L. BERT, Joyce M. MATHIESON and Richard H. PEARCE*
 Department of Pathology, Faculty of Medicine, University of British Columbia, 2211 Wesbrook Place,
 Vancouver, B.C., Canada V6T 1W5

(Received 20 August 1981/Accepted 20 October 1981)

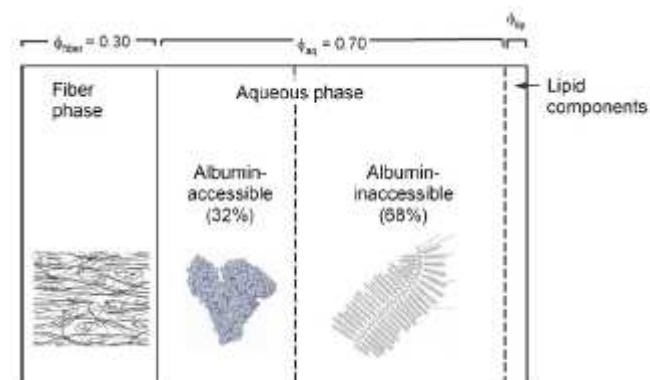


Table 4. Fluid exclusion of albumin by human dermis

Five discs of each human dermis were equilibrated with ¹²⁵I-labelled albumin in phosphate-buffered saline for 14 days at 4°C. The tissue and supernatant fluid were analysed to obtain the following data, as described in detail in the text. The water content of each tissue was determined separately using five additional discs (one was lost for D1).

Property measured	Dermis no. . . .	D1	D2	D3
Weight (g)		0.1294 ± 0.0154	0.1270 ± 0.0084	0.1337 ± 0.0118
Water (g/g fresh wt.)		0.6601 ± 0.0049	0.6241 ± 0.0070	0.6429 ± 0.0116
Collagen (g/g fresh wt.)		0.3036 ± 0.0068	0.3351 ± 0.0057	0.2906 ± 0.0075
Collagen + water (g/g fresh wt.)		0.9636 ± 0.0068	0.9592 ± 0.0057	0.9335 ± 0.0075
Collagen (g/g dry wt.)		0.893 ± 0.020	0.891 ± 0.014	0.814 ± 0.021
Swelling ratio (g of swollen tissue/g fresh wt.)		2.30 ± 0.20	1.89 ± 0.04	2.25 ± 0.17
Albumin (mg/g of free fluid)		34.36 ± 0.27	47.70 ± 0.54	35.22 ± 0.46
Fluid in swollen tissue accessible to albumin (g/g fresh wt.)		1.529 ± 0.215	1.001 ± 0.048	1.438 ± 0.197
Insoluble solids (g/g fresh wt.)		0.3177 ± 0.0056	0.3543 ± 0.0076	0.3352 ± 0.0120
Collagen (g/g of insoluble solids)		0.955 ± 0.021	0.946 ± 0.016	0.867 ± 0.022
Swollen tissue fluid (g/g fresh wt.)		1.982 ± 0.204	1.534 ± 0.044	1.918 ± 0.169
Fluid inaccessible to albumin				
(g/g fresh wt.)		0.452 ± 0.027	0.533 ± 0.012	0.475 ± 0.032
(g/g of insoluble solids)		1.424 ± 0.084	1.505 ± 0.034	1.481 ± 0.095
(g/g of collagen)		1.492 ± 0.105	1.592 ± 0.020	1.638 ± 0.135

The exclusion of human serum albumin by human dermal collagenous fibres and within human dermis

Joel L. BERT, Joyce M. MATHIESON and Richard H. PEARCE*
*Department of Pathology, Faculty of Medicine, University of British Columbia, 2211 Wesbrook Place,
 Vancouver, B.C., Canada V6T 1W5*

(Received 20 August 1981/Accepted 20 October 1981)

TABLE 2
 COMPOSITION OF DERMIS AND SERUM, AND RESULTS OF EQUILIBRATION EXPERIMENTS

Property	Mean \pm SD (No.) for dermis			
	D1	D2	D3	D4
	Composition of dermis			
Water (g/g fresh wt)	0.6474 \pm 0.0059(4)	0.6515 \pm 0.0011(4)	0.7242 \pm 0.0094(4)	0.6464 \pm 0.0035(4)
Fat (g/g fresh wt)	0.0234 \pm 0.0078(4)	0.0324 \pm 0.0045(4)	0.0383 \pm 0.0019(4)	0.0196 \pm 0.0053(4)
Insoluble solids (g/g fresh wt)	0.2812 \pm 0.0052(4)	0.2768 \pm 0.0088(4)	0.2052 \pm 0.0083(3)	0.3004 \pm 0.0071(4)
Water + fat + insoluble solids (g/g fresh wt)	0.9520 \pm 0.0029(4)	0.9606 \pm 0.0043(4)	0.9639 \pm 0.0045(3)	0.9663 \pm 0.0047(4)
Interstitial space, W_1 (g/g fresh wt)	0.6954 \pm 0.0036(4)	0.6909 \pm 0.0053(4)	0.7561 \pm 0.0064(3)	0.6801 \pm 0.0072(4)
Collagen (g/g fresh wt)	0.2467 \pm 0.0180(2)	0.2658 \pm 0.0073(3)	0.1990 \pm 0.0016(3)	0.2756 \pm 0.0051(3)
(g/g insoluble solids)	0.878 \pm 0.064(2)	0.960 \pm 0.027(3)	0.970 \pm 0.008(3)	0.918 \pm 0.017(3)
Albumin (mg/g fresh wt)	7.00 \pm 0.37(4)	8.31 \pm 0.24(4)	6.13 \pm 0.33(4)	4.48 \pm 0.27(4)
Serum albumin (mg/g)	37.33 \pm 1.67(2)	37.86 \pm 0.13(2)	35.54 \pm 0.81(2)	32.64 \pm 0.42(2)
	Equilibration experiments			
Tissue weight (g)	0.1336 \pm 0.0115(3)	0.1269 \pm 0.0188(3)	0.1359 \pm 0.0045(3)	0.1373 \pm 0.0167(3)
Swelling ratio (g swollen tissue/g fresh wt)	1.110 \pm 0.043(3)	1.196 \pm 0.048(3)	1.018 \pm 0.023(3)	1.059 \pm 0.031(3)
Recovery of added radioactivity	0.998 \pm 0.040(2)	1.015 \pm 0.011(2)	1.062 \pm 0.040(3)	1.021 \pm 0.046(3)

BERT, PEARCE, AND MATHIESON

Capillary Density

NOZOL

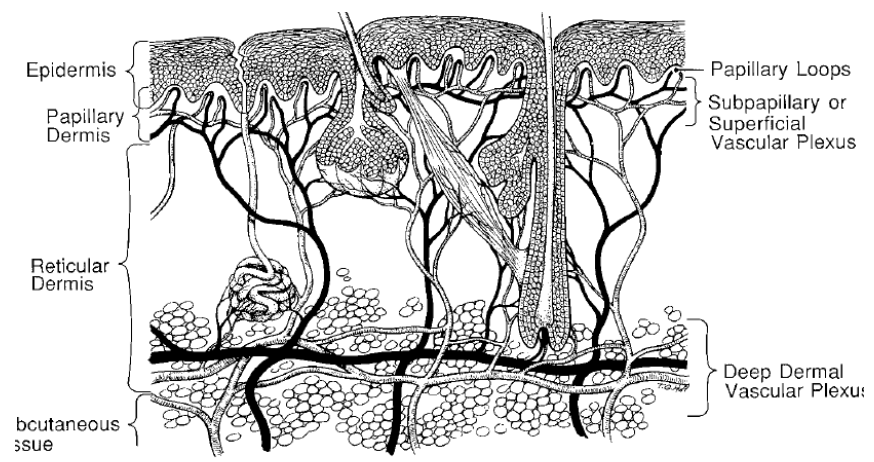


FIG. 1. Schema of the vasculature in two levels of the dermis: papillary and reticular dermi

TABLE III

Differences in Capillary Density of Papillary Dermis Between Head-Face/Neck and Other Regions of the Body Across Six Cadavers

Regions	Grouped Biopsy Sites	Mean ± SD	Significance (p)*
Head Face Neck	1, 2, 3, 4, 5	3.16 ± 0.86	—
Chest Back	6, 15	1.94 ± 0.76	p < 0.11
Arm Forearm Hand	7, 9, 10	1.98 ± 0.81	p < 0.18
Abdomen Buttock	11, 16	1.96 ± 0.46	p < 0.006
Thigh (anterior and posterior)	12, 17	1.48 ± 0.53	p < 0.007
Lower leg	13, 14, 18, 19	1.69 ± 0.70	p < 0.0006

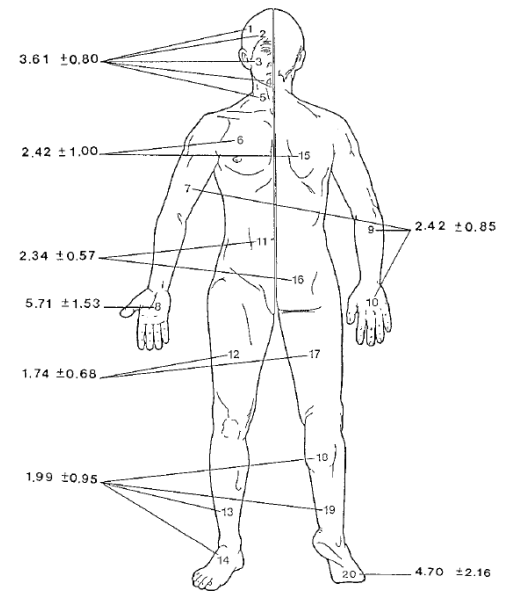
* Significant if p < 0.05.

TABLE IV

Differences in Capillary Density of Reticular Dermis Between Head-Face/Neck and Other Regions of the Body Across Six Cadavers

Regions	Grouped Biopsy Sites	Mean ± SD	Significance (p)*
Head Face Neck	1, 2, 3, 4, 5	1.20 ± 0.42	—
Chest Back	6, 15	0.48 ± 0.27	p < 0.0002
Arm Forearm Hand	7, 9, 10	0.44 ± 0.23	p < 0.02
Abdomen Buttock	11, 16	0.38 ± 0.17	p < 0.001
Thigh (anterior and posterior)	12, 17	0.26 ± 0.23	p < 0.0004
Lower leg	13, 14, 18, 19	0.30 ± 0.25	p < 0.01

* Significant if p < 0.05.



Lymphatic Density

Eleftheria Tsakalozou

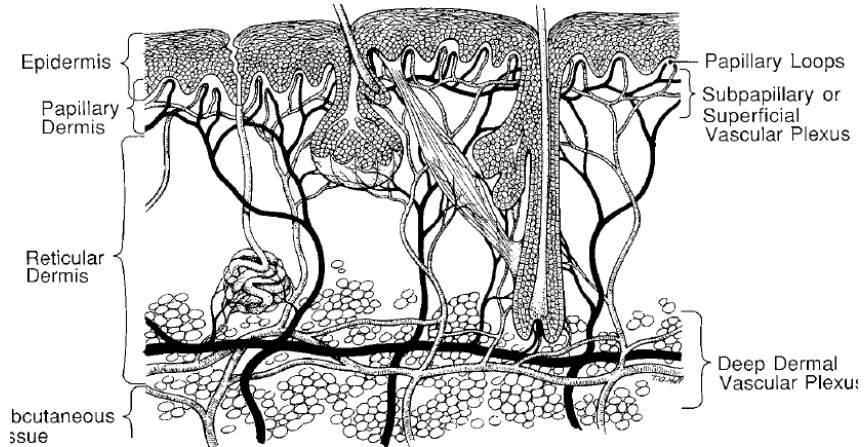


FIG. 1. Schema of the vasculature in two levels of the dermis: papillary and reticular dermi

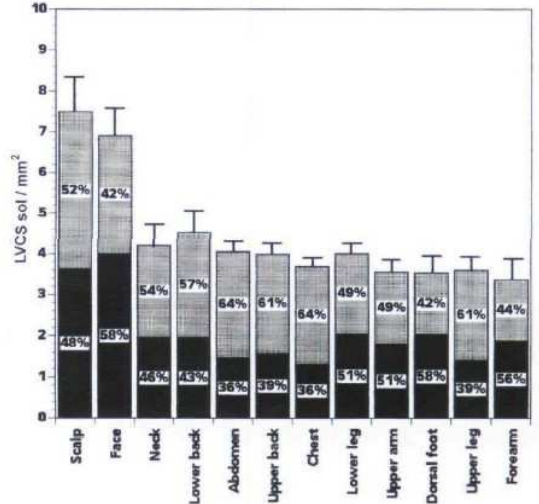
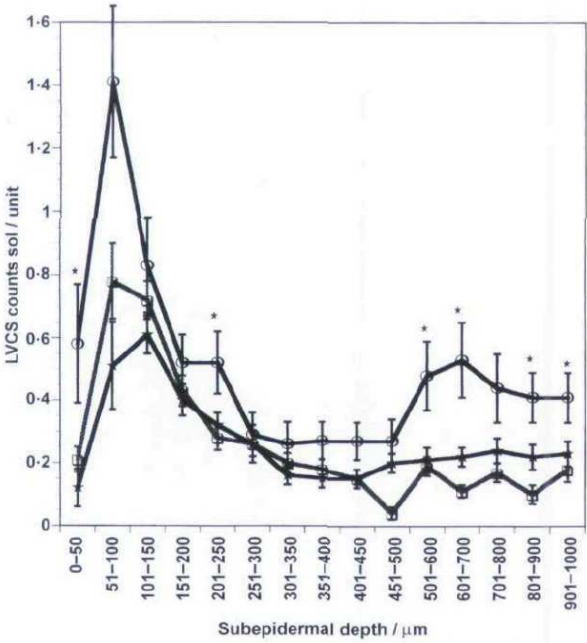


Figure 5. Number of LVCSs per mm^2 subepidermal skin by site of removal. The percentage of the uppermost quarter of the dermis and the deeper part is indicated. The error bars represent the standard error of the mean. Subepidermal depth: \square , 0-250 μm ; \blacksquare , 251-1000 μm .

Fibroblast Density

Variability of fibroblast morphology *in vivo*: a silver

Table 2.
Relative Volumes Including Variation of the Different Compartments of the Dermis

Compartment	Volume %	Reference
Cells	0.3 ± 0.1	This study
Cell water	0.21 ± 0.07	This study
ISF	40 ± 5.2	12
Lymph	0.3–0.4	Calculated from 16 and 26
Blood plasma	8 ± 0.8	14
Excluded	51.4	This study

Validation of a morphometric method for evaluating fibroblast numbers in normal and pathologic tissues

value*
 number of cell processes
 27 (1-7)
 = 100
 23 (2-5)
 = 100

Eleftheria Tsak

FDA - 09/12/19

Table 2. Fibroblast counting by image analysis in the mid-dermis of normal human donors [(a) fixed in Bouin's solution and embedded in paraffin; (b) Cryostat sections] or in hypertrophic scars [(c) fixed in Bouin's solution and embedded in paraffin]

		Normal skin (paraffin sections)									
		no.	1	2	3	4	5	6	7	8	
a)	Donors	age (years)	10	12	15	17	18	20	22	25	
Analyzed Tissue	fibroblasts per 30 fields	(52.5 × 106 μm ³)	410 ± 100	483 ± 48	588 ± 114	368 ± 51	672 ± 74	714 ± 60	436 ± 62	582 ± 78	
Native Tissue	Fibroblasts per mm ³		7800	9200	11200	7000	12800	13600	8292	11070	
Native Tissue	Fibroblasts per mm ³		2364	2788	3394	2121	3879	4121	2513	3355	
Native Tissue	(correcting factor = 1/3.3)		± 577	± 277	± 658	± 294	± 427	± 346	± 358	± 450	
		Normal skin (paraffin sections)									
b)	Donors	no.	1	2	3	4	5	6	7		
Native Tissue	Fibroblasts per mm ³		2816	3890	4444	3701	3231	4638	3975		
Native Tissue			± 75	± 206	± 188	± 79	± 150	± 213	± 181		
		Hypertrophic scars (paraffin sections)									
c)	Donors	no.	1		2						
			'normal'	'rich'	'normal'	'rich'					
Native Tissue	Fibroblasts per mm ³		4710	10 096	4823	10 890					
Native Tissue			± 562	± 639	± 579	± 398					

Fibroblast Density

Variability of fibroblast morphology *in vivo*: a silver impregnation study on human digital dermis and subcutis

G. E. K. NOVOTNY AND C. GNOTH

Department of Neuroanatomy, University of Düsseldorf, Moorenstrasse 5, 4000 Düsseldorf 1, Germany

(Accepted 5 April 1991)

Validation of a morphometric method for evaluating fibroblast numbers in normal and pathologic tissues

Table 1. Measurements of fibroblasts in digital skin of a 54-year-old male*

	Cell body size (µm)		Length of longest cell process (µm)	Number of cell processes
	Greater diameter	Lesser diameter		
Papillary dermis	7.30 (1.8)	3.7 (1.0)	22.15 (5.9)	5.27 (1.7)
	(n = 46)		(n = 100)	(n = 100)
Subcutis	15.6 (4.5)	8.2 (2.8)	75.9 (21.8)	6.3 (2.5)
	(n = 50)		(n = 100)	(n = 100)

* Means with SD in parentheses.

Table 2. Fibroblast counting by image analysis in the mid-dermis of normal human donors [(a) fixed in Bouin's solution and embedded in paraffin; (b) Cryostat sections] or in hypertrophic scars [(c) fixed in Bouin's solution and embedded in paraffin]

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	(correcting factor = 1/3.3)		± 577	± 277	± 658	± 294	± 427	± 346	± 358	± 450	

		Normal skin (paraffin sections)							
		no.	1	2	3	4	5	6	7
b)	Donors								
Native Tissue	Fibroblasts per mm ³		2816	3890	4444	3701	3231	4638	3975
			± 75	± 206	± 188	± 79	± 150	± 213	± 181

		Hypertrophic scars (paraffin sections)				
		no.	1 'normal'	'rich'	2 'normal'	'rich'
c)	Donors					
Native Tissue	Fibroblasts per mm ³		4710	10 096	4823	10 890
			± 562	± 639	± 579	± 398

Table 2. Relative Volumes Including Variation of the Different Compartments of the Dermis

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Excluded	51.4	This study

Compound Screen

SV-Alfentanil simCYT

GI Tract | Lung | **Skin** | Vaginal Tract

MechDermA Model

Multi-phase multi-layer (MPML) MechDermA Model

Single Layer Dermis Model

Depth Resolved Dermis Model

Permeability | Formulation

	Whole SC	Top 25% SC Layers	Upper Middle 25% SC Layers	Lower Middle 25% SC Layers	Bottom 25% SC Layers
Stratum Corneum (SC) Permeability Scalar	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>

Eleftheria

FDA - 09/12/2019

DRDM Screens

Compound Screen

Dermis Parameters (DRDM)		Global	Dermis 1	Dermis 2	Dermis 3	Dermis 4
Diffusivity (cm ² /h)	D _{unbound}	<input type="text" value="0.0066408224"/>	<input type="text" value="0.0066408224"/>	<input type="text" value="0.0066408224"/>	<input type="text" value="0.0066408224"/>	<input type="text" value="0.0066408224"/>
	D _{bound}	<input type="text" value="7.2E-05"/>	<input type="text" value="7.2E-05"/>	<input type="text" value="7.2E-05"/>	<input type="text" value="7.2E-05"/>	<input type="text" value="7.2E-05"/>
Fraction Unbound in Dermis ISF (fu _{ISF})		<input type="text" value="0.9417925126"/>	<input type="text" value="0.9417925126"/>	<input type="text" value="0.9417925126"/>	<input type="text" value="0.9417925126"/>	<input type="text" value="0.9417925126"/>
$D_{\text{effective}} = fu_{\text{ISF}} * D_{\text{unbound}} + (1 - fu_{\text{ISF}}) * D_{\text{bound}}$						
Effective Diffusivity (cm ² /h)		<input type="text" value="0.0062584677"/>	<input type="text" value="0.0062584677"/>	<input type="text" value="0.0062584677"/>	<input type="text" value="0.0062584677"/>	<input type="text" value="0.0062584677"/>
Fraction non-ionized (pH 7.4) (fni _{Dermis})		<input type="text" value="0.8881842302"/>	<input type="text" value="0.8881842302"/>	<input type="text" value="0.8881842302"/>	<input type="text" value="0.8881842302"/>	<input type="text" value="0.8881842302"/>
Capillary Wall Permeability (P _{mem}) (cm/s)		<input type="text" value="3.0978466899"/>	<input type="text" value="3.0978466899"/>	<input type="text" value="3.0978466899"/>	<input type="text" value="3.0978466899"/>	<input type="text" value="3.0978466899"/>
$PS = P_{\text{mem}} * SA_{\text{Exchange}} * f_{\text{niDermis}}$						
Effective Permeability Surface Area Product (PS) (ml/h per cm ³)		<input type="text" value="9.9341270876"/>	<input type="text" value="2.6344528872"/>	<input type="text" value="2.6344528872"/>	<input type="text" value="2.6344528872"/>	<input type="text" value="2.6344528872"/>
Fraction Unbound Intracellular Fibroblast (fu _{cell})		<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>	<input type="text" value="1"/>

Elef FDA -

Capillary Permeability

Unbound

Capillary endothelium permeability (Ibrahim *et al.*, 2012) :

$$\log P_{\text{mem}} = 1.64 \log K_{\text{oct}} - 1.37 \text{MW}^{1/3} + 2.82$$
$$n = 37; s = 0.71; r^2 = 0.8951 \quad (24)$$

Permeability Surface Area Product = Capillary permeability * Exchange Surface area * f_{ni}

Bound

Two pore theory, matched with our biologics module (negligible impact for small molecules)

Removal by the lymph (<1% of the clearance even for highly bound drugs)

Could be more important for larger drugs, i.e., delivered by microneedles or intradermal injection

IBRAHIM, R., NITSCHKE, J. M. & KASTING, G. B. 2012. Dermal clearance model for epidermal bioavailability calculations. *J Pharm Sci*, 101, 2094-108.

Diffusion in the Dermis

Partitioning, diffusivity and clearance of skin permeants in mammalian dermis

Kosmas Kretsos¹, Matthew A. Miller, Grettel Zamora-Estrada, Gerald B. Kasting*

College of Pharmacy, University of Cincinnati Academic Health Center, P.O. Box 670004, Cincinnati, OH 45267-0004, USA

Received 27 October 2005; received in revised form 1 June 2007; accepted 5 June 2007

Available online 23 June 2007

***In Silico* Prediction of Percutaneous Absorption and Disposition Kinetics of Chemicals**

Longjian Chen • Lujia Han • Ouarda Saib • Guoping Lian

Current Model:
(Modified Chen)

$$D_{de}(\text{m}^2/\text{s}) = \frac{10^{-8.15-0.655\log(MW)}}{0.68 + \frac{0.32}{f_u} + 0.025K_{mw}}$$

Diffusion in the Dermis

- Measured effective diffusivity value were collected form the literature. N = 34.

Mechanistic framework:

$$D_{Dermis} = f u_{ISF} * D_{unbound} + (1 - f u_{ISF}) * D_{Bound} \quad (1)$$

Diffusion in the Dermis

Wilke Chang 1955:

$$D_{unbound} (\text{cm}^2/\text{s}) = \left(\frac{7.4 * 10^{-10} * T_{Dermis} * (MW\phi_{ISF})^{0.5}}{\eta_{ISF} * V_A^{0.6}} \right) \quad (2)$$

Where:

T_{Dermis} = Absolute Temperature in the Dermis Value = 310.15 K

$MW\phi_{ISF}$ = Solvent Molecular Weight * Association parameter (assumed the same as water);
Value = 40.68 Da

η_{ISF} = Viscosity of ISF (assumed the same as lymph measured by Bouta et al., 2014);
Value = 0.0181 dyne × s/cm²

V_A = Molar Volume of the Molecule (predicted from MW by method described in Ibrahim et al., 2012).

Diffusion in the Dermis

Diffusion of bound drug assumed to be the same as albumin

Direct measurement of interstitial convection and diffusion of albumin in normal and neoplastic tissues by fluorescence photobleaching

(fluorescence recovery/interstitial fluid flow/macromolecular diffusion)

SRIKANTH R. CHARY AND RAKESH K. JAIN*

Department of Chemical Engineering, Carnegie Mellon University, Pittsburgh, PA 15213-3890

Communicated by John M. Prausnitz, May 1, 1989

Self-diffusion in a hyaluronic acid–albumin–water system as studied by NMR

Andrey Filippov,^{a,b,*} Marina Artamonova,^a Maya Rudakova,^a Roustam Gimatdinov^a and Vladimir Skirda^a

Plasma pharmacokinetics and interstitial diffusion of macromolecules in a capillary bed

LAWRENCE J. NUGENT AND RAKESH K. JAIN

Department of Chemical Engineering, Carnegie-Mellon University, Pittsburgh, Pennsylvania 15213

Aqueous Diffusivity = 0.0032 cm²/h

Effective Diffusivity in ISF = 7.2 x 10⁻⁵ cm²/h

Diffusion in the Dermis

$f u_{ISF}$

$$f u_{Dermis} > f u_{2\% \text{ solution}} > f u_{4\% \text{ solution}} > f u_{plasma} > f u_{predicted}$$

The fraction of drug unbound to albumin in the interstitial fluid of the dermis can be predicted by Equation 3.

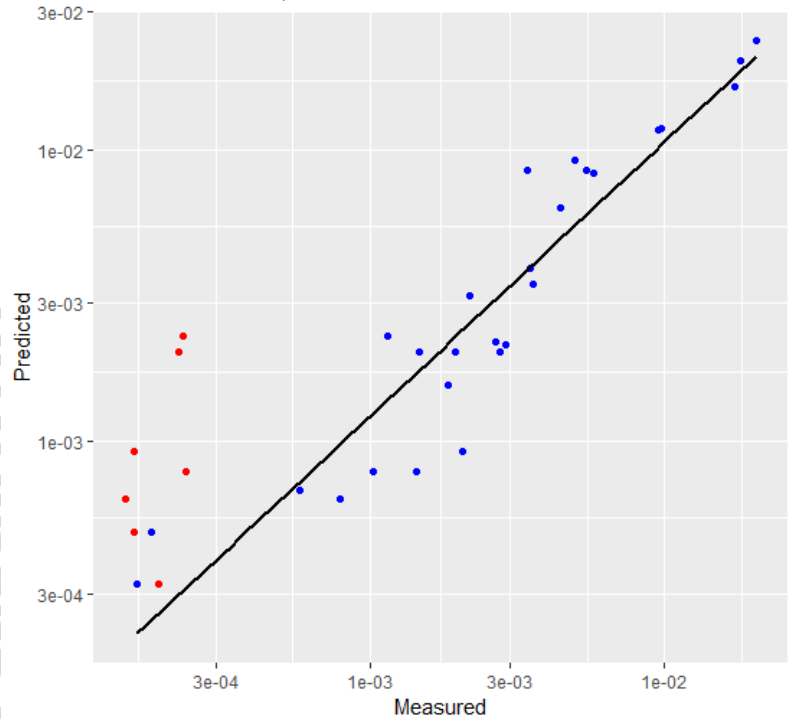
$$f u_{ISF} = \frac{1}{1 + \frac{[P]_{Dermis}}{K_D}} \quad (3)$$

$[P]_{Dermis}$ = The abundance of albumin in the dermis ISF; Value = 454.3 μM (Bert et al., 1986)

K_D = The affinity constant between the drug and albumin. This can be calculated by rearranging Equation 3 and replacing $f u_{ISF}$ and $[P]_{Dermis}$ with values for plasma, where measured values of $f u_{plasma}$ are available.

Diffusion in the Dermis

Full dataset , N=27

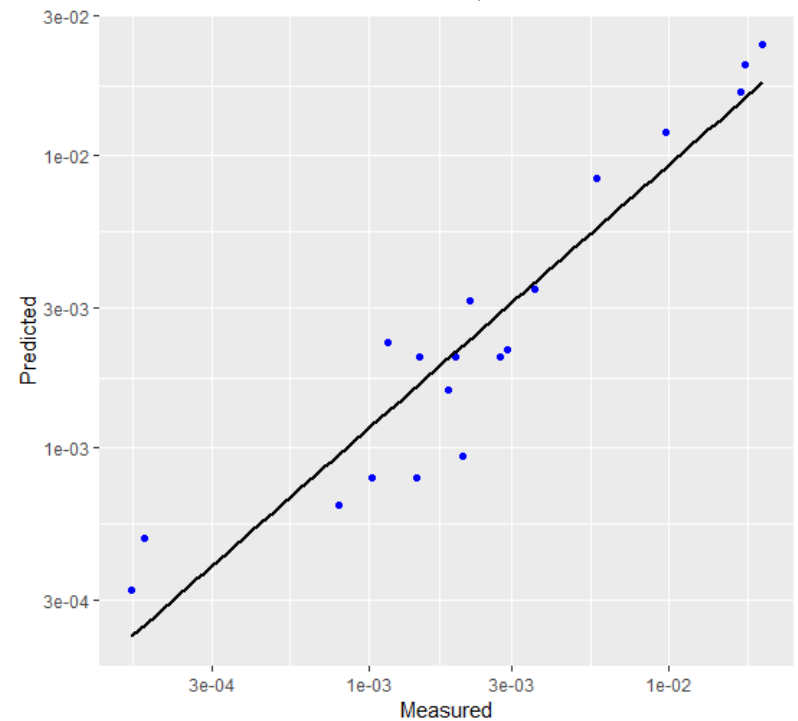


Clarke 2019:
 $R^2 = 0.94$
 Slope = 1.12
 NRMSE = 9.4%

Modified Chen:
 $R^2 = 0.96$
 Slope = 1.17
 NRMSE = 9.1%

(Trained with 21 of these observations)

Subset for Measure fu, N=19



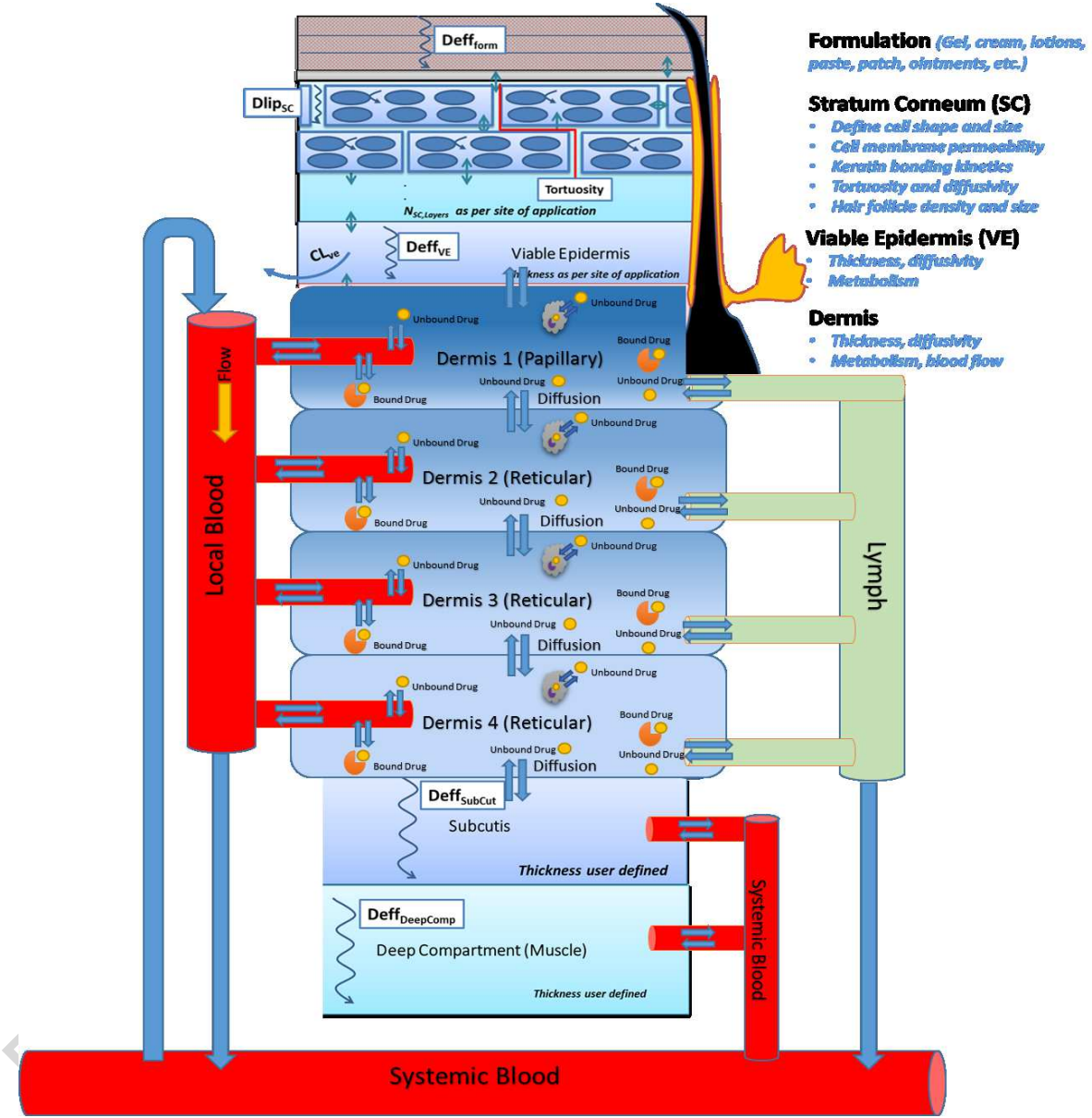
Clarke 2019:
 $R^2 = 0.98$
 Slope = 1.12
 NRMSE = 6.6%

Modified Chen:
 $R^2 = 0.98$
 Slope = 1.18
 NRMSE = 8.3%

(Trained with 13 of these observations)

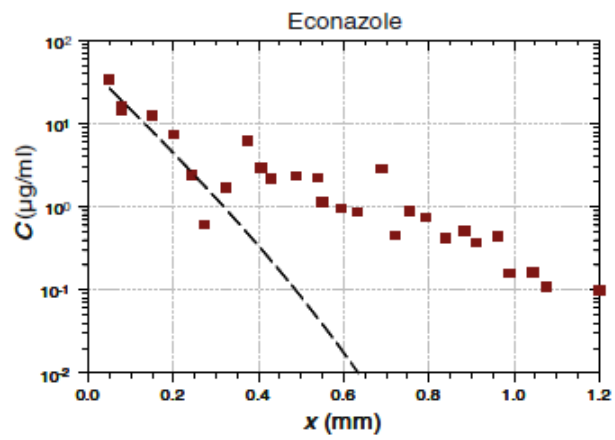
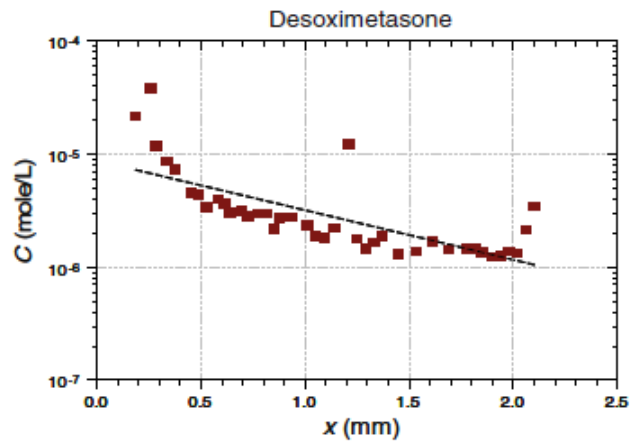
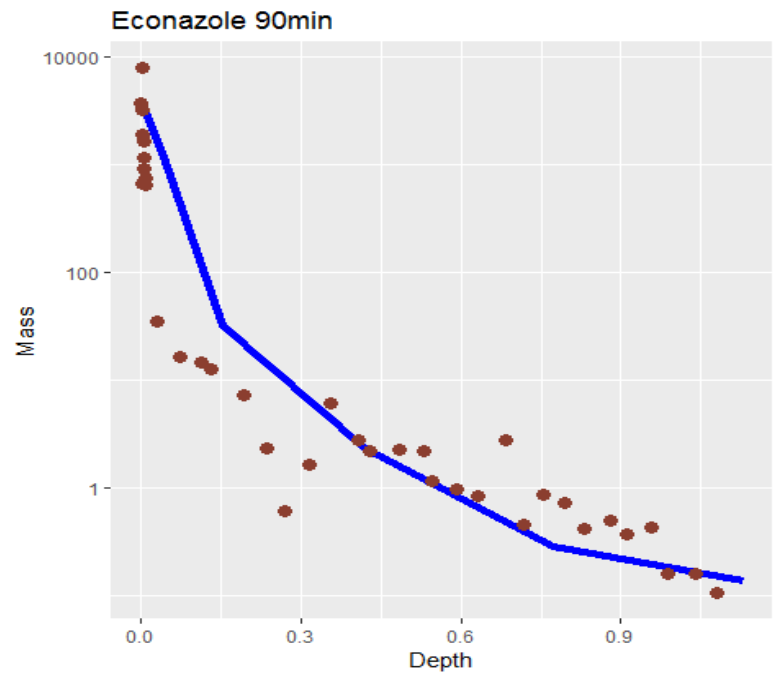
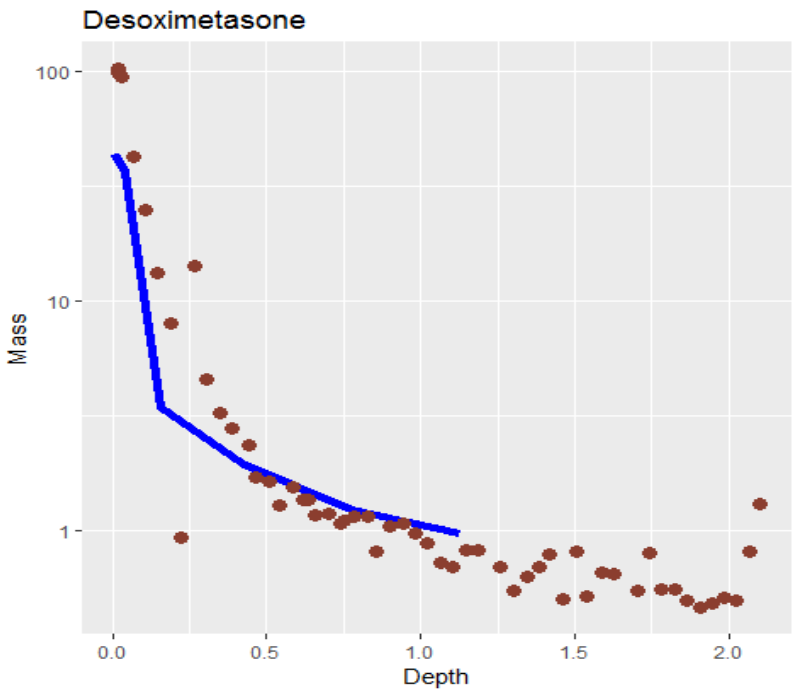
Depth Resolved Dermis Model (DRDM)

Eleftheria Tsakalozou



Initial DRDM Verification (in R prototype model)

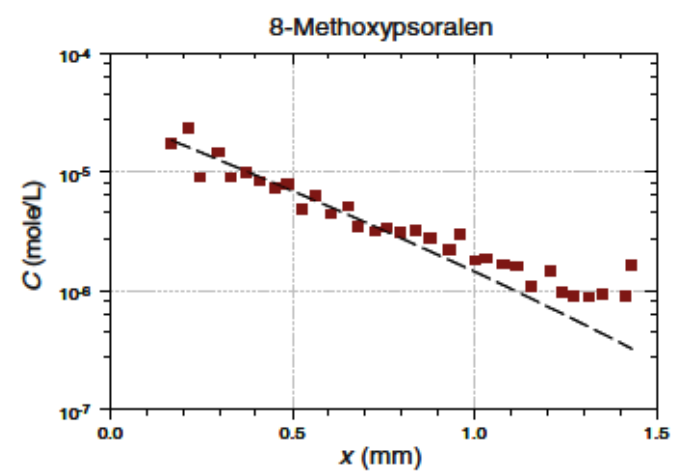
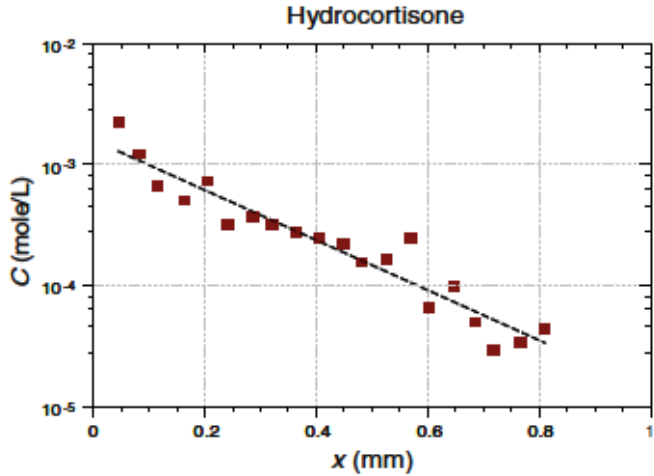
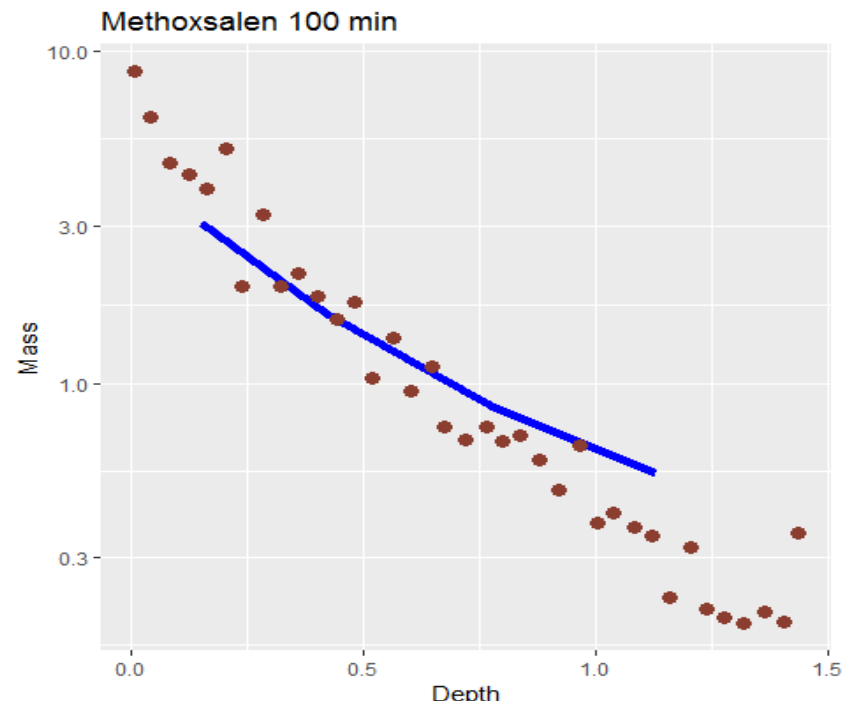
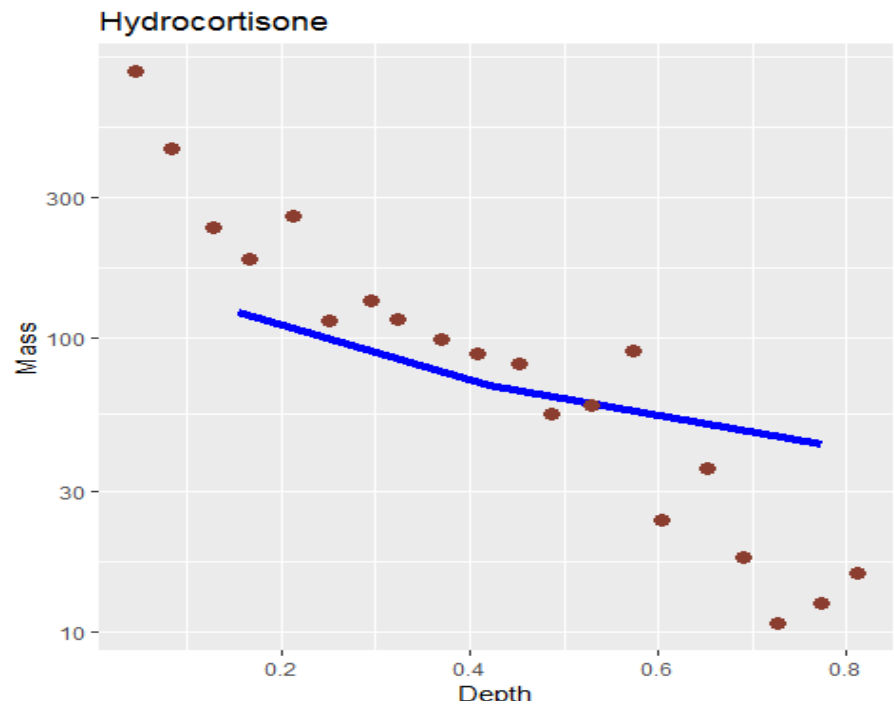
C_{initial} was fitted, as these studies had limited information about dosing or formulation



Anissimov & Roberts

Initial DRDM Verification (in R prototype model)

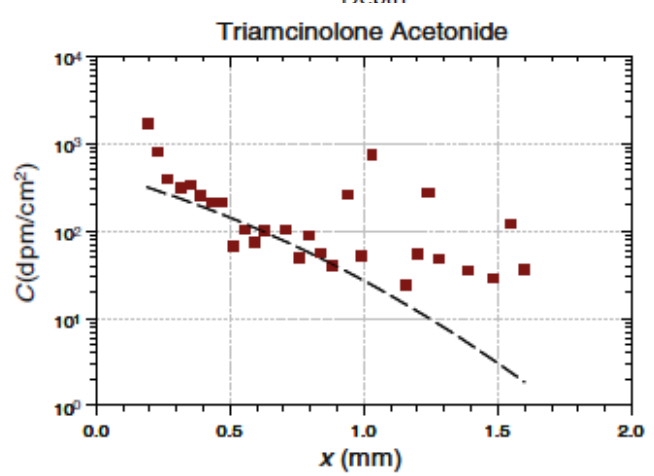
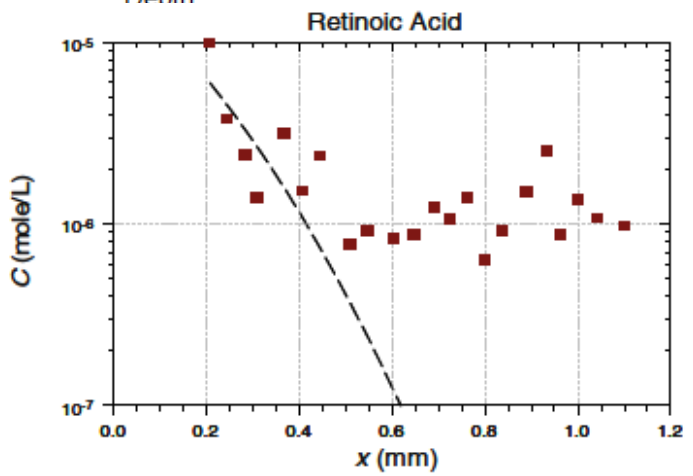
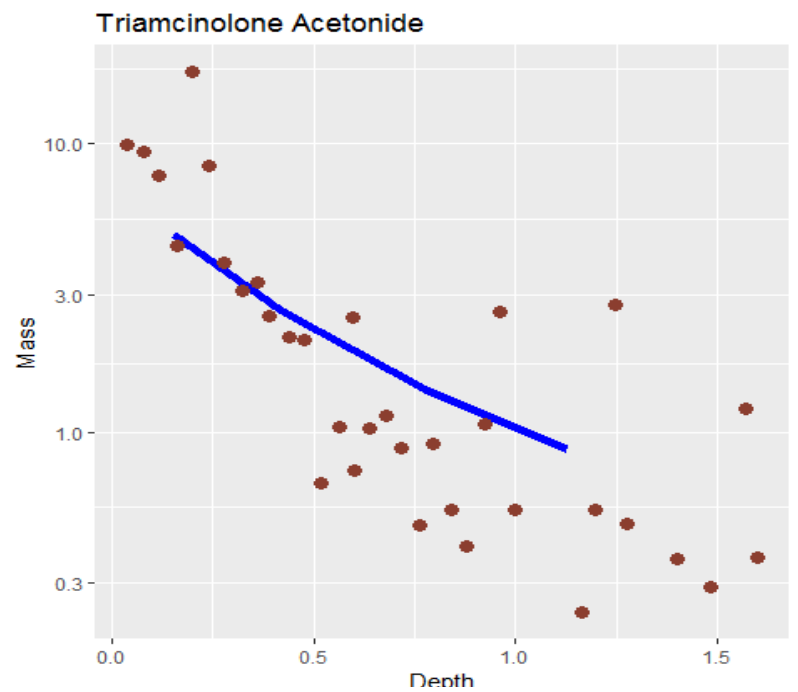
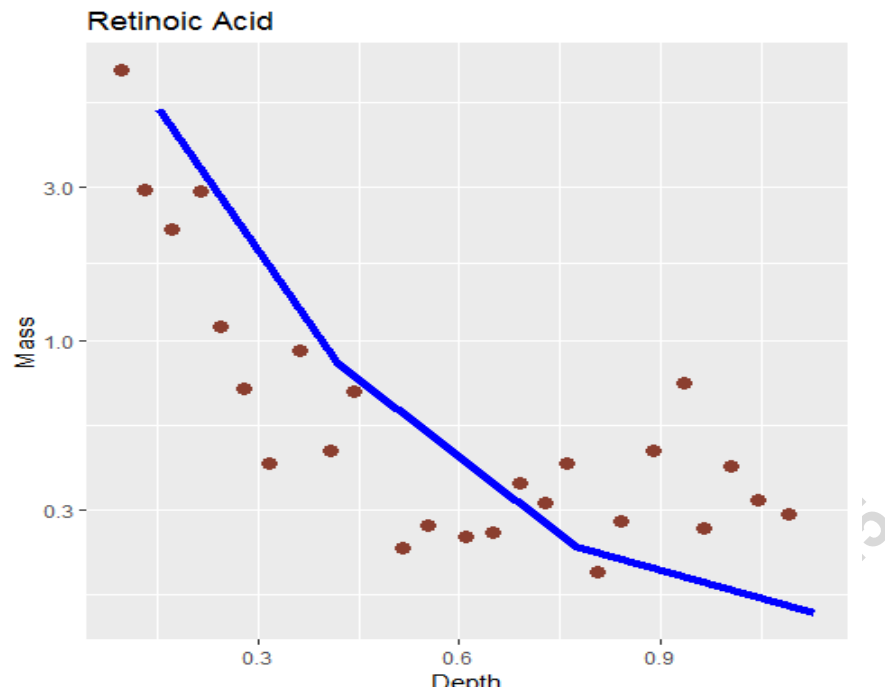
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Anissimov
& Roberts

Initial DRDM Verification (in R prototype model)

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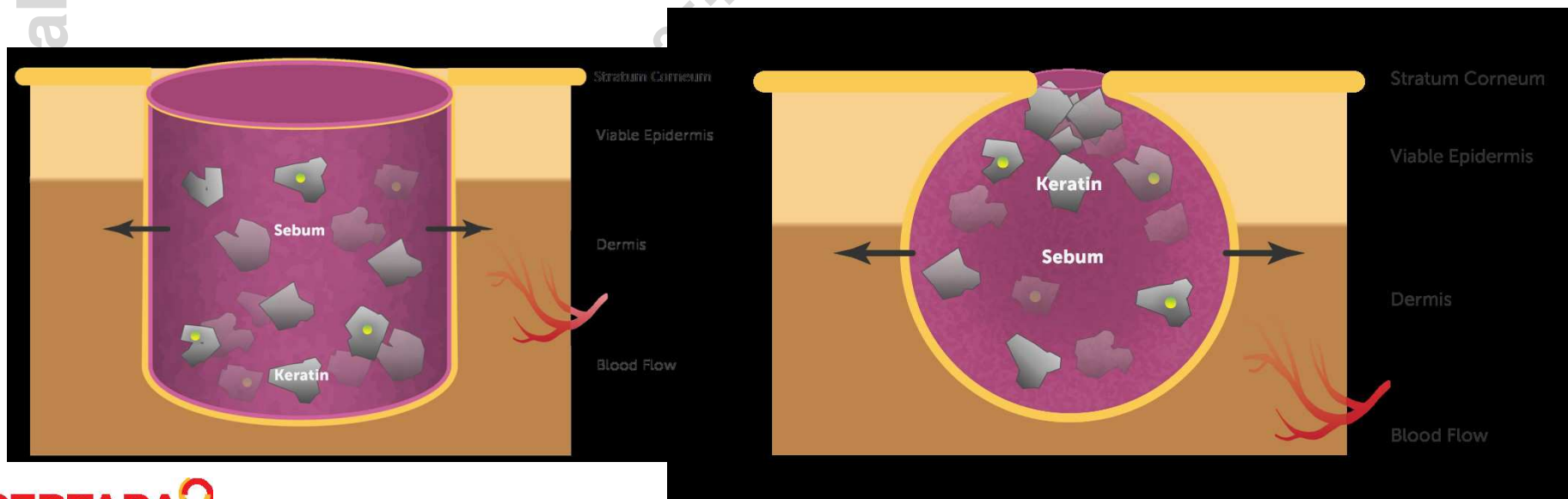
Eleftheria
Anissimov
& Roberts

Comedone Model



Comedones

- Drug partition into sebum based on the accessible area
- Drug can bind to keratin in the Comedone
- Drug can only partition out of the Comedone into the Dermis
- When DRDM is active, Comedone can end in any of the four compartments based on the defined volume
- Within each individual, Comedones are all the same size. But different individuals can have different volume.



Comedones - Screens

Sim-Healthy Volunteers



MechDerma Model Multi-Phase Multi-Layer (MPML) MechDerma Model

Regional subcutis blood flow scalar*

Muscle

Thickness (mm)

Regional muscle blood flow scalar*

Comedones

Closed Comedones

Mean CV (%)

Number per cm²

Pore Radius (μm)

Volume (μL)

D sebum Scalar

Keratin Volume Fraction***

Open Comedones

Mean CV (%)

Number per cm²

Pore Radius (μm)

Volume (μL)

D sebum Scalar

Keratin Volume Fraction***

Cracks in the Stratum Corneum

Mean CV (%)

Number per cracks per cm²

Width of cracks (μm)

Depth of cracks (% of total SC thickness)

*Regional blood flow scalar is a multiplier to the area corrected total skin tissue blood flow to account for regional differences from average skin tissue blood flow

**Tortuosity value is calculated using the Johnson model based on the PopRep corneocyte dimensions of the top 25% SC layer. This tortuosity model is recommended to be used when the diffusion coefficient SC lip (DC_{SClip}) is calculated by the Johnson 1996 model.

***Used in conjunction with K_D keratin to predict binding

Lua Scripting

Custom Dermal - Physiology Parameter(s)

Elefti FDA - 03

Comedones - Screens





Addition of keratin volume fraction on the population screen

Corneocyte width (μm)		Corneocyte length (μm)		Hydration level (% water volume)		Keratin volume fraction	SC lipid viscosity (centipose)	
Mean	CV (%)	Mean	CV (%)	Mean	CV (%)	Mean	Mean	CV (%)
27.7	9.2	34.2	4.1	35.5	17	0.645	75	0
27.7	9.2	34.2	4.1	46.1	11	0.539	75	0
27.7	9.2	34.2	4.1	56.75	7	0.4325	75	0
27.7	9.2	34.2	4.1	67.4	4	0.326	75	0


Addition of an option to use K_D as opposed to f_u . This will make binding dependent on keratin volume fraction. And therefore each bin in the SC can have different binding.


Keratin Binding Kinetics

Steady State

fusc Fraction Unbound in SC  0.1949986504   $K_D, \text{keratin}$  0.0006729879

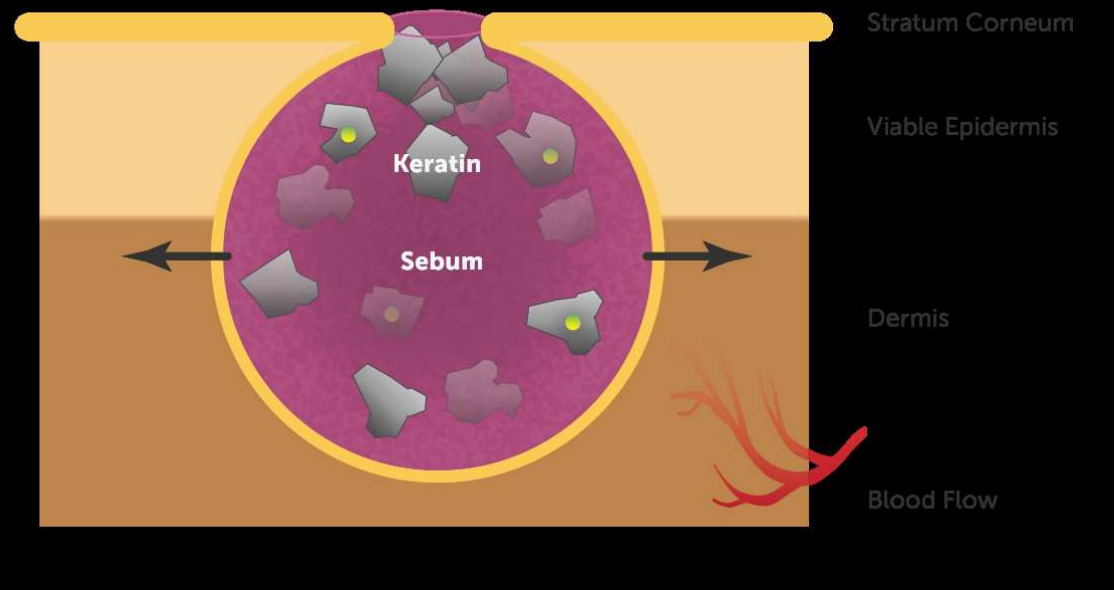
Dynamic Adsorption/Desorption Kinetics

Corneocyte membrane permeability (cm/h)  1E-05

Fraction of drug non-ionised in corneocyte ($f_{ni, \text{corn}}$)  0.6661394245 CV (%) 30

Closed Comedones

- Modelled as a truncated sphere



✔ Comedones

	Closed Comedones	
	Mean	CV (%)
Number per cm ²	0	40
Pore Radius (μm)	65.4	83
Volume (μL)	0.56	43
<i>D_{sebum}</i> Scalar	1	0
Keratin Volume Fraction***	1E-05	0

Open Comedones

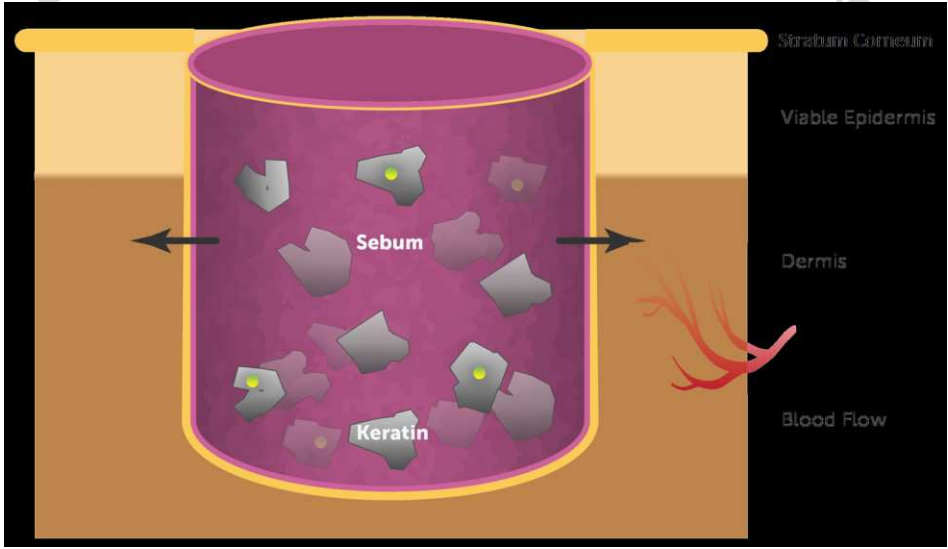
- Modelled as a cylinder

Comedones

	Closed Comedones		Open Comedones	
	Mean	CV (%)	Mean	CV (%)
Number per cm ²	0	40	0	40
Pore Radius (μm)	65.4	83	500	40
Volume (μL)	0.56	43	0.56	43
<i>D sebum</i> Scalar	1	0	1	0
Keratin Volume Fraction***	1E-05	0	1E-05	0

kaloizou

45



CERTARA[®]

Simcyp

Updates to Psoriasis Dermal Population for V19



Psoriasis Dermal

First implemented in Version 17 of the Simcyp Simulator.

This population file describes the involved skin of patients with Plaque Psoriasis.

If a population parameter is being updated, there are three possible reasons for this:

- New Data has become available in the literature after the last literature search was completed in 2016.
- New Data has been found that was not picked up in the 2016 literature search
- New Models are available in Simcyp that can now be parameterized for Psoriasis Plaque

Parameters altered in a Psoriatic Plaque

- Surface pH ↓
- Surface Temperature ↑
- Corneocyte Surface Area ↓
- Corneocyte thickness ↑
- Number of Stratum Corneum layers ↑
- Cracks in the Stratum Corneum
- Tortuosity ↓
- Stratum Corneum hydration ↓
- Viable Epidermis Thickness ↑
- Dermis Thickness ↑
- Capillary Volume Fraction in the Dermis ↑
- Lymph Volume Fraction in the Dermis ↑
- Lymph Flow in the Dermis ↑
- Interstitial Fluid Volume Fraction in the Dermis ↑
- Blood Flow in the Dermis ↑
- Blood Flow in the Subcutis ↑
- Albumin concentration in plasma ↓
- Albumin concentration in the Dermis ↓

New for V19

Reanalysed for V19

Same as V17

Surface Temperature

CUTANEOUS BLOOD FLOW IN PSORIASIS MEASURED BY ¹³³XENON CLEARANCE*

ALLAN NYFORS, M.D. AND HANS W. ROTHENBERG, M.D., D.P.H. (LONDON)

Increased average temperature, Mean 33.7°

TABLE I

Blood flow calculated from the initial slope in curves recorded for 20-30 minutes

Patients			Skin blood flow (ml/100 g X min)		Skin temperature			Controls			Skin blood flow
No./name	Age	Sex	normal	psoriasis	normal	psoriasis	diff.	No./name	Age	Sex	
1. H.O.	64	m	6.5	8.8	31.6	33.0	1.4	1. N.D.	48	f	3.1
2. S.C.	23	m	16.2	24.3	31.0	32.5	1.5	2. V.H.	58	f	4.4
3. F.M.	66	m	8.8	13.9	33.0	35.5	2.5	3. E.J.	67	f	5.4
4. A.L.	59	f	4.1	6.6	32.0	34.0	2.0	4. J.N.	41	f	6.1
5. A.N.	18	f	3.7	6.5	32.0	33.0	1.0	5. N.M.	31	m	6.1
6. S.P.	13	f	3.5	6.4	31.0	31.5	0.5	6. N.C.	42	f	7.5
7. E.H.	44	m	6.8	12.1	31.5	35.0	3.5	7. N.M.	27	f	8.1
8. J.P.	36	f	4.8	9.7	34.0	35.0	1.0	8. N.W.	62	m	8.1
9. H.B.	56	m	13.1	26.9	—	—	—	9. P.N.	18	m	8.5
10. E.L.	59	m	2.0	4.4	33.2	33.4	0.2	10. M.A.	29	m	7.5
11. A.N.	71	f	9.2	20.2	32.6	33.8	1.2				
12. M.L.	14	f	6.1	15.1	33.0	35.0	2.0				
13. A.L.	59	m	3.6	11.8	32.0	33.0	1.0				
14. C.L.	32	f	2.9	10.3	31.8	33.2	1.4				
15. L.P.	32	f	1.1	4.5	32.0	33.5	1.5				
Total	646		92.4	181.5	450.7	471.4	20.7		423		64.8
Mean	43		6.16	12.1	32.2	33.7	1.5		42		6.5

M 6.16 12.1
s² 17.50 48.34
σ 5.73
t 3.06
0.01 > p > 0.001

Eleftheria Tsakalozou

FDA - 09/12/20-

Corneocyte Dimensions

- Corneocytes are thicker with a smaller surface area, similar volume.
- Low patient numbers, but all tell a similar story.
- We can use these dimensions to calculate tortuosity (feature added in V18)

Three-dimensional analyses of individual corneocytes with atomic force microscope: morphological changes related to age, location and to the pathologic skin conditions

Nobuo Kashibuchi¹, Yoshikazu Hirai¹, Kenichiro O’Goshi² and Hachiro Tagami²
1Pola Laboratories, Pola Chemical, Inc. Yokohama, Japan and 2Department of Dermatology, Tohoku University School of Medicine, Sendai, Japan

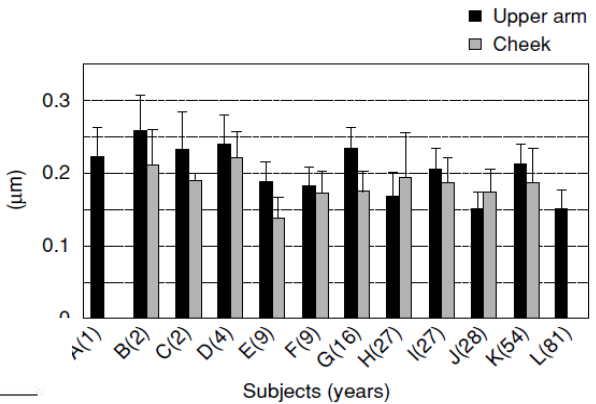
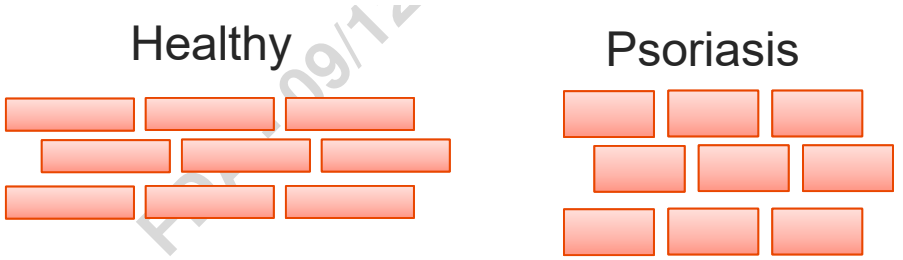


TABLE 3. Parameters of corneocytes from the involved and uninvolved sites of psoriatic patients

Parameter	Dimension	Subject 9, age 63 (Female)		Subject 10, age 48 (Female)		Subject 11, age 36 (male)	
		Involved	Uninvolved	Involved	Uninvolved	Involved	Uninvolved
Mean thickness	µm	0.389 ± 0.09	0.237 ± 0.04	0.462 ± 0.107	0.235 ± 0.041	0.827 ± 0.186	0.263 ± 0.056
Projected area	µm ²	1113 ± 139	1615 ± 88	1019 ± 135	1448 ± 115	1020 ± 123	1060 ± 89
Surface area	µm ²	1156 ± 139	1649 ± 87	1048 ± 141	1471 ± 113	1060 ± 122	1095 ± 95
Volume	µm ³	426 ± 85	382 ± 64	465 ± 101	339 ± 58	841 ± 204	276 ± 61
Flatness index		3.06 ± 1.08	7.03 ± 1.34	2.33 ± 0.62	6.36 ± 1.33	1.36 ± 0.59	4.27 ± 1.22

Mean thickness of the corneocytes of various aged subjects at various locations. Mean thickness of the corneocytes from the flexor surface of the upper arm decreased with age. In contrast, the corneocytes from the cheek showed no such a relation of its great individual differences. Data are reported as SD (n = 20 corneocytes).

Data are reported as mean ± SD, n = 20.



Corneocyte Dimensions

- Corneocytes are thicker with a smaller surface area, similar volume.
- Low patient numbers, but all tell a similar story.
- We can use these dimensions to calculate tortuosity (feature added in V18)

Surface Area Measurements of Psoriatic Corneocytes: Effects of Intralesional Steroid Therapy

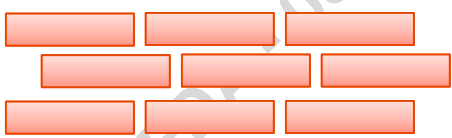
HERBERT GOLDSCHMIDT, M.D.

The Department of Dermatology, University of Pennsylvania Medical School (Duhring Laboratories), Philadelphia, Pennsylvania, U.S.A.

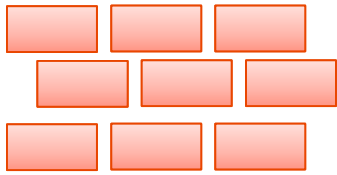
Mean corneocytes surface areas (μm^2) and standard deviations in 11 psoriatic patients before and after treatment

Patient	Age	Sex	Area	Psoriasis										
				Normal (Control)		Before treatment		After treatment						
				Mean	SD	Mean	SD	2 weeks		4 weeks		6 weeks		
1	38	M	Knee	790	102	440	101	580	100	631	84	750	96	
2	70	M	Knee	974	146	599	83	632	77	663	89	862	111	
3	76	M	Knee	805	111	551	92	568	95	756	80	801	110	
4	61	M	Knee	668	116	489	67	577	119	659	123	651	82	
5	22	M	Lower leg	604	62	560	90	583	76	612	106	602	81	
6	36	F	Lower leg	678	77	618	63	680	80	697	111	687	95	
7	49	F	Gluteal	798	58	609	89	747	124	754	129	784	76	
8	50	F	Elbow	676	89	564	108	577	61	618	75	654	100	
9	38	F	Elbow	713	97	506	68	556	80	662	88	721	58	
10	47	F	Elbow	620	101	511	62	568	66	607	76	590	98	
11	33	M	Forearm	781	94	442	68	615	101	595	78	712	82	

Healthy



Psoriasis



Corneocyte Dimensions

Surface Area Measurements of Psoriatic Corneocytes: Effects of Intralesional Steroid Therapy

HERBERT GOLDSCHMIDT, M.D.

The Department of Dermatology, University of Pennsylvania Medical School (Duhring Laboratories), Philadelphia, Pennsylvania, U.S.A.

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1	38	M	Knee	790	102	440	101	580	100	631	84	750	96
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7	49	F	Gluteal	798	58	609	89	747	124	754	129	784	76
8	50	F	Elbow	676	89	564	108	577	61	618	75	654	100
9	38	F	Elbow	713	97	506	68	556	80	662	88	721	58
10	47	F	Elbow	620	101	511	62	568	66	607	76	590	98
11	33	M	Forearm	781	94	442	68	615	101	595	78	712	82

Surface area measurements (Before Treatment) Plaque vs Healthy controls
 Ratio = 0.73 - agrees with the Kashibuchi data (0.79).

Final Ratios:

Thickness : 2.25

Surface Area : 0.75



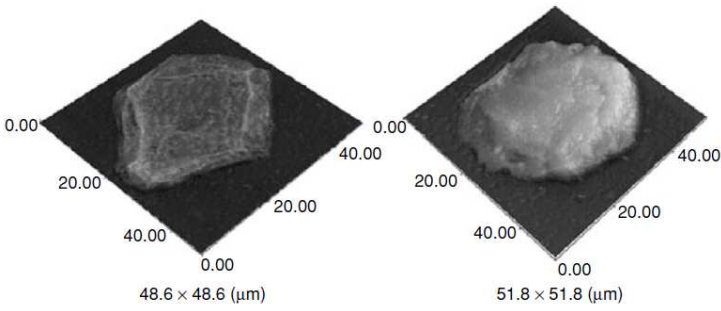
Corneocyte Dimensions

Final Ratios:

Thickness : 2.25

Surface Area : 0.75

Fig. 12. The appearance of corneocyte obtained from involved site or uninvolved site of a psoriatic patient. Corneocytes from involved site (right) had thicker and rounded form than those from uninvolved site (left). In contrast to the corneocytes from uninvolved site from AD patient, corneocytes from uninvolved site from psoriatic patient had a similar shape to normal corneocyte.

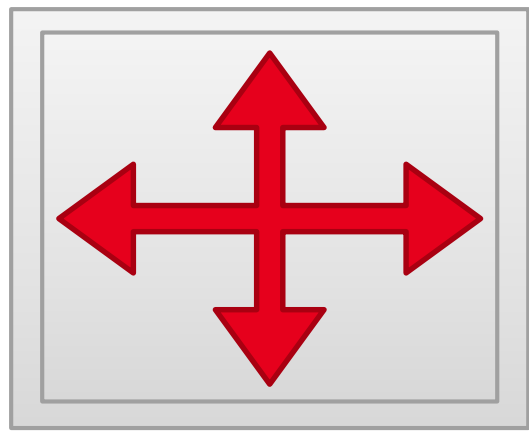


Assumptions:

- Topological Aspect ratio remains the same (to scale)
- Shape is cuboidal

Example:

Volar Forearm	(μm)	
	Healthy	Plaque
Thickness	0.59	1.35
Width	29.6	26.3
Length	36.4	32.3



Number of SC layers

Measurement of epidermal thickness in a patient with psoriasis by computer-supported image analysis

No studies measuring number of layers specifically.

Therefore calculated from SC thickness and corneocyte thickness

Mean ratio of SC thickness from healthy to plaque = **2.37**

(Palm omitted from calculation)

Table 2. Epidermal thickness measurements of psoriasis patients and controls according to biopsy location.

Biopsy location	Full epidermal thickness (µm)	Stratum corneum thickness (µm)	Rete length (µm)	Suprapapillary epidermal thickness (µm)	Dermal papilla distance (µm)
Patient elbow (11)	500	133	361	60	51
Control elbow (9)	326	73	225	88	69
Patient knee (22)	610	87	491	72	40
Control knee (16)	208	36	157	65	37
Patient leg (10)	532	91	447	53	56
Control leg (10)	195	37	159	69	45
Patient trunk (8)	365	64	307	53	46
Control trunk (7)	204	23	180	40	63
Patient palmar region (13)	663	192	388	78	35
Control palmar region (15)	207	45	162	43	46

The number of subjects in each group is given in parentheses. The data are reported as medians in micrometers.

Hydration

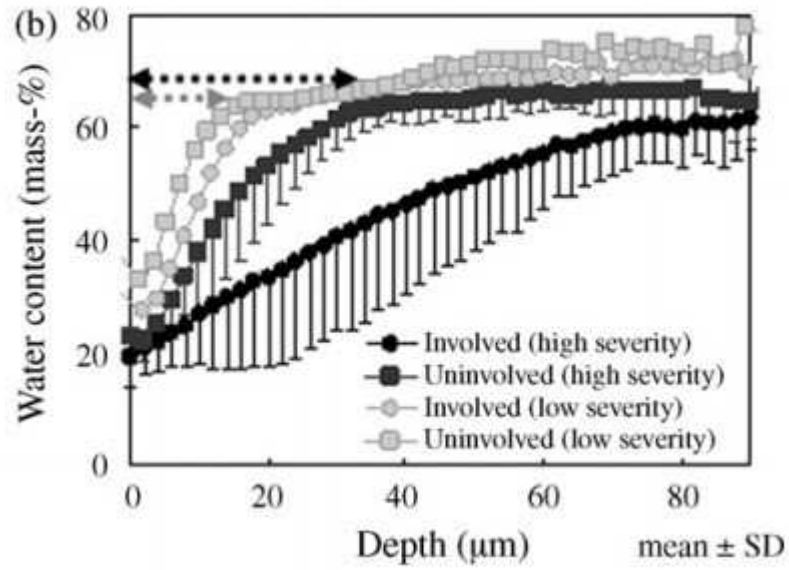
A proton NMR study on the hydration of normal *versus* psoriatic stratum corneum: linking distinguishable reservoirs to anatomical structures

Cornelia Laule^{a,b*}, Sumia Tahir^a, Charmaine L. L. Chia^a, Irene M. Vavasour^b, Neil Kitson^c and Alex L. MacKay^{a,b}

In vivo characterization of the structure and components of lesional psoriatic skin from the observation with Raman spectroscopy and optical coherence tomography: A pilot study

Table 1. Hydration of the normal and psoriatic stratum corneum (SC) samples

Sample	Hydration (g H ₂ O/g SC)
Normal	
1a	0.91
1b	0.51
Psoriatic	
1	0.25
2	0.22
3a	0.54
3b	0.54
4a	0.57
4b	0.63
4c	0.80
4d	0.42
5a	0.23
5b	0.35
5c	0.44
5d	0.43



Number of SC layers

SC N-layers was calculated from the measured thickness of total SC, corneocyte thickness and hydration.

Will also affect keratin binding as keratin content = 1 - water content

Example hydration for forearm:

	Healthy	Plaque
Bin1	33.9	29.72
Bin2	44.7	33.32
Bin3	55.5	35.81
Bin4	66.4	41.75

Example (male) :

Site	Healthy µm	Plaque µm	Healthy N Layers	Plaque N Layers
Forehead	11.68	27.68	12	16
Forearm	18.04	42.76	22	28
Outer Forearm	20.77	49.23	22	28
Upper arm	17.61	41.74	18	23
Face	8.74	20.72	13	17
Low Leg	25.68	60.87	22	28
Upper Leg	21.06	49.92	18	23
Back	14	33.18	17	22

Tortuosity

Tortuosity of lipid diffusion pathway of SC User Input

12.7

18

Johnson**

2335.056

Johnson tortuosity will automatically recalculate from corneocyte dimensions:

$$\tau^* = 1 + \frac{2g}{h} \ln\left(\frac{d}{2s}\right) + \frac{Ndt}{sh} + \left(\frac{d}{1+\omega}\right)^2 \left(\frac{\omega}{hg}\right) (N-1) \quad (7)$$

Simcyp Default of 12.7 has been scaled to reflect this difference based on the ratio calculated by Johnson. Average new value ~ **6**

Viable Epidermis

- In Psoriasis the viable epidermis is thicker on average but has deep rete ridges.
- There is good data available for this: (more than shown here)

Original Article

Cellular Features of Psoriatic Skin: Imaging and Quantification Using In Vivo Reflectance Confocal Microscopy

E. A. W. Wolberink,^{*} P. E. J. van Erp, M. M. Teussink,
P. C. M. van de Kerkhof, and M. J. P. Gerritsen

Department of Dermatology, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands

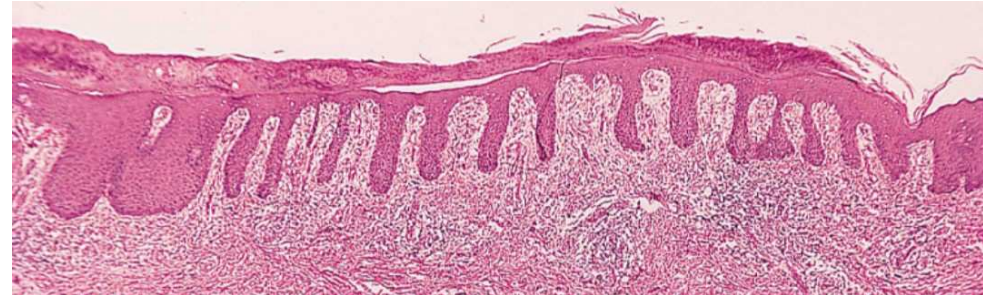


Table 2
Shape Descriptors of Dermal Papillae in Psoriatic and Uninvolved Skin

	Psoriasis		Uninvolved		T-test	P-value
	Mean	SD	Mean	SD		
Area (μm^2)	4186.5	1496.9	3095.8	2082.6	1.3	0.23
Mean gray value	41.7	8.2	50.5	13.2	-2.9*	0.02
Mode gray value	34.7	5.9	40.9	9.5	-3.2*	0.02
Perimeter (μm^2)	247.2	48.7	211.0	70.9	1.3	0.25
Feret (μm^2)	80.5	15.7	70.4	24.7	1.1	0.33
MinFeret (μm^2)	63.9	12.8	51.3	15.3	2.0	0.91
Circularity	0.79	0.02	0.75	0.03	3.7**	<0.01

Circularity, mean and mode gray value of dermal papillae are statistically significant RCM parameters in psoriatic and uninvolved skin. (* $P < 0.01$, ** $P < 0.001$).

Table 1
Quantification and Correlation of Parameters in Psoriatic and Uninvolved Skin

Feature	RCM				Histology			
	Psoriasis		Uninvolved		Psoriasis		Uninvolved	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Dermal Papillae (number/mm ²)	293.8**	132.9	238.2*	90.9	100.4**	35.5	83.8*	14.7
Parakeratosis surface %	45.0**	42.8	0	0	50.0**	41.1	0	0
Height SP (μm)	86.8*	30.8	47.5*	10.6	90.0*	24.7	60.0*	12.7
Height SC (μm)	41.6**	28.2	12.4**	6.7	48.5**	26.3	25.4**	7.2
Capillaries (number/mm ²)	8.8	3.4	1.3	1.5	122.3	57.2	51.3	47.9
Inflammatory cells EF (number/mm ²)	12.6*	5.0	3.1*	2.4	63.9*	43.0	13.2*	31.8
Inflammatory cells T (number/mm ²)					319.6*	226.6	18.1*	10.9
SG surface %					48.8	32.3	100	0
Laser power SG (mW)	3.2	1.5	2.4	2.3				
Refractivity SG	98.0	28.1	80.5	27.4				
Height PD (μm)			38.8	5.7			28.0	11.4
Height EH (μm)			82.4	16.3			79.4	18.8

Dermal papillae number, parakeratosis, amount of inflammatory cells and SP and SC height are significantly correlated to histology. (SP, suprapapillary plate; SC, stratum corneum; EF, *en face* sections; T, transverse sections; SG, stratum granulosum; PD, papillary dermis; EH, epidermal height, * $P < 0.01$, ** $P < 0.001$).

Measurement of epidermal thickness in a patient with psoriasis by computer-supported image analysis

Table 2. Epidermal thickness measurements of psoriasis patients and controls according to biopsy location.

Biopsy location	Full epidermal thickness (μm)	Stratum corneum thickness (μm)	Rete length (μm)	Suprapapillary epidermal thickness (μm)	Dermal papilla distance (μm)
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Patient palmar region (13)	663	192	388	78	35
Control palmar region (15)	207	45	162	43	46

The number of subjects in each group is given in parentheses. The data are reported as medians in micrometers.

Viability Epidermis Thickness

A Simple Method for the Evaluation of Epidermal Thickness Variations in Psoriasis Vulgaris

Psoriasis Area and Severity Index (PASI) given for each patient

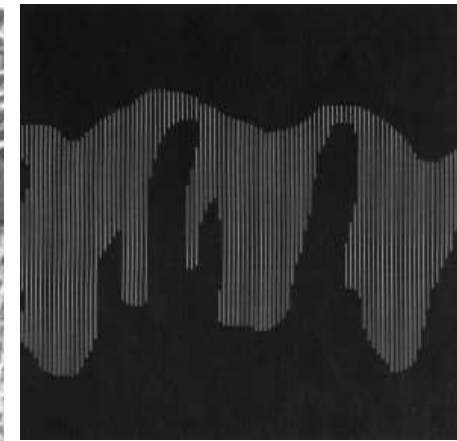
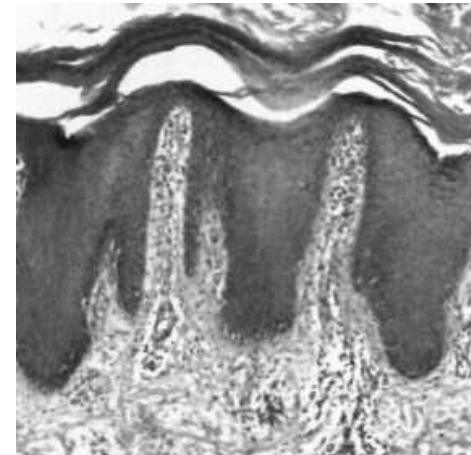


Table 1 Pretreatment Lesional and Nonlesional, and Post-treatment Lesional Biopsy, the Values of Thickness of Epidermis (micron) and,

Case no.	Min			Max			Mean		
	Les	Nonl	PL	Les	Nonl	PL	Les	Nonl	PL
1	29	16	13	264	96	73	165.48	45.18	38.66
2	32	20	18	157	64	53	86.59	37.28	34.12
3	27	16	14	235	92	88	131.47	45.45	43.52
4	19	17	14	171	72	64	103.66	35.58	42.10
5	36	22	18	235	72	70	138.46	35.95	32.24
6	35	25	22	204	99	101	102.75	53.15	50.90
7	24	19	15	294	123	151	144.46	67.26	71.91
8	32	27	10	365	61	75	213.92	41.68	34.73
9	32	24	17	254	81	82	122.23	43.68	52.68
10	42	9	11	239	78	80	147.18	31.97	33.45
11	34	17	11	236	86	83	121.14	43.62	42.38
12	19	18	11	218	48	46	138.37	31.65	31.03
13	12	16	14	292	113	88	189.06	50.52	40.25
14	50	19	13	246	66	58	157.52	35.60	28.36
15	24	19	16	184	82	153	109.41	43.72	72.08
16	15	27	14	216	105	65	109.98	51.44	41.90
17	26	25	18	299	67	69	160.37	50.24	46.12
18	38	16	13	335	78	79	160.60	40.10	38.23
19	26	19	16	322	84	83	162.82	48.51	42.32
20	18	18	17	217	101	93	110.00	46.07	45.15
21	32	16	16	342	107	98	174.71	38.02	39.42
22	21	16	15	325	104	98	147.52	47.16	45.30

Les = pretreatment lesional, Nonl = pretreatment nonlesional, PL = post-treatment lesional, CV = coefficient of variation, Pre = pretreatment, Post = post-treatment, PASI = Psoriasis Area and Severity Index.

PASI Scores

Case no.	SD			CV			PASI	
	Les	Nonl	PL	Les	Nonl	PL	Pre	Post
1	69.23	19.43	15.18	41.84	43.01	39.27	11.9	0.4
2	31.47	10.43	9.32	36.34	27.98	27.32	9.6	0.3
3	58.28	18.83	16.32	44.33	41.43	37.50	11.2	0.7
4	51.58	15.36	13.50	49.76	43.17	32.07	12.3	0.6
5	72.01	13.14	14.05	52.01	36.55	43.58	14.7	0.7
6	60.93	17.94	21.27	59.30	33.75	41.79	8.7	0.1
7	101.94	28.86	34.30	70.57	42.91	47.70	14.2	0.4
8	118.27	8.03	15.50	55.29	19.27	44.63	21.7	0.9
9	78.31	10.86	15.43	64.07	24.86	29.29	8.9	0.4
10	68.61	14.56	15.85	46.62	45.54	47.38	6.9	0.1
11	68.07	18.94	17.36	56.19	43.42	40.96	8.9	0.5
12	59.57	7.44	7.15	43.05	23.51	23.04	14.2	0.6
13	82.93	28.02	16.72	43.86	55.46	41.54	17.1	1.2
14	76.85	10.27	10.05	48.79	28.85	35.44	13.2	1.2
15	54.98	18.38	32.12	50.25	42.04	44.56	21.4	1.7
16	54.42	19.94	12.10	49.48	38.76	28.88	6.9	0.3
17	101.05	9.95	13.99	63.01	19.80	30.33	11.2	0.6
18	100.34	15.48	13.25	62.48	38.60	34.66	12.1	0.2
19	108.87	17.80	10.67	66.87	36.69	25.21	13.1	1.1
20	61.98	18.92	16.90	56.35	41.07	37.43	11.7	0.9
21	106.33	19.11	16.10	60.86	50.26	40.84	12.5	1.4
22	102.73	21.05	18.50	69.64	44.64	40.84	16.3	1.3

Viable Epidermis Thickness

Several studies, various techniques inc. ultrasound, biopsy and confocal microscopy.

Ratio of 2.5-fold to healthy

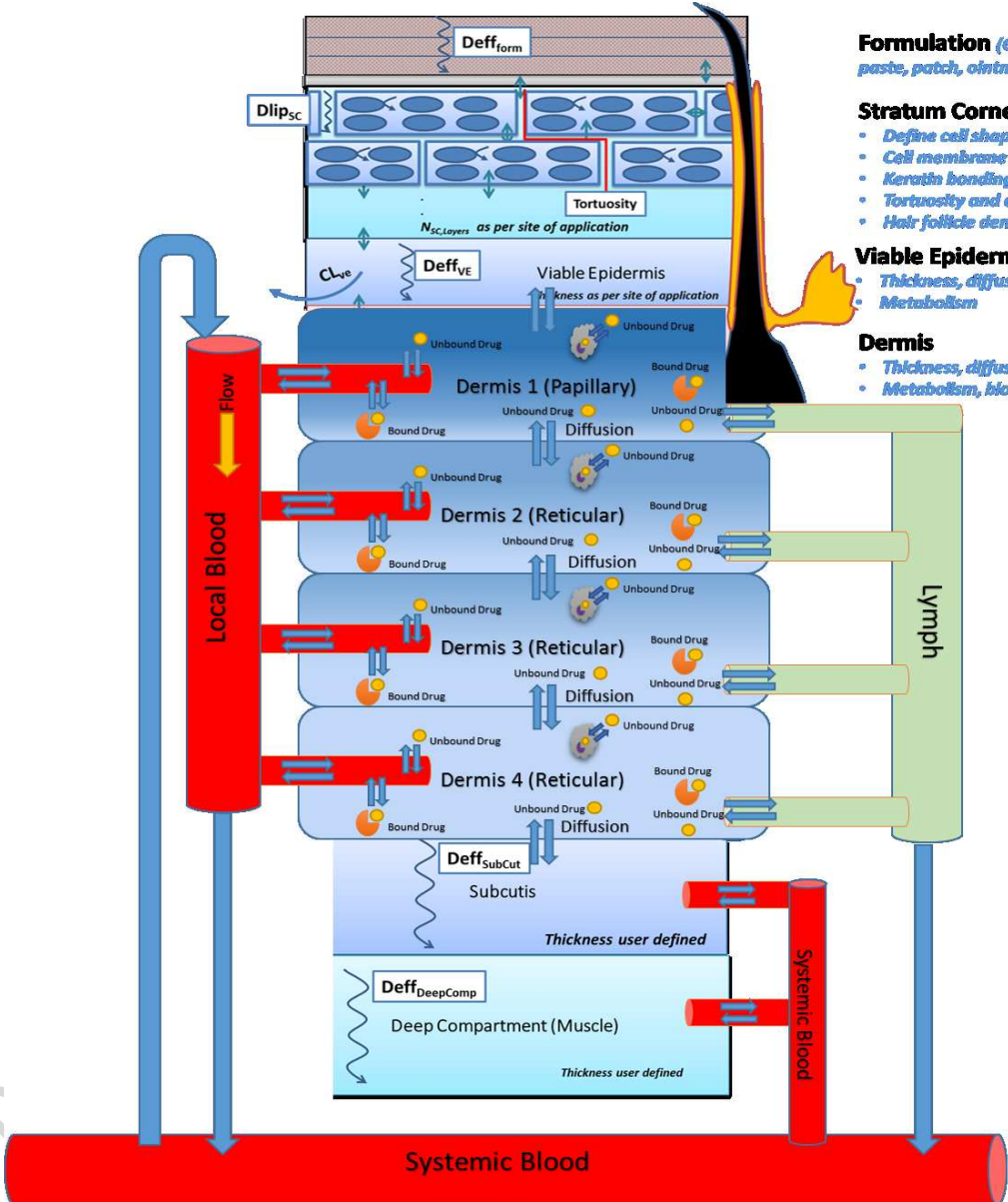
Assumption:

Thickness is half way between the bottom of the supra-papillary plate and the bottom of the rete ridge.

Body site	Healthy	Plaque
Back	45.66	118.17
Fore head	50.05	129.53
Inner Fore arm	41.96	108.59
Outer Fore arm	53.57	138.64
Upper Forearm	51.80	134.06
Face	46.87	121.30
Leg (Lower)	57.36	148.45
Leg (Upper)	40.00	103.52

Depth Resolved Dermis Model (DRDM)

Eleftheria Tsakalozou



Formulation (Gel, cream, lotions, paste, patch, ointments, etc.)

Stratum Corneum (SC)

- Define cell shape and size
- Cell membrane permeability
- Keratin bonding kinetics
- Tortuosity and diffusivity
- Hair follicle density and size

Viable Epidermis (VE)

- Thickness, diffusivity
- Metabolism

Dermis

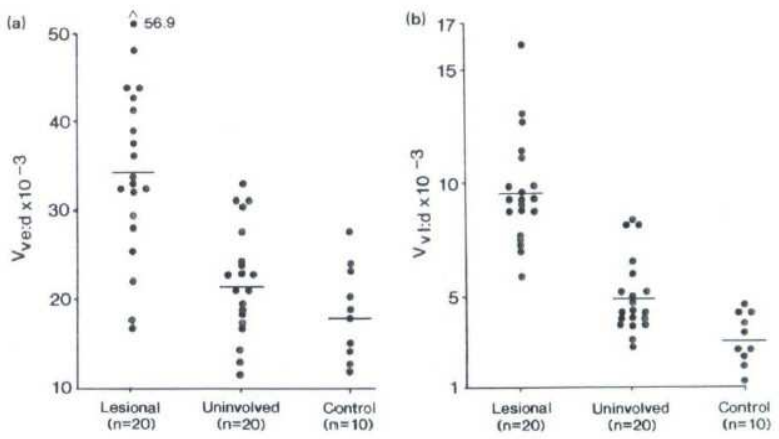
- Thickness, diffusivity
- Metabolism, blood flow

FF

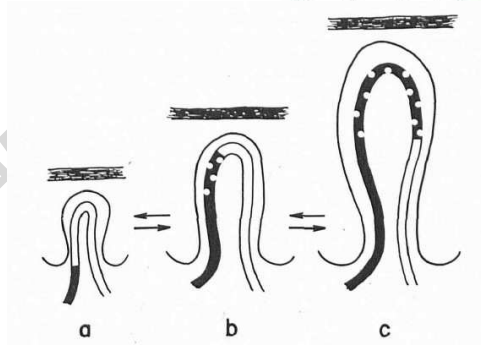
Capillary Volume Fraction

Quantification of microvascular changes in the skin in patients with psoriasis

S.P.BARTON,* M.S.ABDULLAH AND R.MARKS
 Department of Dermatology, University of Wales College of Medicine, Heath Park, Cardiff CF4 4XN, U.K.
 *Boots Pharmaceuticals Division, Nottingham NG2 3AA, U.K.
 Accepted for publication 29 January 1992



Psoriasis patients					Control subjects			
No.	Age	Sex	Body site	Biopsy site	No.	Age	Sex	Biopsy site
101	34	F	L. elbow	Margin	1	34	F	L. elbow
102	40	M	R. thigh	Centre	2	27	F	L. elbow
103	62	F	R. elbow	Margin	3	29	M	L. elbow
104	33	M	L. elbow	Margin	4	30	M	Back
105	29	M	L. elbow	Margin	5	60	F	Abdomen
106	29	M	L. elbow	Margin	6	29	M	Abdomen
107	60	F	Abdomen	Margin	7	65	F	R. elbow
108	30	M	Back	Margin	8	67	M	L. elbow
109	65	F	L. elbow	Margin	9	65	F	L. elbow
110	36	M	Back	Margin	10	10	M	R. thigh
111	76	M	L. elbow	Centre				
112	29	M	Abdomen	Centre				
113	48	M	L. buttock	Margin				
114	37	M	Back	Centre				
115	35	M	L. elbow	Centre				
116	40	M	L. knee	Centre				
117	35	F	L. foot, med. aspect	Centre				
118	41	F	L. elbow	Centre				
119	26	F	L. elbow	Centre				



- Data measuring capillary endothelial and luminal volume fractions in the dermis can be used to parameterise the DRDM model.
- Biopsies taken from various body sites and sexes, however data between groups could not be separated.

Capillary Volume Fraction

In vivo volumetric imaging of microcirculation within human skin under psoriatic conditions using optical microangiography

Imaging of Vessel Density

Cannot be used directly, but agrees with Barton Data (1.5 – 2 x increase between uninvolved and lesion)

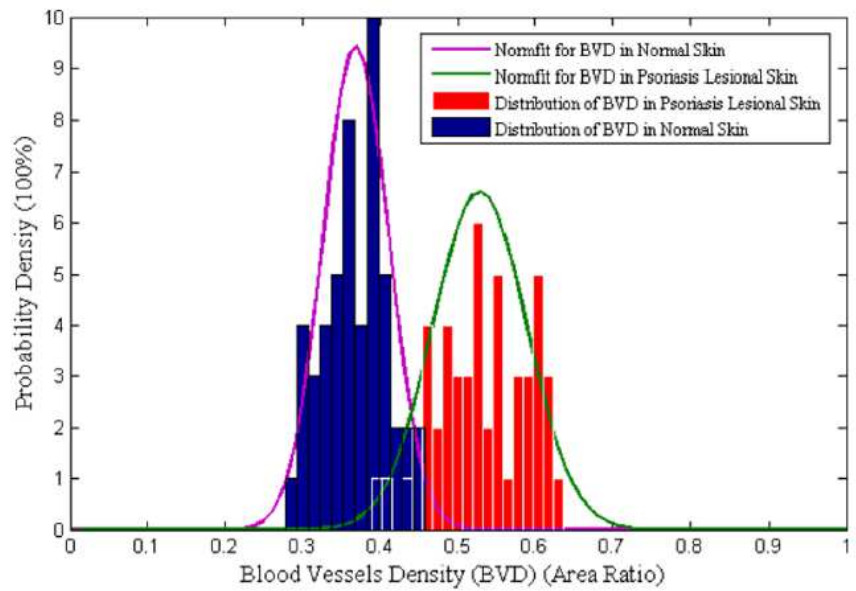
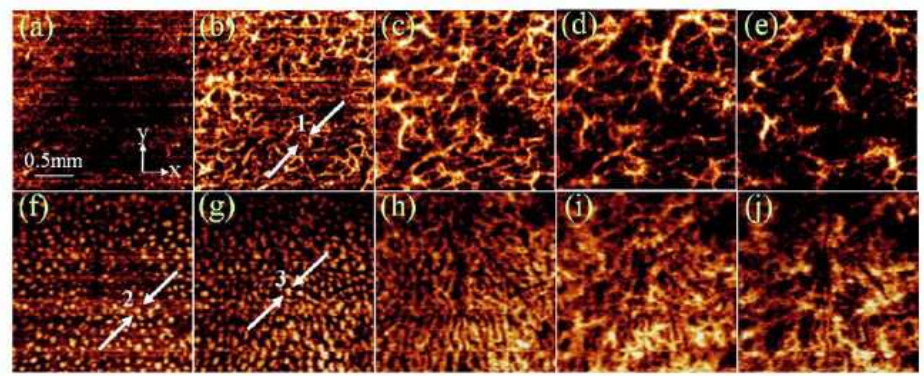


Fig. 7. Histogram distribution for the blood vessel density (BVD) values obtained from the normal skin and psoriatic skin. Blue bars are for the normal skin and the red bars for psoriatic lesion skin. The bell-curves are obtained by fitting the BVD values in each group to the normal distribution function.



Capillary Diameter

Assessment of dermal papillary and microvascular parameters in psoriasis vulgaris using *in vivo* reflectance confocal microscopy

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ULTRASTRUCTURE OF THE CAPILLARY LOOPS IN THE DERMAL PAPILLAE OF PSORIASIS

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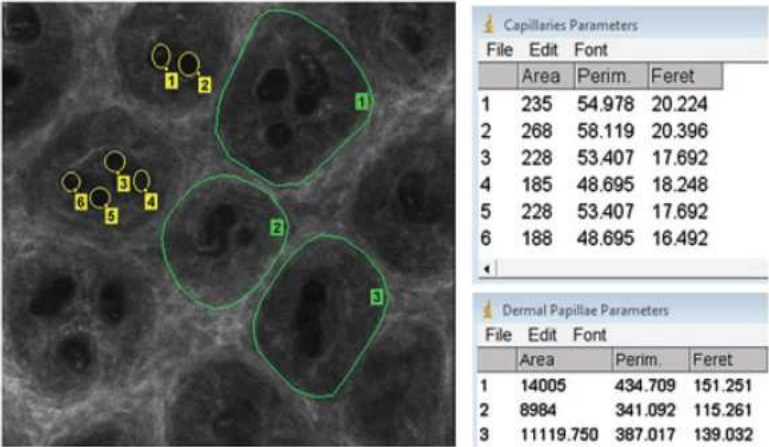


Figure 1. Micromorphological parameters of dermal papillae and dermal capillaries. The parameters of the dermal papillae (green contour lines) and of capillary vessels (yellow circles) were measured using Image J 1.45 analysis software. The parameters investigated were area, Perim and Feret. Perim, perimeter; Feret, Feret's diameter.

TABLE I. Dimensions of intrapapillary capillary loops (outside endothelial tube diameters)

	Ascending limb (μm)	Crest (μm)	Descending limb (μm)
Pustular psoriasis (15 loops)	6-10 ^a	8.8-16 ^b	8.5-17 ^c
Pustular psoriasis after treatment (4 loops)	6-10	8-9	8-10
Psoriasis vulgaris (25 loops)	6-9.6	8-16	7.2-17 ^c
Psoriasis vulgaris after treatment (6 loops)	6-8.4	6-8.4	7.2-12
Normal skin	3.5-6	3.5-6	3.5-6

^a One loop of the 15 had an ascending limb of 14-16 μm .
^b One loop of the 15 had a crest of 18-24 μm .
^c In 2 loops the lower half of the descending limb increased in width to 20-30 μm .

Transvascular Leakage

> Transvascular leakage

Exchange of Macromolecules between Plasma and Skin Interstitium in Extensive Skin Disease

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Departments of Clinical Physiology and Dermatology, The Finsen Institute, Copenhagen, Denmark

TABLE III.

Case No.	¹²⁵ I-TER _{alb} ^a (% IVM·hr ⁻¹)
1	9.1
2	7.6
3	6.4
4	6.8
5	7.0
6	7.3
7	7.5
8	7.3
9	11.4
10	8.8
11	8.6
Patients Mean	8.0
± SD	1.4
Controls Mean	5.1
± SD	1.2
p <	0.001

^a TER = transcapillary escape rate; alb = a ¹²⁵I-albumin and ¹²⁵I-IgG.

Microvascular Protein Leakage in Extensive Skin Diseases: Aspects of the Transport Mechanisms

ANNE-MARIE WORM, M.D., AND NIELS ROSSING, M.D.

Departments of Clinical Physiology and Dermatology, The Finsen Institute, Copenhagen, Denmark

Transvascular transport and distribution of fluid and protein in psoriasis

Bent Staberg, M.D., Anne-Marie Worm, M.D., Per Klemp, M.D., and Niels Rossing, M.D.
Copenhagen, Denmark

PV/IVF	TER (% IVM × hr ⁻¹)	A _b /A _s (% × hr ⁻¹)	
		I	II
0.33	8.3	9.9	14.1
0.31	7.6	5.1	29.9
0.32	7.5	1.6	16.1
0.33	5.8	3.3	6.2
0.40	3.5	8.9	11.9
0.29	6.8	7.3	27.6
0.23	8.6	4.7	9.9
0.29	6.4	4.4	11.5
—	—	—	—
—	—	—	—
0.31	6.8	5.7	15.9
0.05	1.6	2.8	8.5
			<0.02
0.39	5.1	8.3	
0.06	1.2	2.6	
<0.01	<0.05	NS	<0.02

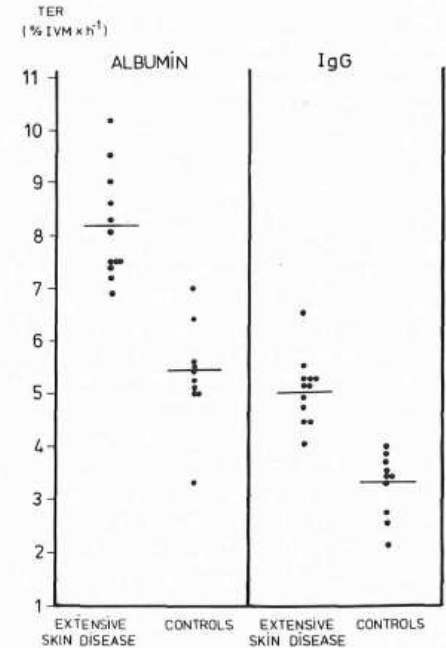


Fig 1. Transcapillary escape rate of albumin and IgG in patients with extensive skin disease and controls. The solid line represents the mean value.

Elefth

FDA - 09/17

Interstitial Fluid

> ISF Volume

Plasma and Interstitial Fluid Volume in Extensive Skin Disease

ANNE-MARIE WORM, M.D., AND HANS-HENRIK PARVING, M.D.

The Departments of Clinical Physiology and Dermatology, The Finsen Institute and the Department of Clinical Physiology, Bispebjerg Hospital, Copenhagen, Denmark

The mean values of plasma volume (PV), extracellular fluid volume (ECV), interstitial fluid volume (IFV), and PV/IFV ratio in 17 patients with extensive skin disease and 18 normal subjects

		PV				ECV				IFV				PV
		ml	ml/kg	ml/cm	l/m ²	ml	ml/kg	ml/cm	l/m ²	ml	ml/kg	ml/cm	l/m ²	IFV
Patients	Mean	2916	37.5	17.1	1.54	11407	146	66.8	6.01	8439	109	49.8	4.47	0.35
	± SD	487	6.5	2.3	0.17	8630	22	9.4	0.55	1456	17	7.8	0.48	0.05
Controls	Mean	3110	45.0	18.0	1.69	10666	155	61.7	5.82	7556	110	43.8	4.13	0.42
	± SD	525	4.7	2.5	0.18	1823	21	9.2	0.75	1371	18	7.2	0.62	0.05
p <		NS	0.001	NS	0.05	NS	NS	NS	NS	NS	NS	NS	NS	0.001

The volumes are given in absolute figures (ml) and related to body weight (ml/kg), body height (ml/cm) and body surface area (l/m²).

Transvascular transport and distribution of fluid and protein in psoriasis

Bent Staberg, M.D., Anne-Marie Worm, M.D., Per Klemp, M.D., and Niels Rossing, M.D.
Copenhagen, Denmark

Table I. Clinical data, plasma volume (PV), extracellular fluid volume (ECV), interstitial fluid volume (IFV), transcapillary escape rate of albumin (TER), appearance rate of labeled albumin in blister fluid (A_b/A_s) from uninvolved (I) and involved (II) skin in ten patients with psoriasis

Case No.	Surface area (m ²)	Skin involvement (%)	PV (liter/m ²)	ECV (liter/m ²)	IFV (liter/m ²)
1	2.18	80	1.93	7.71	5.78
2	2.08	70	1.85	7.77	5.92
3	1.88	40	1.35	5.57	4.22
4	2.01	30	1.35	5.43	4.08
5	1.93	40	1.87	6.56	4.69
6	1.88	40	1.34	5.96	4.62
7	1.83	60	1.09	5.85	4.76
8	1.73	50	1.47	6.54	5.07
9	2.15	40	—	—	—
10	1.95	60	—	—	—
<i>Patients</i>					
Mean	1.96	51	1.53	6.42	4.89
±S.D.	0.14	16	0.31	0.91	0.67
p					
<i>Controls*</i>					
Mean	1.80		1.64	5.91	4.27
±S.D.	0.19		0.15	0.75	0.67
p	NS		NS	NS	NS

* Values from Rossing N, Worm A-M: Clin Physiol 1:275, 1981.

> Lymph Flow

Lymphatic albumin clearance from psoriatic skin

Bent Staberg, M.D., Per Klemp, M.D., Michael Aasted, M.D., Anne-Marie Worm, and Per Lund, M.D.
Copenhagen, Denmark

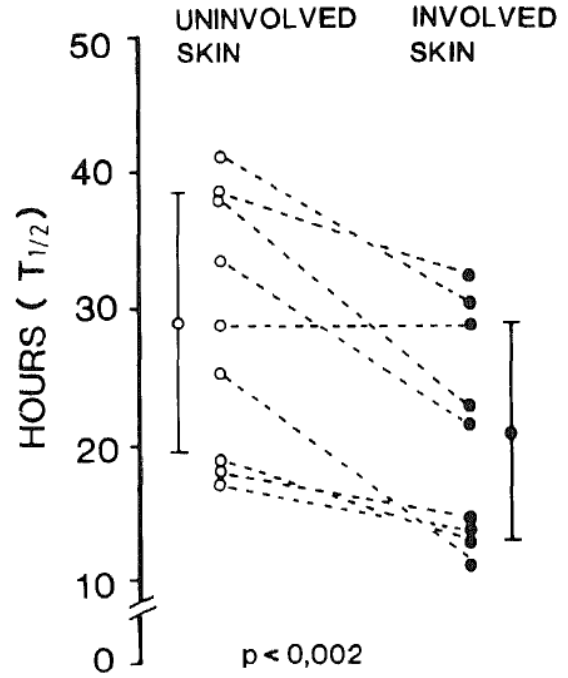


Fig. 2. Radiolabelled albumin clearance ($T_{1/2}$) in uninvolved (○) and involved (●) skin of nine patients with psoriasis. Vertical bars represent means \pm S.D.

> Lymph vessel density

Increased vessel density in psoriasis: involvement of lymphatic vessels in the papillary dermis

Table 2 Density of lymphatic vessels in skin of psoriatic plaque and normal skin control

	Psoriasis, n = 32 (n mm ⁻¹) ^a	Control, n = 32 (n mm ⁻¹) ^a	P-value ^b
Lymphatics (all sizes) ^c	10.01 \pm 6.09	3.58 \pm 1.31	< 0.001
Inside dermal papillae ^d	3.36 \pm 3.62	0.01 \pm 0.03	< 0.001
Lymphatics < 10 μ m ^c	3.58 \pm 3.52	1.47 \pm 0.94	0.002
Inside dermal papillae ^d	1.25 \pm 2.02	0.01 \pm 0.02	< 0.001
Lymphatics 10–17 μ m ^c	4.03 \pm 2.68	1.47 \pm 0.89	< 0.001
Inside dermal papillae ^d	1.48 \pm 1.60	0.00 \pm 0.03	< 0.001
Lymphatics 18–24 μ m ^c	1.58 \pm 1.49	0.33 \pm 0.25	< 0.001
Inside dermal papillae ^d	0.48 \pm 0.59	0.00 \pm 0.00	< 0.001
Lymphatics > 24 μ m ^c	0.82 \pm 0.67	0.32 \pm 0.51	< 0.001
Inside dermal papillae ^d	0.15 \pm 0.28	0.00 \pm 0.00	< 0.001

^aMean \pm SD. ^bAsymptotic significance level (two-tailed) Mann-Whitney U-test. ^cAll stained vessel segments above the upper horizontal blood plexus. ^dPositive segments between elongated rete ridges.

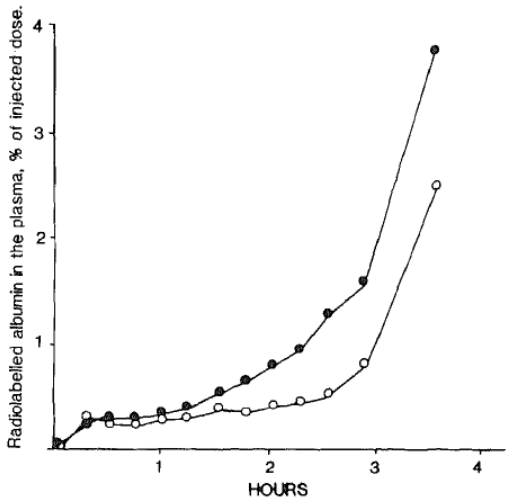
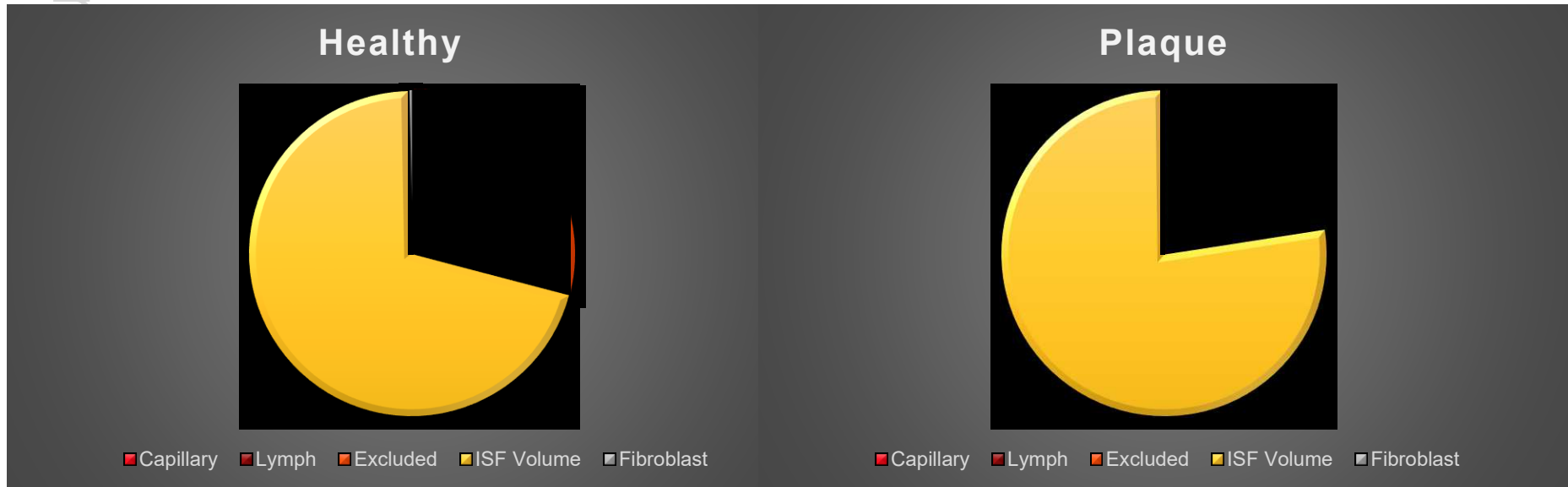


Fig. 3. An example of the plasma appearance rate during the resting period of radiolabelled albumin injected intradermally in uninvolved (○) and involved (●) skin of a patient with psoriasis. About 3 hours after the injection the patient was allowed to move.

Volume Fractions

Volume fraction of Dermis Components for DRDM



Physiology

Dermis

Total Thickness (µm)

Regional dermis blood flow scalar*

DRDM Parameters

	Vessel Radii (µm)				Volume Fraction (cm ³ /cm ³) †						
	Layer Thickness (%)*	Capillary Wall Thickness (µm)	Capillary Lumen	Capillary External	Capillary †		Lymph †		Fibroblast	Excluded	Interstitial Fluid
					Mean	CV (%)	Mean	CV (%)	Mean	Mean	
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Dermis 2	<input type="text" value="29.25"/>	<input type="text" value="4.4"/>	<input type="text" value="7.66"/>	<input type="text" value="12.06"/>	<input type="text" value="0.014"/>	<input type="text" value="37.54"/>	<input type="text" value="0.0341"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.24298"/>	<input type="text" value="0.706"/>
Dermis 3	<input type="text" value="29.25"/>	<input type="text" value="4.4"/>	<input type="text" value="7.66"/>	<input type="text" value="12.06"/>	<input type="text" value="0.014"/>	<input type="text" value="37.54"/>	<input type="text" value="0.0341"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.27298"/>	<input type="text" value="0.706"/>
Dermis 4	<input type="text" value="29.25"/>	<input type="text" value="4.4"/>	<input type="text" value="7.66"/>	<input type="text" value="12.06"/>	<input type="text" value="0.014"/>	<input type="text" value="37.54"/>	<input type="text" value="0.0341"/>	<input type="text" value="98.98"/>	<input type="text" value="0.003"/>	<input type="text" value="0.24298"/>	<input type="text" value="0.706"/>

Lymph Flow Scalar

Fraction of ISF accessible by albumin

Albumin concentration in accessible spce (g/L)

* % of the total dermis thickness. † Capillary and lymph volume fractions are that occupied by lumen + endothelium. ‡ Volume of substituent per cm³ of tissue



FDA - 091

Dermis Blood Flow

- Many studies available with various techniques employed
- Consistent finding of $>$ blood flow, ranging from 1.7 – 13.5 fold.

Assumption:

Because all studies found minimal difference between uninvolved and healthy BF (<1.5 fold).

Uninvolved was assumed “healthy” to calculate ratios where healthy control was not used

Study	Method	Final Ratio to 'healthy'
NYFORS 1970	Xe washout	1.70
Klemp 1983	Xe washout	10.16
Klemp 1985	Xe washout	4.10
Klemp 1985	Xe washout	13.50
Ferguson 1961	I Washout	2.45
Yosopivitch 2003	LDV	2.88
Goh 2004	LDPI	2.78
Hern 2005	LDPI	9.08
Khan 1987	LDV	6.69
Ascheim 1966	Na washout	1.79
	Mean Ratio	5.52

Subcutis Blood Flow

Two studies by Klemp *et al*, Xe washout draw a similar conclusion.

Study	Method	Final Ratio to healthy
Klemp 1985a	Xe washout	1.571
Klemp 1985b	Xe washout	1.776
	Mean	1.67

Eleftheria Tsakalozou

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Albumin

Several studies suggest a decreased albumin content in both serum and ISF.

	N	Serum	ISF
Staberg	10	0.889	0.754
Worm	2	0.841	0.850
Worm	6	0.841	0.681
Sheikh	100	0.918	
Weighted Mean Ratio to healthy		0.910	0.740

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FDA - 09/12/2019 21

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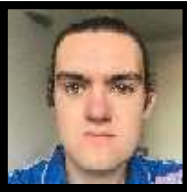
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Questions?



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