



Biopharmaceutic In Vitro In Vivo Extrapolation (IVIV_E) Informed Physiologically-Based Pharmacokinetic Model of Ritonavir Norvir Tablet Absorption in Humans Under Fasted and Fed State Conditions

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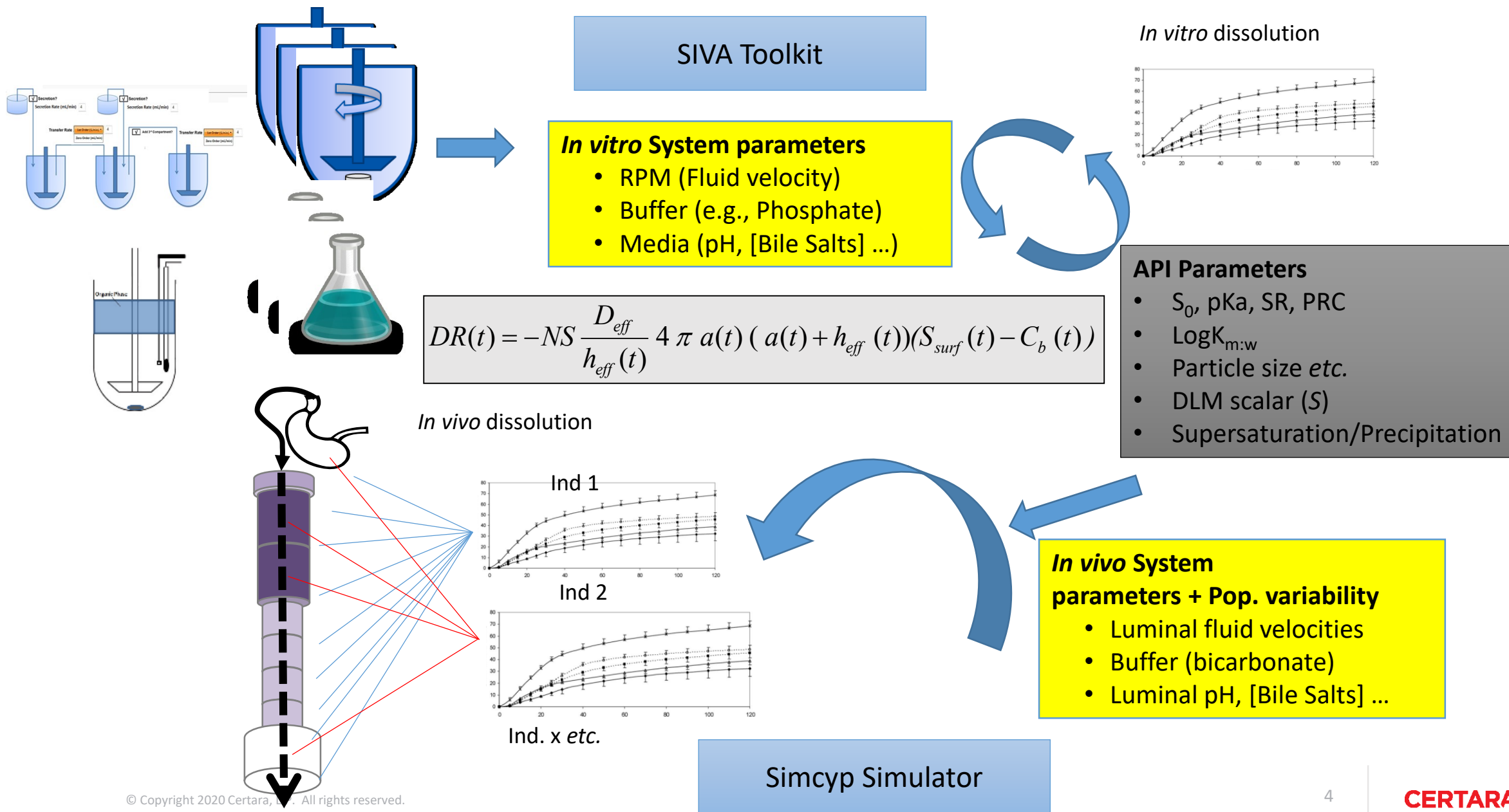
Simcyp Consortium Members Webinar
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Outline of the Presentation

- Aim, Background and Challenges
- Model (dis)qualification in a stepwise manner
- Results and Discussion
- Conclusions and Limitations

The objective of this research is to develop a mechanistic absorption and disposition model of ritonavir which could link the biopharmaceutic properties of the drug/drug product with its DDI risk potential

Running Theme – Biopharmaceutics *In vitro In vivo* Extrapolation (IVIVE)



Biopharmaceutics IVIVE – Many Case Studies

molecular
pharmaceutics

Article

In Silico Modeling Approach for the Evaluation of Gastrointestinal Dissolution, Supersaturation and Precipitation of Posaconazole

Bart Hens, Shriram M. Pathak, Amitava Mitra, Nikunj Kumar Patel, Bo Liu, Sanjay Kumar Patel, Masoud

molecular
pharmaceutics

Article

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Model-Based Analysis of Biopharmaceutic Experiments To Improve Mechanistic Oral Absorption Modeling: An Integrated *in Vitro in Vivo* Extrapolation Perspective Using Ketoconazole as a Model Drug

Shriram M. Pathak,^{*,†,⊕} Aaron Ruff,[‡] Edmund S. Kostewicz,[‡] Nikunj Kumar Patel,[†] David B. Turner,[†] and Masoud Jamei[†]



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Journal of Pharmaceutical Sciences

journal homepage: www.jpharmsci.org

Pharmacokinetics, Pharmacodynamics and Drug Transport and Metabolism

Biopharmaceutic *IVIVE*—Mechanistic Modeling of Single- and Two-Phase *In Vitro* Experiments to Obtain Drug-Specific

Parameters for Incorporation

Shriram M. Pathak^{1, *}, Kerstin J

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Article

Biopharmaceutic *In Vitro In Vivo* Extrapolation (IVIV_E) Informed Physiologically-Based Pharmacokinetic Model of Ritonavir Norvir Tablet Absorption in Humans Under Fasted and Fed State Conditions

Sumit Arora,^{*} Amita Pansari, Peter Kilford, Masoud Jamei, Iain Gardner, and David B. Turner

Challenges with the PBPK model development with Ritonavir

- Complex pharmacokinetic profile – mechanism-based inhibitor of CYP3A4/5, inducer of CYP3A4/5, P-gp substrate and also an inhibitor
- Lack of intravenous data (parameter identifiability issues) and information regarding fraction metabolized by CYP3A4
- Commercial formulation is an amorphous solid dispersion (ASD). Dissolution of ASDs under non-sink conditions is complex with the possibility of several phase transitions within the dissolving medium and the ASD matrix
- Being a weak base, parameterization of models which adequately capture interplay between supersaturation and precipitation of ritonavir in gut lumen can be complex

We adopted a Model (dis)qualification approach in a stepwise manner until consistency with clinical data is obtained

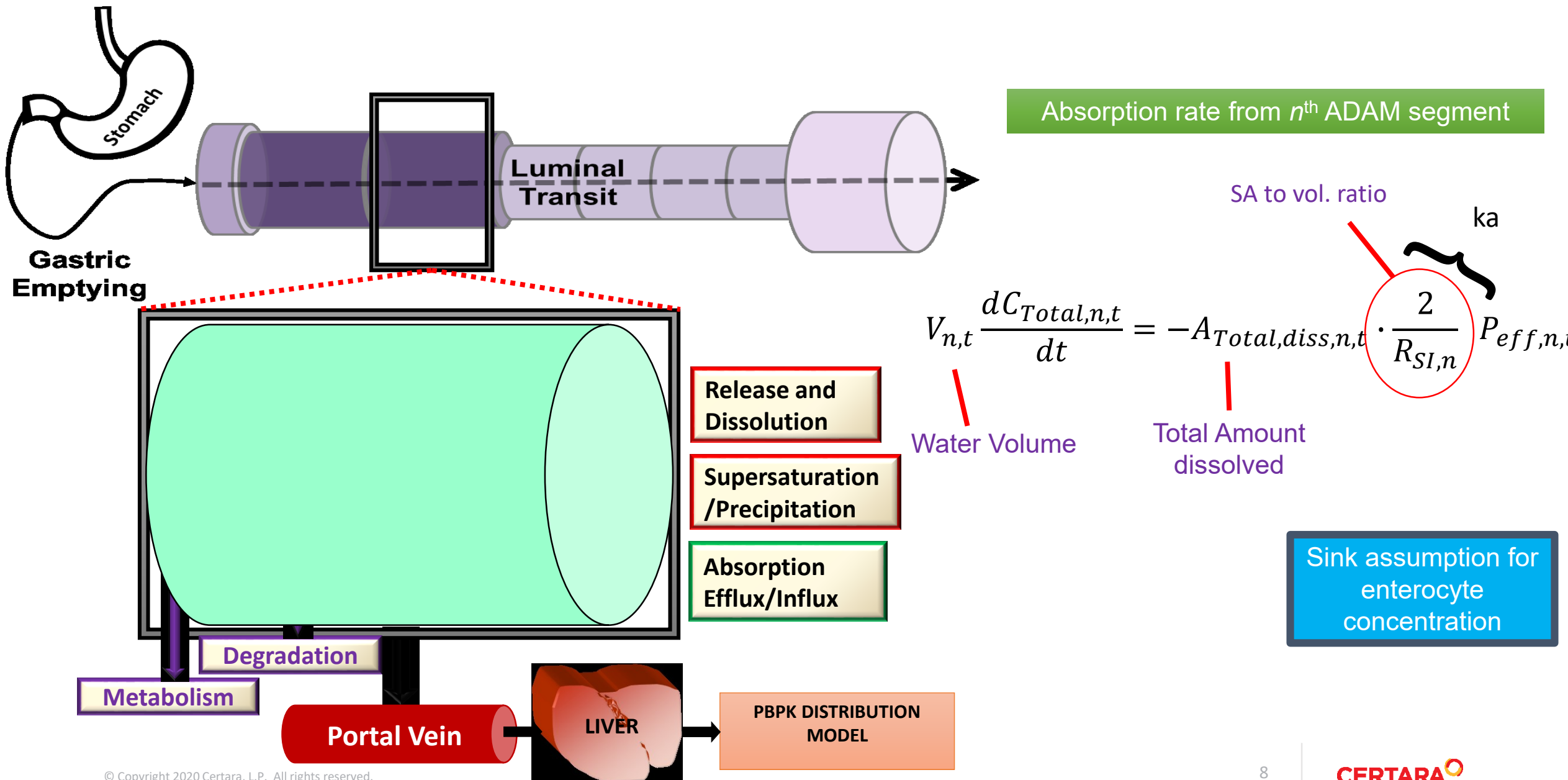
Can we simulate ritonavir in amorphous state?

- Norvir[®] 100 mg tablet - amorphous solid dispersion of ritonavir with copovidone (PVP –VA) (15% drug load)
- T_g of ritonavir 50°C^a – indicative of inherent stability of amorphous ritonavir at room temperature
- Hancock^b classified ritonavir as molecule with low propensity for crystallization (MW – 721 Da, 18 rotatable bonds)
- Indulkar et al^c – Nucleation induction time (time taken to reduce initial supersaturation concentration by 2.5%) of ritonavir – 700 min in composite SIF and around 300 min in FaSSIF-V1

Hence, it is quite likely that ritonavir remains in amorphous state during its transit in gut

^a Zhou D et al, J Pharm Sci, 2007;96(1):71-83, ^b Hancock J Pharm Sci 2017, 106(1):28-30, ^c Indulkar et al Pharm Res (2018) 35:158

Advanced Dissolution Absorption and Metabolism(ADAM) Model



Two Models for Handling Dissolution of Particles

Permeability Formulation

Formulation Type Transit Times Diffusion Layer Model Luminal Degradation

Model Two Solid States Fraction in Dose (%) Solid State 1: 70 Solid State 2: 30

General Aqueous Phase Solubility Bile Micelle Mediated Solubility Supersaturation and Precipitation Excipient-Mediated Solubility

Diffusion Layer Model (DLM) Equation

$$DR(t) = -NS \frac{D_{eff}(t)}{h_{eff}(t)} 4\pi a(t)(a(t) + h_{eff}(t))(S_{surface}(t) - C_{bulk}(t))$$

The equation is applied to each particle size bin and the dissolution rate summed over all bins.

DR(t)	Rate of dissolution at time t	h _{eff} (t)	Effective diffusion layer thickness at time t
N	Number of particles	a(t)	Radius of particle at time t
S	Empirical scalar (DLM Scalar)	S _{surface} (t)	Solubility at particle surface time t
D _{eff} (t)	Effective diffusion coefficient at time t	C _{bulk} (t)	Concentration of drug in bulk solution at time t

DLM Model Options

Mass Balance Only Model Particle Population Balance Model

Include bile salt micelle-mediated component

Include excipient-bound component

MBO
v17 and before
Retained in v18 for
backward compatibility

PPB
From v18
More Mechanistic
More features to consider

Particle Population Balance (PPB) Model

- The main idea is to track the number of particles per bin.

Particles

$$\frac{dN_{i,j}(t)}{dt} = - \frac{[G_i(a_{j+1}, t)N_{i,j+1} - G_i(a_j, t)N_{i,j}]}{\Delta_j} + M$$

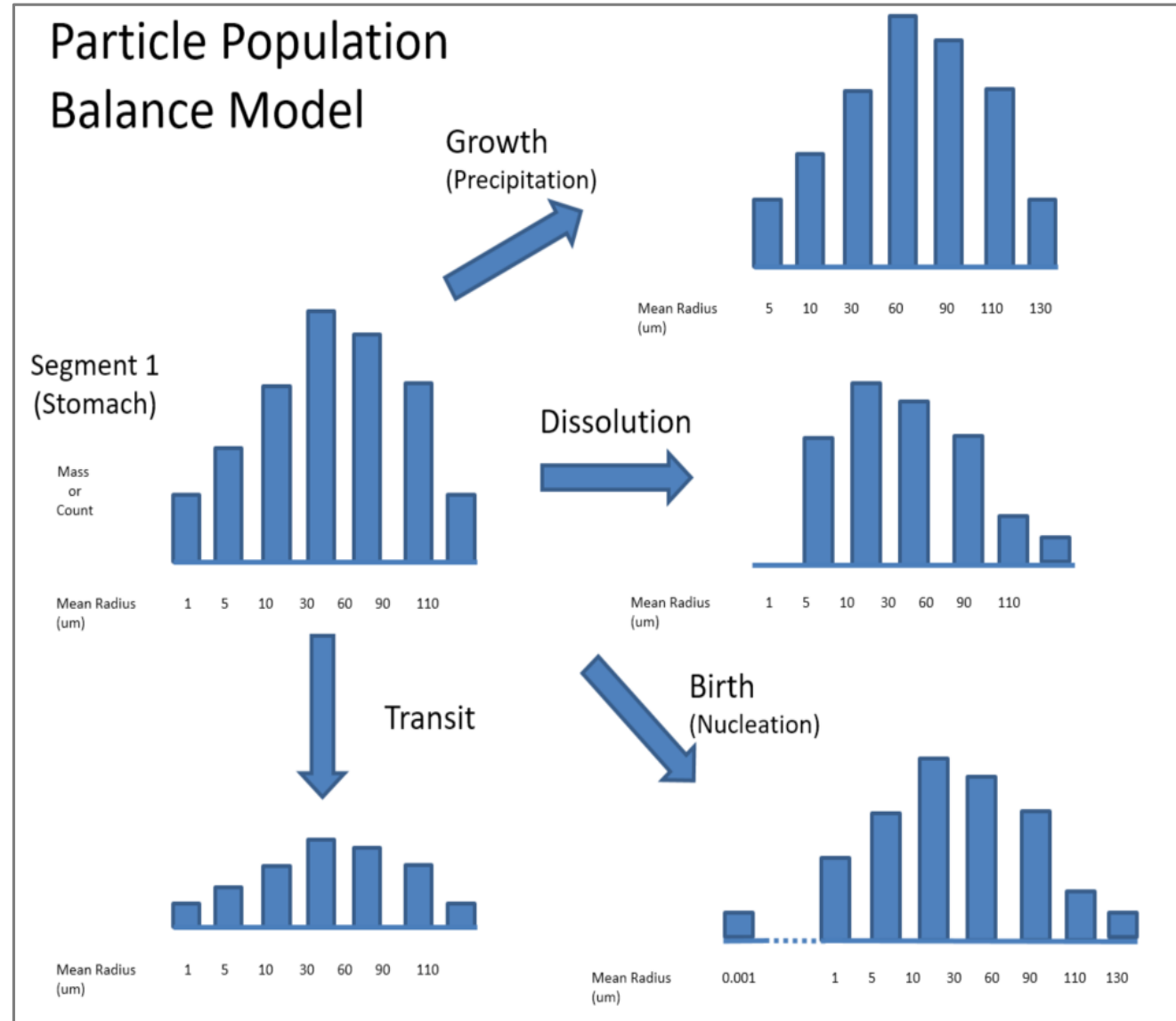
$$M = \begin{cases} -K_{t,1}N_{1,j} & i = 1 \\ K_{t_{i-1}}N_{i-1,j} - K_{t_i}N_{i,j} & i = 2 \text{ to } 8 \end{cases}$$

$$\Delta_j = a_{j+1} - a_j$$

$$N_j = n_j \Delta_j$$

$$G(a, t) = \frac{da}{dt} = - \frac{D_{eff}}{\rho h} \frac{(a+h)(C_s - C_i)}{a}$$

Different dissolution/growth function?

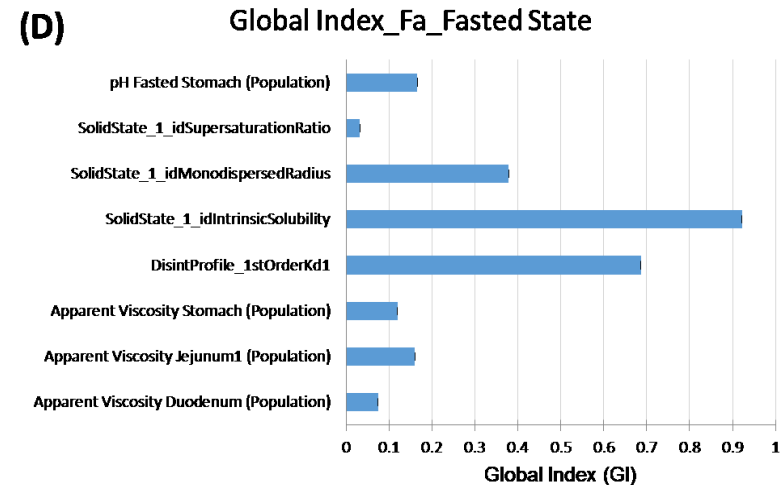
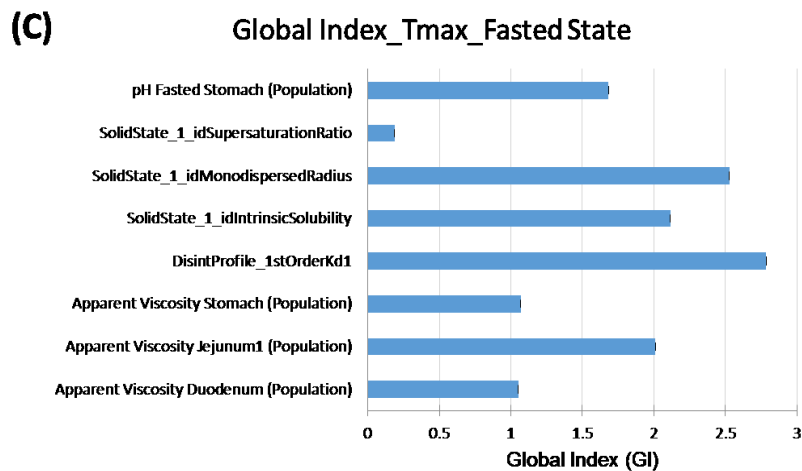
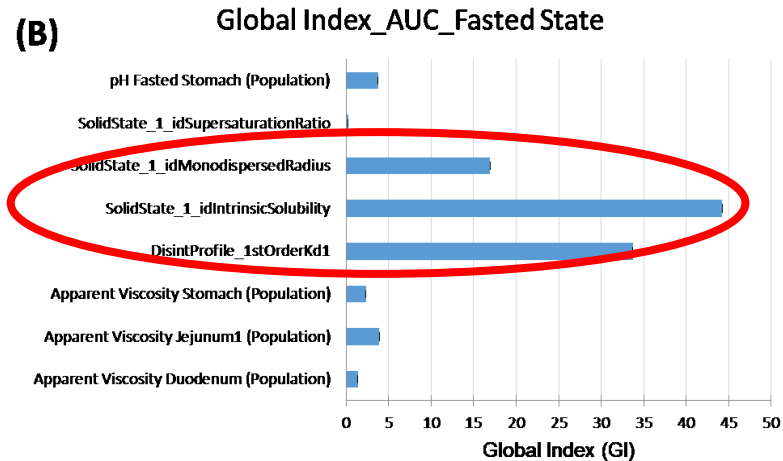
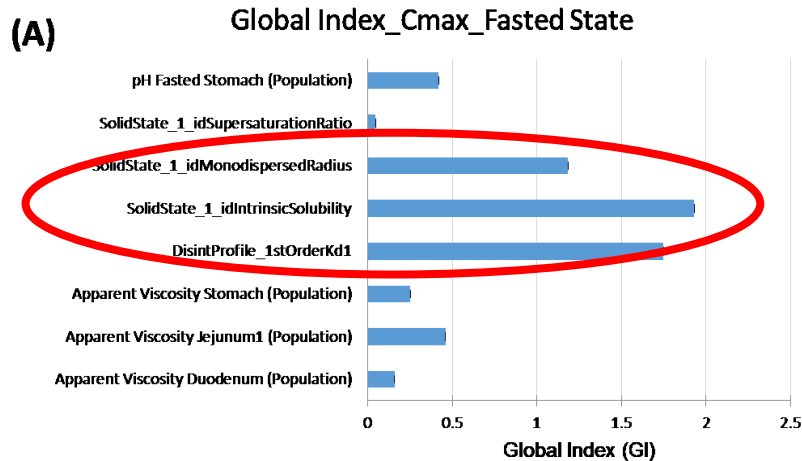


Main Functionalities - MBO vs PPB

	Mass Balance Only (MBO)	Particle Population Balance (PPB)
Mass balance	Y	Y
Particle count	Fixed	Particle “death” and “birth”
Particle bin handling (Polydispersed)	Discrete with gaps	Discrete with or without gaps
Maximum number of PSD bins	10 (V17), 1000 (V18)	1,000
Two PSDs	N	Y
Two solid states	N	Y (in dose and precipitating)
Excipient-mediated Solubility	Y	Y
pH-dependent & Bile Micelle Solubility	Y	Y
Particle Surface Solubility	Y	Y
Immediate/Modified/Control/Extended Release Formulations	Y	Y
Food Staggering Model	N	Y
Custom Dosing with different doses	N	Y
Segregated Transit Time model	Y	Y
First Order Precipitation	Y	Y
Mechanistic Precipitation	N	Y
Nucleation	N	Y
Luminal degradation	Y	Y

Global Sensitivity Analysis - Introduced in Simcyp V19

- The Morris method was employed which is appropriate for initial screening of parameters influencing model outcomes



GSA Analysis of chosen parameters on (A) C_{max} (B) AUC (C) T_{max} and (D) Fraction Absorbed (Fa) of ritonavir systemic exposure following administration of 100 mg Norvir® Tablet in the fasted state.

Sequential Modeling of *In Vitro* Biopharmaceutical Experiments

SIVA 3

Aqueous Solubility Modelling



$$S_{(BS)Tot} = \left([BS] \cdot \frac{S_0}{C_{H_2O}} \cdot K_{m:w,unionised} + S_0 \right) + \left([BS] \cdot \frac{S_i}{C_{H_2O}} \cdot K_{m:w,ionised} + S_i \right)$$

Biorelevant Solubility Modelling

Confirmed Intrinsic Solubility & Solubility Factors



$$S_{(BS)Tot} = \left([BS] \cdot \frac{S_0}{C_{H_2O}} \cdot K_{m:w,unionised} + S_0 \right) + \left([BS] \cdot \frac{S_i}{C_{H_2O}} \cdot K_{m:w,ionised} + S_i \right)$$

USP-2 Dissolution Modelling

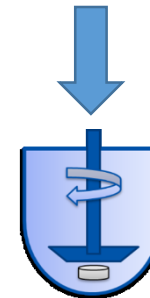
Confirmed Bile Micelle Partition Coefficients



$$DR(t) = -NS \frac{D_{eff}}{h_{eff}(t)} 4\pi a(t) (a(t) + h_{eff}(t)) (S_{surface}(t) - C_{bulk}(t))$$

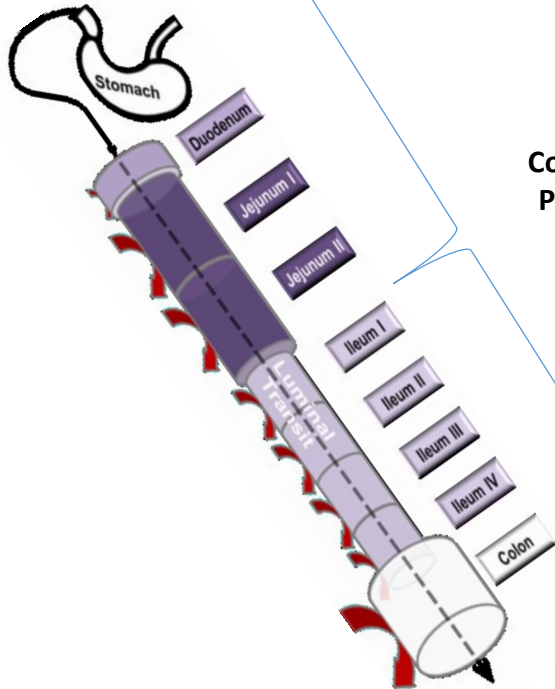
pH Shift Experiment Modelling

Confirmed Delayed Release and DLM Parameters



Critical Supersaturation Ratio ^c	1
Precipitation Rate Const. (1/h) ^d	0.001

Confirmed Precipitation Parameters



Confirm and/or Estimate Parameters

Sequential Modeling of *In Vitro* Biopharmaceutic Experiments

Aqueous Solubility Modelling



$$S_{(BS)Tot} = \left([BS] \cdot \frac{S_0}{C_{H_2O}} \cdot K_{m:w,unionised} + S_0 \right) + \left([BS] \cdot \frac{S_i}{C_{H_2O}} \cdot K_{m:w,ionised} + S_i \right)$$

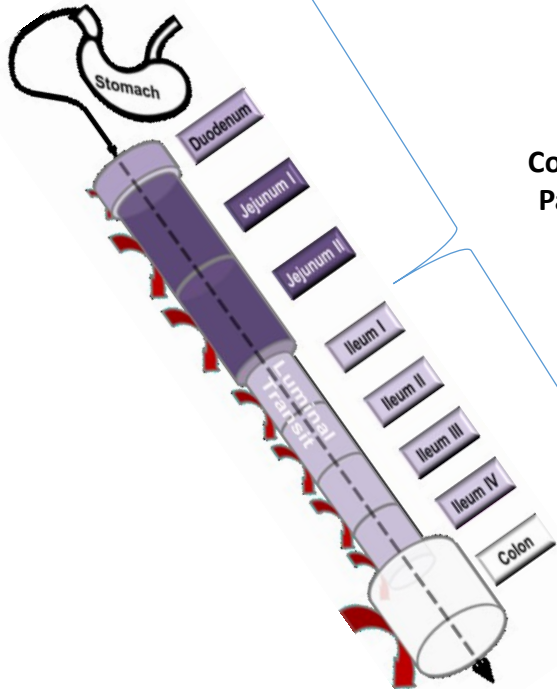
Biorelevant Solubility Modelling



$$S_{(BS)Tot} = \left([BS] \cdot \frac{S_0}{C_{H_2O}} \cdot K_{m:w,unionised} + S_0 \right) + \left([BS] \cdot \frac{S_i}{C_{H_2O}} \cdot K_{m:w,ionised} + S_i \right)$$

Confirmed Intrinsic Solubility & Solubility Factors

Confirmed Bile Micelle Partition Coefficients



Modeling of *In Vitro* Solubility Experiments

Global Solubility Equation v19

Aqueous Phase Solubility

$$S_{Tot}(t) = S_o \cdot S_{o_scalar}(t) \cdot \left(1 + \frac{[BS](t)}{C_{H_2O}} \cdot Km:w,u\right) + S_i(t) \cdot \left(1 + \frac{[BS](t)}{C_{H_2O}} \cdot Km:w,i\right) + S_{bound,excip}(t)$$

Bile Micelle Phase Solubility

Excipient "Phase" Solubility

Apply bile micelle mediated solubilisation enhancement?

Inhibitor 1

Excipient

Reduces to: $S_{Tot}(t) = S_o \cdot S_{o_scalar}(t) + S_i(t)$ Aqueous Phase Solubility

where

Monoprotic Base $S_i(t) = S_o \cdot S_{o_scalar}(t) \cdot (10^{pKa-pH(t)})$

Diprotic Base $S_i(t) = S_o \cdot S_{o_scalar}(t) \cdot (10^{pKa1-pH(t)} + 10^{pKa2-pH(t)} + 10^{pKa1+pKa2-2pH(t)})$

Monoprotic Acid $S_i(t) = S_o \cdot S_{o_scalar}(t) \cdot (10^{pH(t)-pKa})$

Diprotic Acid $S_i(t) = S_o \cdot S_{o_scalar}(t) \cdot (10^{pH(t)-pKa1} + 10^{pH(t)-pKa2} + 10^{2pH(t)-pKa1-pKa2})$

Ampholytes $S_i(t) = S_o \cdot S_{o_scalar}(t) \cdot (10^{pH(t)-pKa1} + 10^{pKa2-pH(t)} + 10^{pKa2-pKa1})$

where, for Ampholytes only, pKa1 is acidic and pKa2 is basic

Modeling of *In Vitro* Solubility Experiments

Aqueous Solubility Modelling

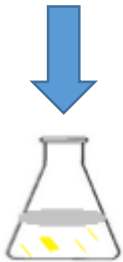


Confirmed Intrinsic solubility, pKa & Solubility Factors

$$S_{Tot}(t) = S_o \cdot S_{o_{scalar}}(t) \cdot \left(1 + \frac{[BS](t)}{C_{H_2O}} \cdot Km:w, u\right) + S_i(t) \cdot \left(1 + \frac{[BS](t)}{C_{H_2O}} \cdot Km:w, i\right) + S_{bound, excip}(t)$$

Free base amorphous solubility, pH 7.4 = 38.5 µg/mL (Xu et al 2017)
 Solubility in 0.1 N HCL = 4 mg/mL (Law 2001 JPharmSci 90:1015)
 Solubility Factor = 103.896 (Optimized with SIVA 3)

Biorelevant Solubility Modeling

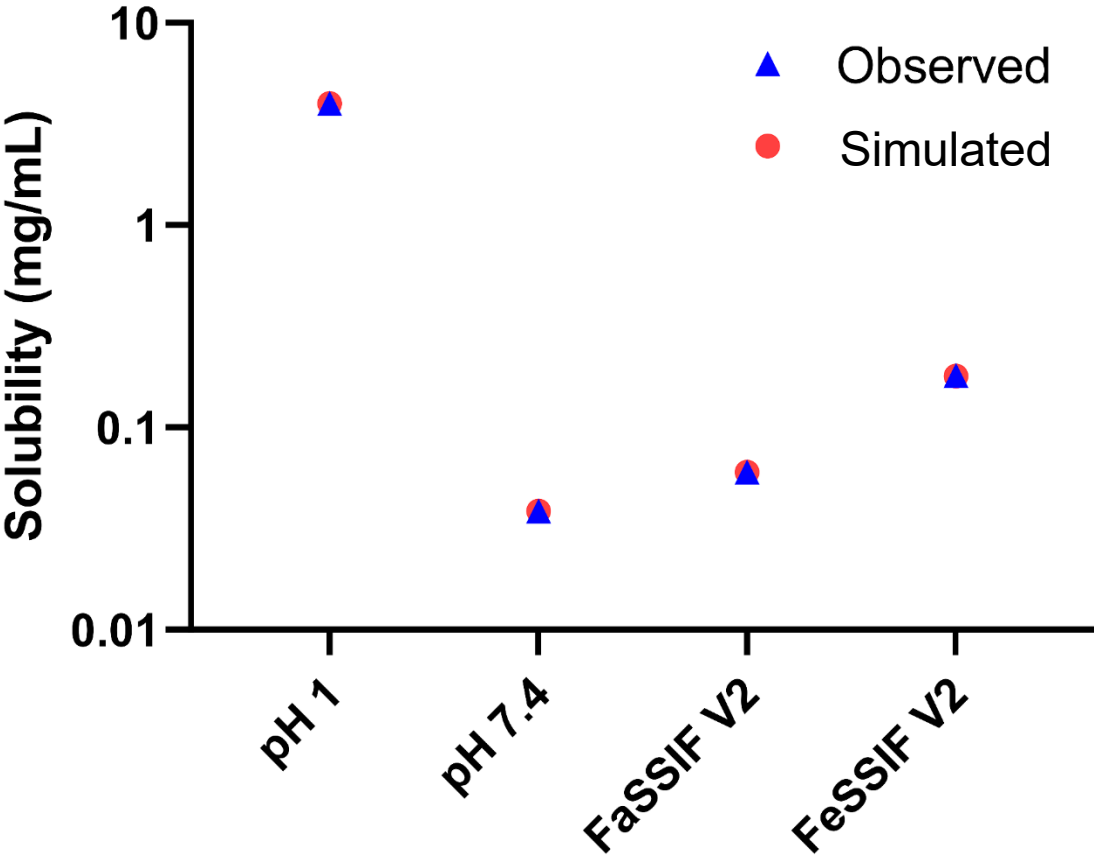


Confirmed Bile Micelle Partition Coefficients

$$S_{Tot}(t) = S_o \cdot S_{o_{scalar}}(t) \cdot \left(1 + \frac{[BS](t)}{C_{H_2O}} \cdot Km:w, u\right) + S_i(t) \cdot \left(1 + \frac{[BS](t)}{C_{H_2O}} \cdot Km:w, i\right) + S_{bound, excip}(t)$$

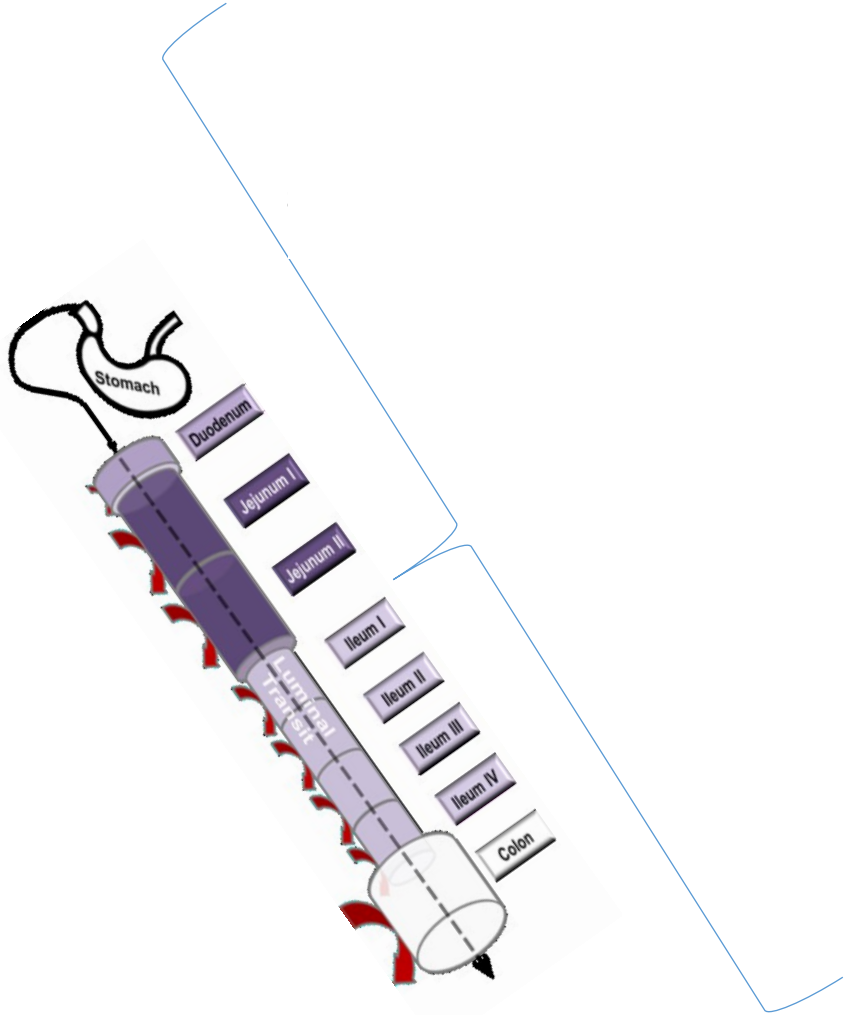
Media	pH	Conc. of Bile Salts (mM)	Solubility (µg/mL)	Experimental Data
FaSSIF V2	6.5	3	60	Xu et al, Mol. Pharmaceutics, 2017, 14 (11), pp 3801–3814
FeSSIF-V2	5.8	10	180	Xu et al, Mol. Pharmaceutics, 2017, 14 (11), pp 3801–3814
		Km/w (neutral)	3.606	Optimized with SIVA 3
		Km/w (ion)	NA	Optimized with SIVA 3

Modeling of *In Vitro* Solubility Experiments



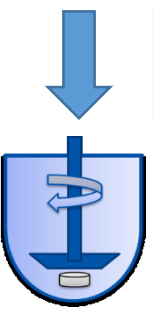
Solubility values in aqueous and biorelevant media are nicely captured

Sequential Modeling of *In Vitro* Biopharmaceutical Experiments



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USP-2 Dissolution Modelling



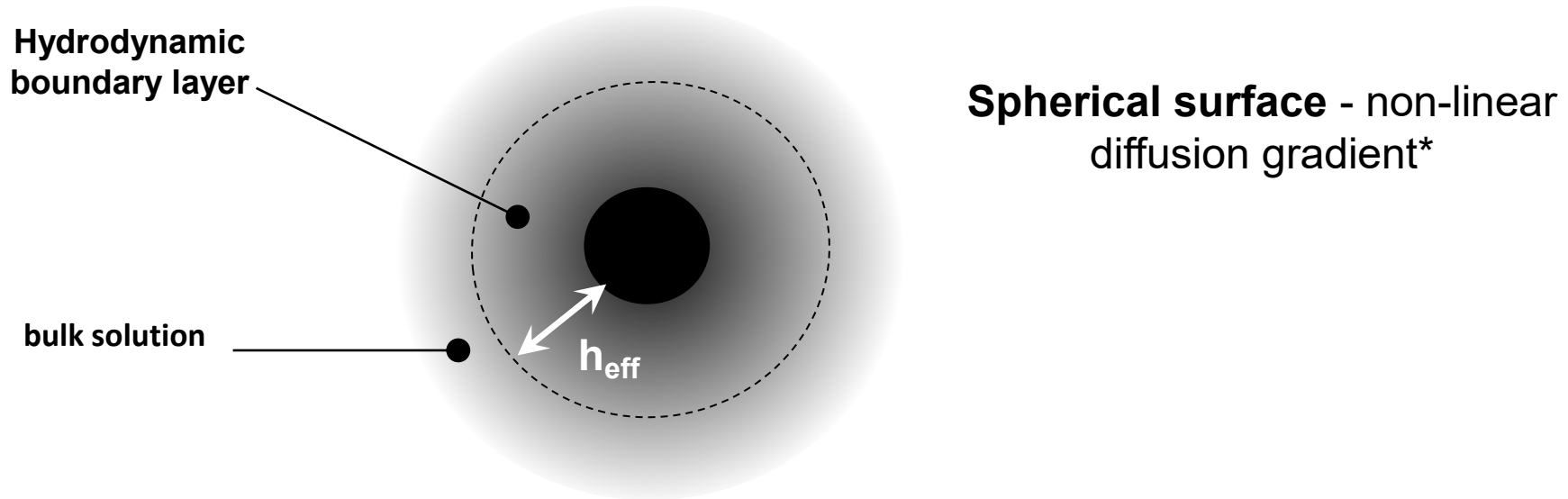
$$DR(t) = -NS \frac{D_{eff}}{h_{eff}(t)} 4\pi a(t) (a(t) + h_{eff}(t)) (S_{surface}(t) - C_{bulk}(t))$$

Confirmed Delayed Release and DLM Parameters

Confirm and/or Estimate Parameters

Mechanistic Dissolution Modelling: Diffusion Layer Models (DLM)

Assumption: Rate limiting step is diffusion across a hydrodynamic boundary layer surrounding the particles of effective thickness h_{eff}



General Form of the Dissolution Rate Equation

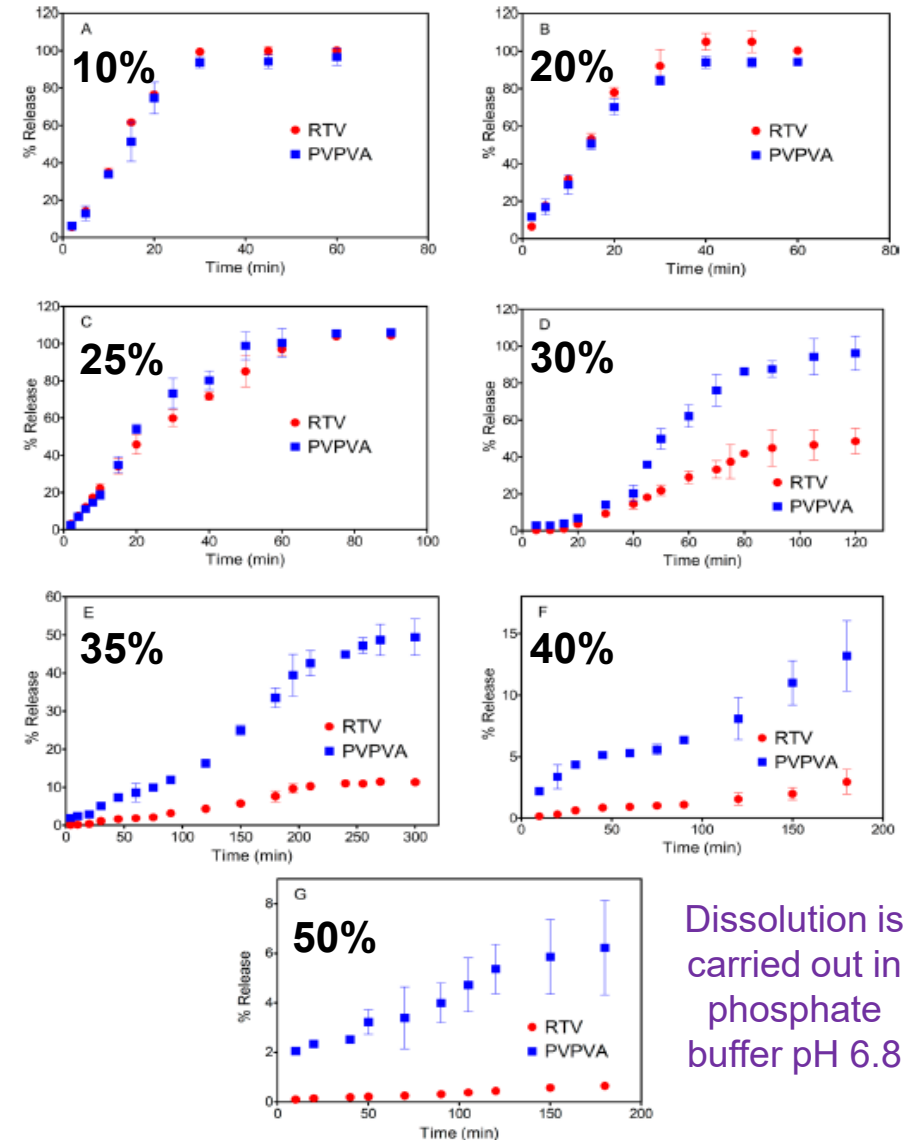
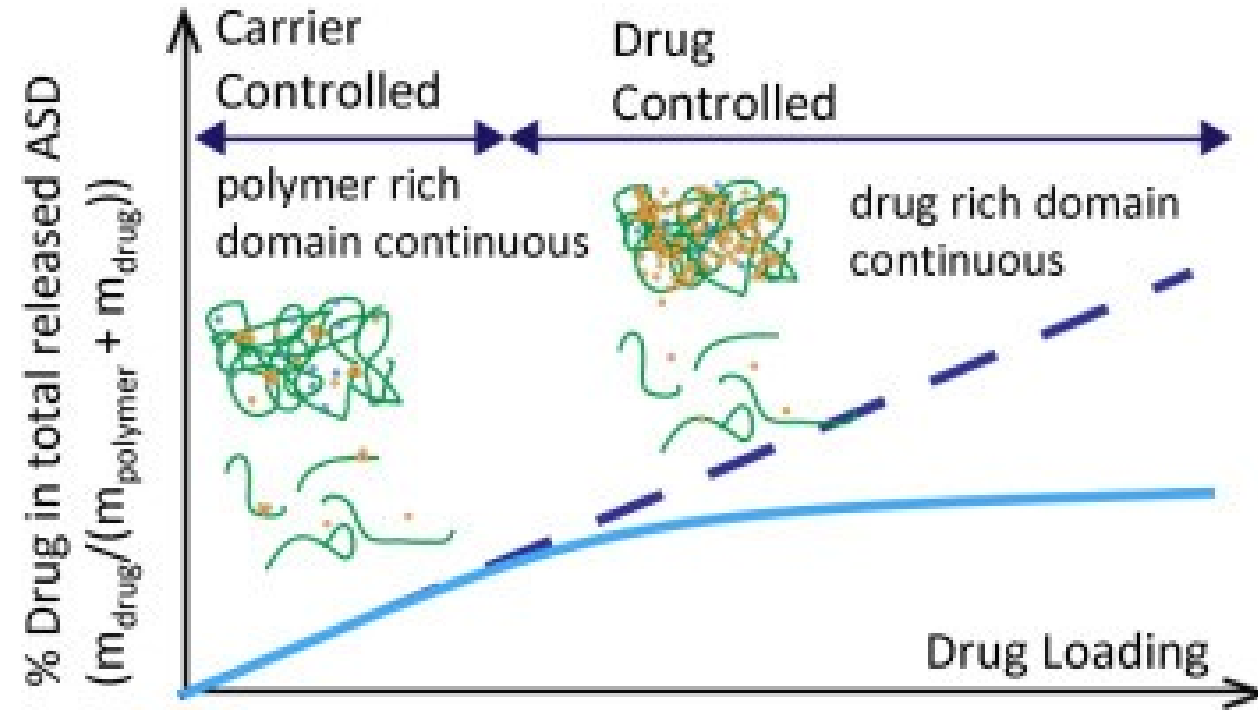
$$DR(t) = \sum_{NBINS}^{i=1} -N_i S_{DR} \frac{D_{eff}(t)}{h_{eff,i}(t)} 4\pi a_i(t) (a_i(t) + h_{eff,i}(t)) (S_{surface}(t) - C_{bulk}(t))$$

a – particle radius; C_{bulk} – bulk lumen concentration; $S_{surface}$ – solubility at particle surface; D_{eff} – effective diffusion coefficient; DR – Dissolution rate; h_{eff} – effective diffusion layer thickness; N – number of particles in PS bin i ; S_{DR} – dissolution DLM scalar; t - time

(Noyes & Whitney, 1897; *Wang & Flanagan, 1999; Further discussion: Sugano, 2010)

Understanding release of drug from amorphous solid dispersion

The release of the drug from the amorphous solid dispersion can be either polymer controlled or drug controlled

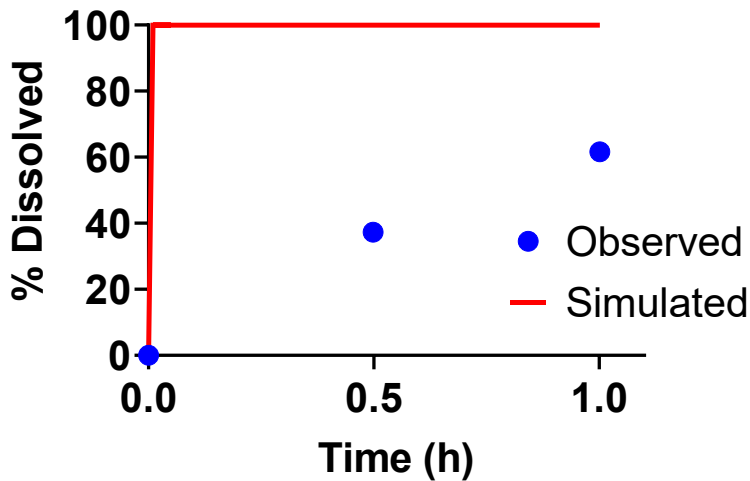


Dissolution is carried out in phosphate buffer pH 6.8

Modeling *In Vitro* Dissolution Experiments in Simulated Gastric Fluid (pH 1.2)

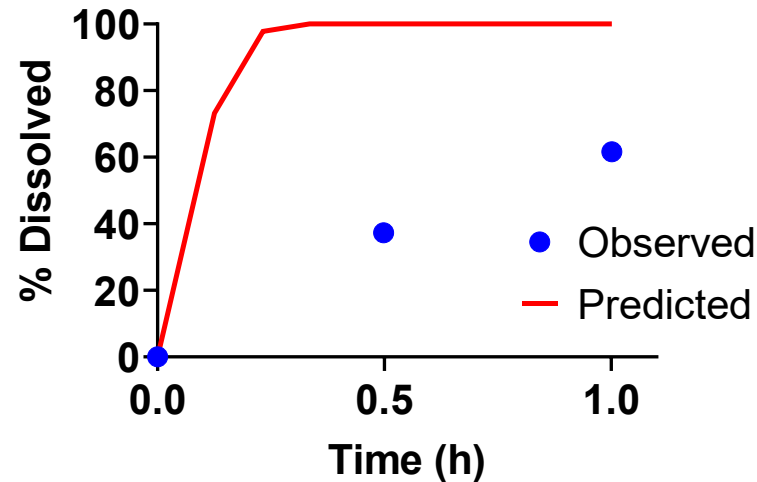
- Ritonavir is a weak base showing high solubility in the acidic pH
- Particle size of the API in commercial batches used in *in vitro* or *in vivo* studies are rarely reported in literature

Scenario 1



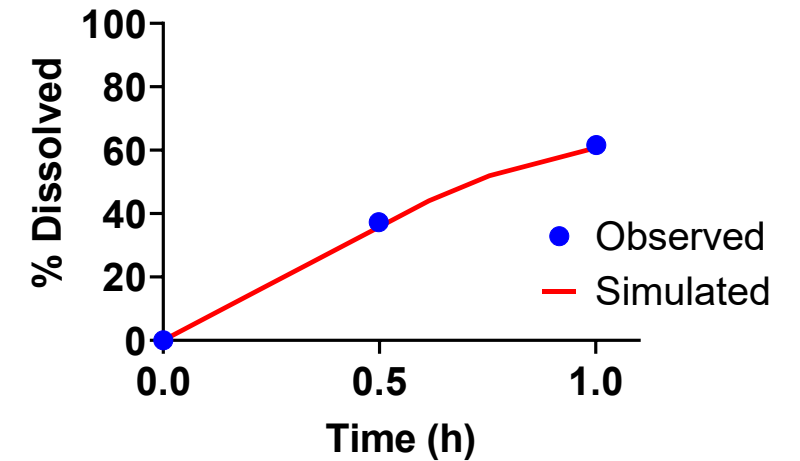
Input particle size – 10 µm
(Default in Simcyp)

Scenario 2



Input particle size – 100 µm

Scenario 3



Fitting a first order release constant
Input particle size – 10 µm

$$\%F_{release,t} = \%F_{max} \left(1 - e^{-kd_1 \cdot (t-t_{lag})} \right)$$

$$F_{max} (\%) = 100\%$$

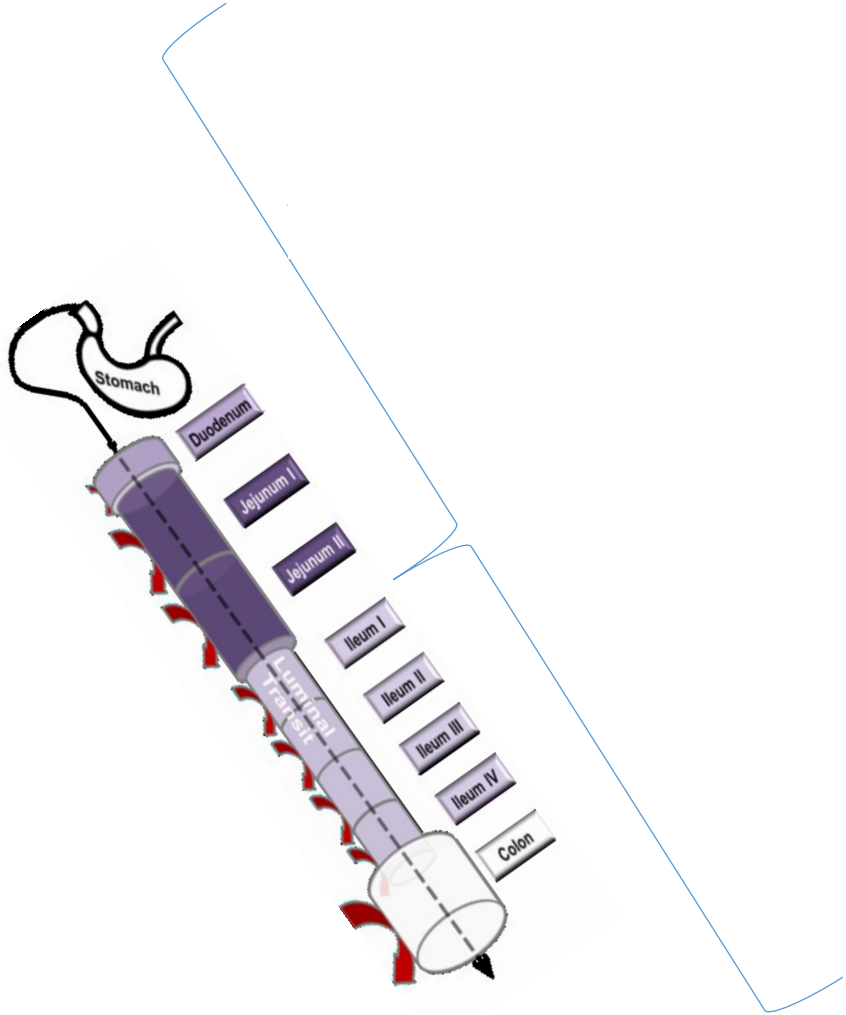
$$K_d (h^{-1}) = 0.017$$

$$T_{lag} (h) = 0$$

Parameters
used in the
PBPK
model

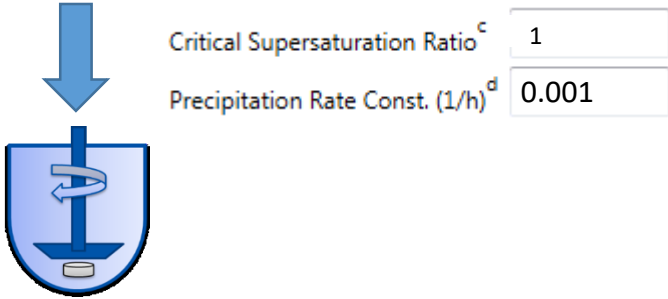
Use of default particle size should be justified via modeling of *in vitro* experiments

Sequential Modeling of *In Vitro* Biopharmaceutical Experiments



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pH Shift Experiment Modelling

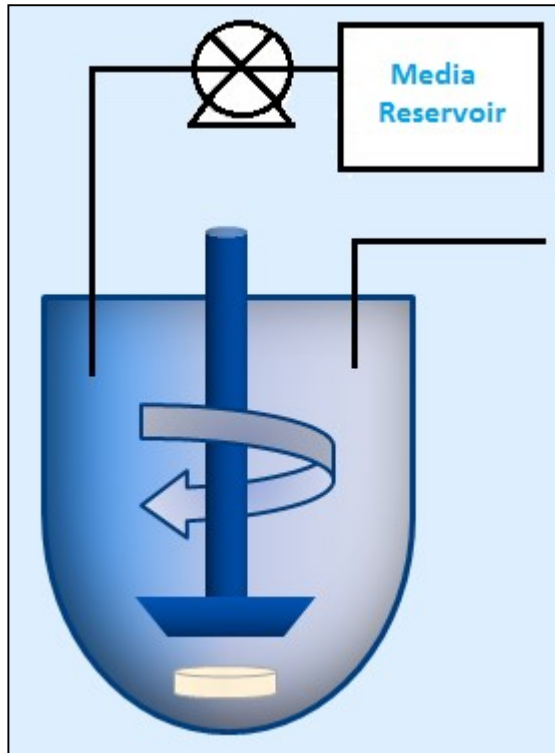


Confirmed Precipitation Parameters

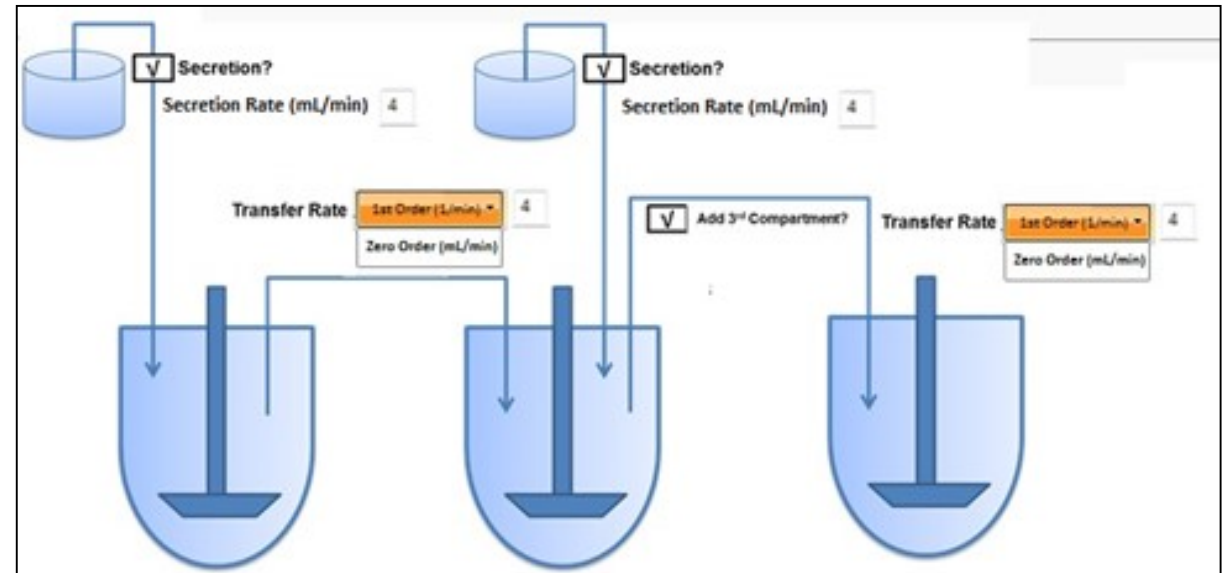
Confirm and/or Estimate Parameters

In Vitro Experiments for Assessing Supersaturation Behaviour

Static - spiked solutions, media change, pH change
solvent shift (two-phase)



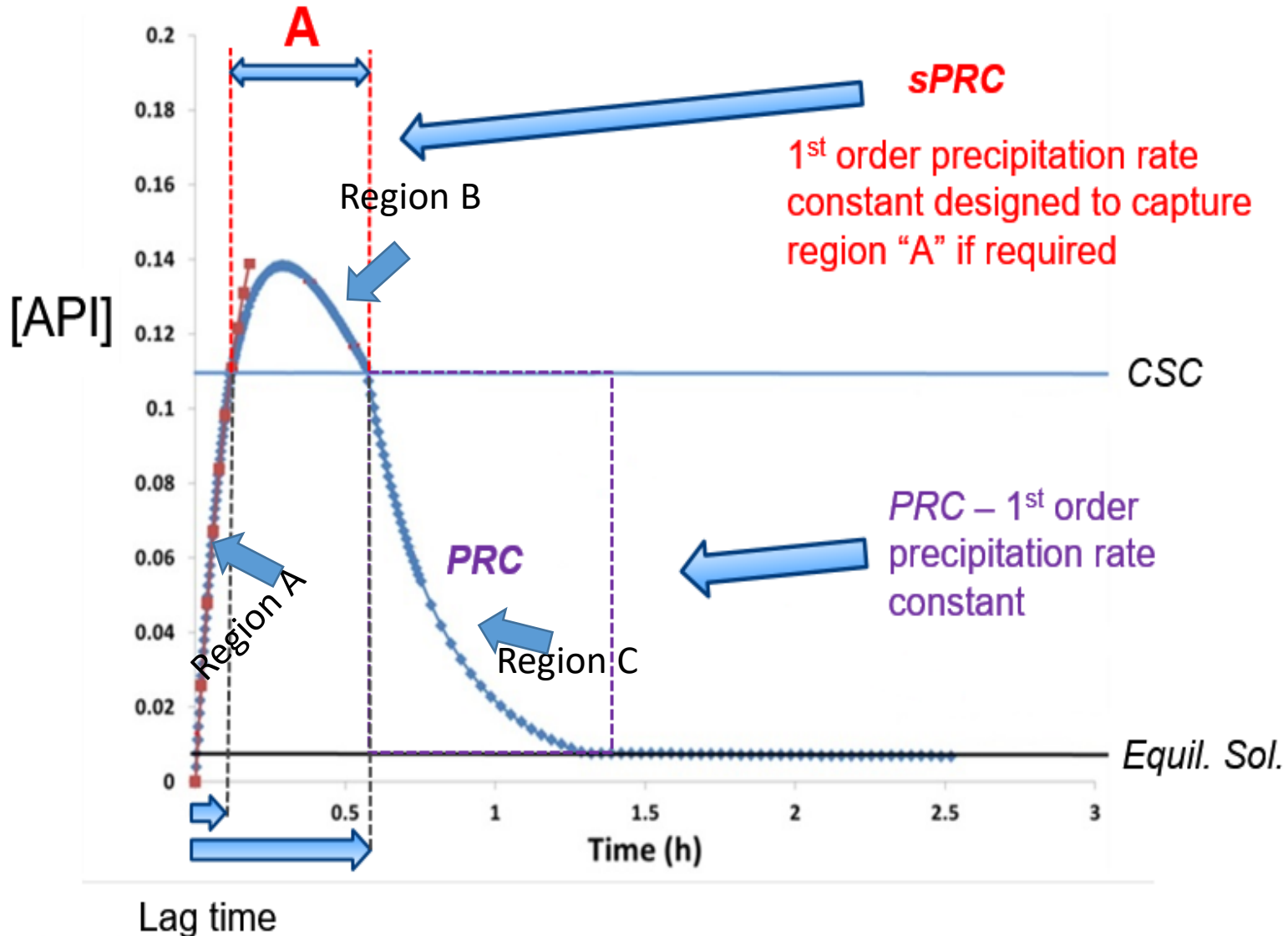
Dynamic - continuous transfer experiments



+ more complex models such as TNO TIN-1

Empirical Supersaturation/Precipitation Model

Time Concentration Profile Receiver Compartment



Empirical Model

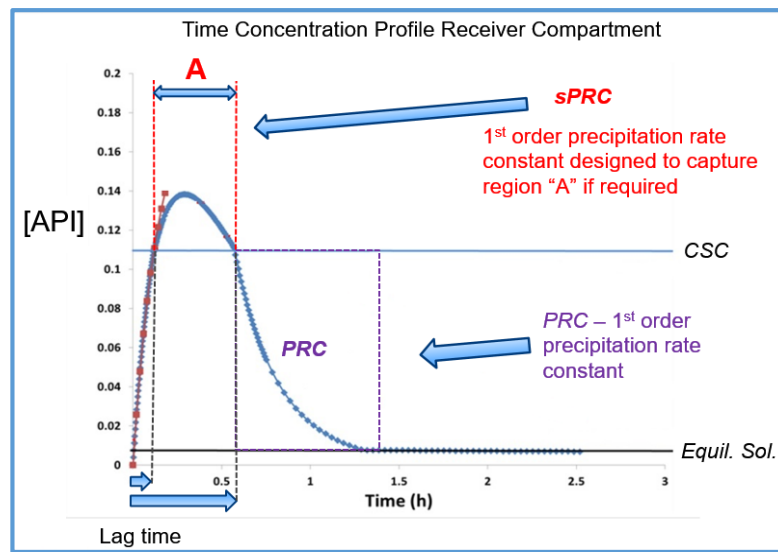
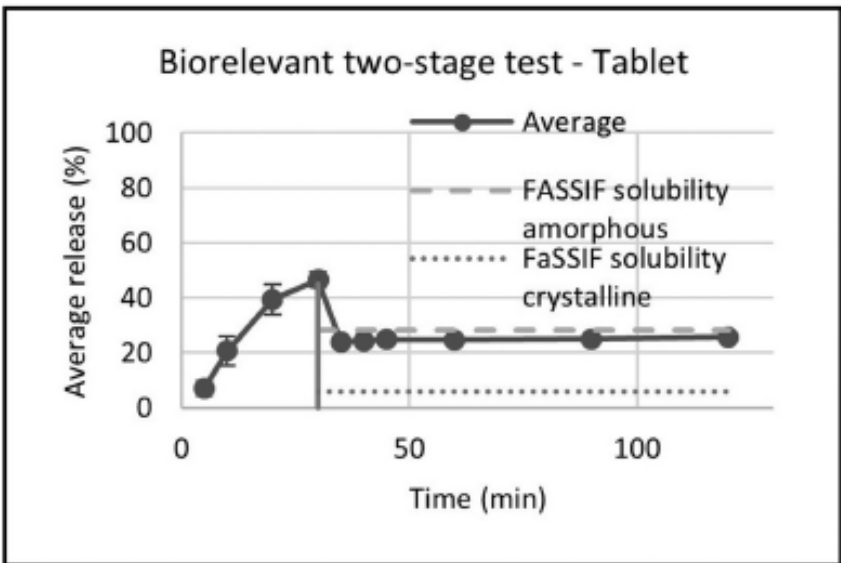
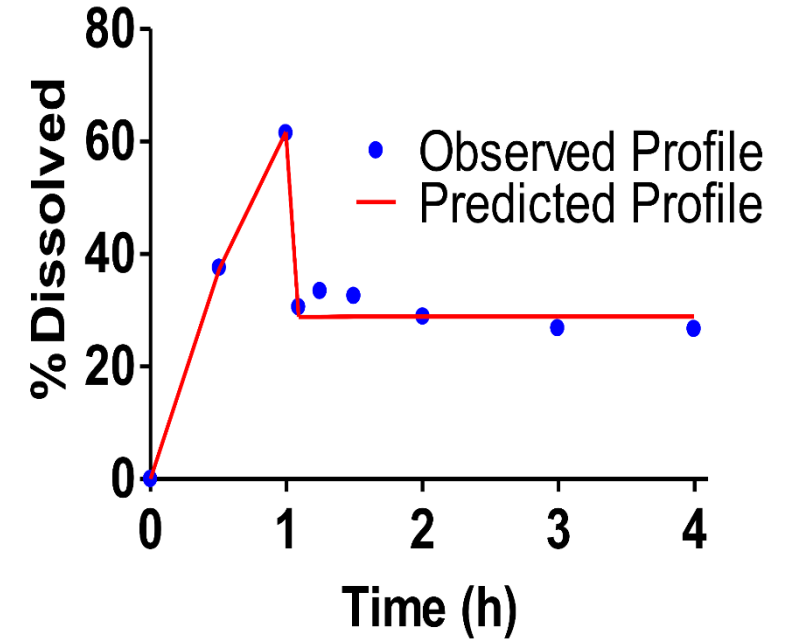
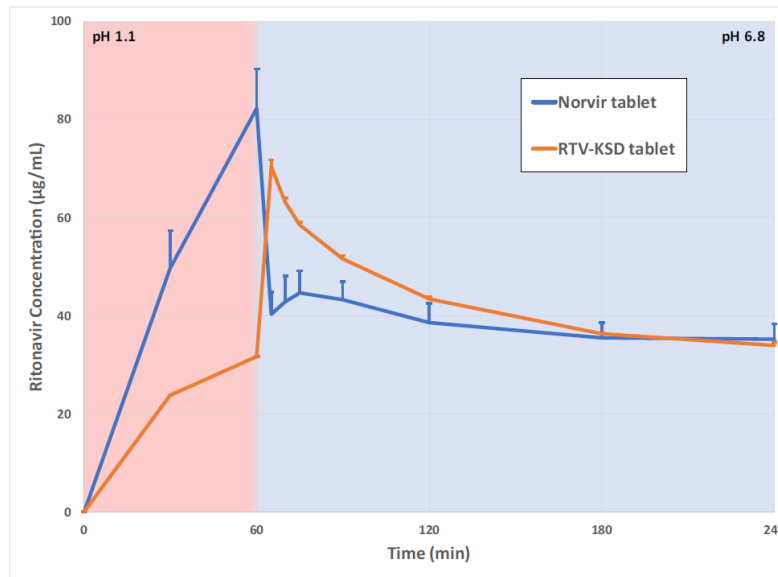
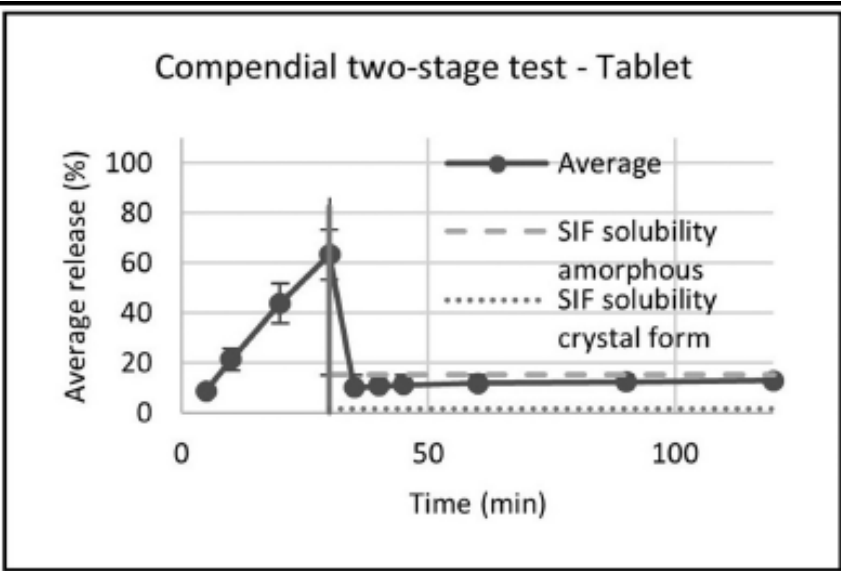
IF supersaturated conditions encountered
THEN

Dissolution stops
Precipitation can only begin
when CSC is reached
CSC is a critical conc. at which
precipitation starts
 [Drug] may continue to rise due to
 slow permeation of drug from skin
Supersaturated conc. may
exceed CSC (CSR x Eq.Sol)

CSC – Critical Supersaturation Concentration
 CSR – Critical Supersaturation Ratio
 PRC – Precipitation Rate Constant (1/h)
 sPRC – Secondary Precipitation Rate Constant (1/h)

Modeling *In Vitro* Drug Supersaturation and Precipitation

Ritonavir precipitates to its amorphous state and does not crystallize



CSR – 1
sPRC – 1000 (1/h)
PRC – 0.001 (1/h)

Parameters used in the PBPK model

Modeling Permeability of Ritonavir in the PBPK Model – Mechanistic Permeability (MechPeff) Model

MechPeff model predicts regional Peff depending on different morphology in each gut segment

$$P_{eff,n} = \left(\left((P_{Trans,0} \cdot f_{neutral,UBL,pH,n} + P_{para,n}) \cdot ACC_n \cdot MVE_n \cdot fu_{UBL,n} \right)^{-1} + (P_{UBL,n})^{-1} \right)^{-1} \cdot FEp,n$$

Mechanistic modelling of *in vitro* experiments (SIVA 4)

Ionisation effects - pH of the UBL

Free fraction effects linked to bile and/or excipient solubilisation

Input Parameters

In the present model, logP of Ritonavir was used to predict regional $P_{Trans,0}$ paracellular

Compound type and pKa

$\log P_{ow}$ or $\log P_{PAMPA}$ but only if predicting $P_{trans,0}$ from these values

$P_{trans,0}$ – predicted, calibrated from predicted Papp, MechP (SIVA 4)

Solubility – aqueous and bile micelle mediated, calibrated/confirmed
– essential for free fraction calculations

MVE – maximum villous surface area expansion

ACC – additional surface area scalar - automatically calculated (Oliver, Rowland et al 1998)

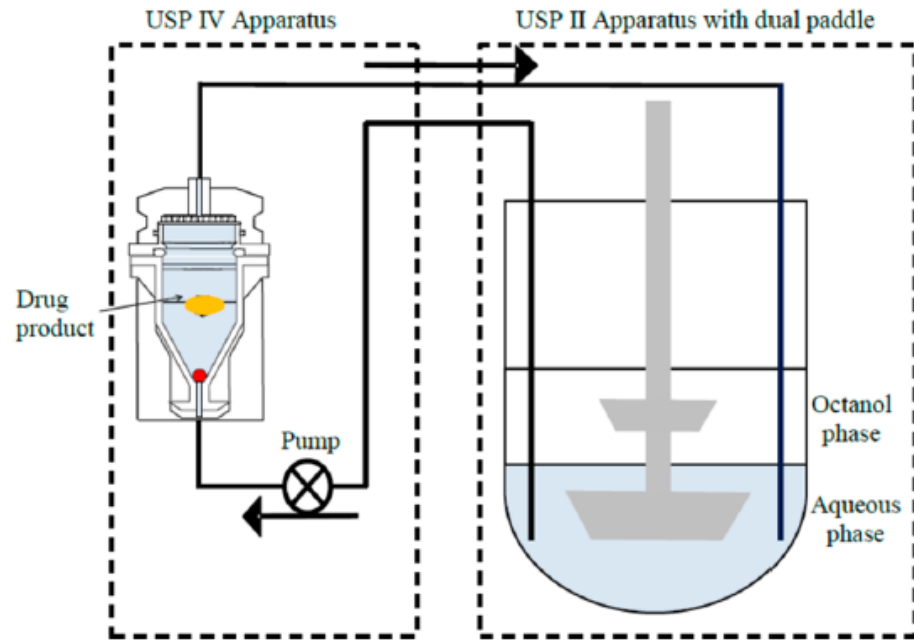
FEp – plicae circulares surface area expansion (humans)

P_{para} – paracellular permeability via Renkin function

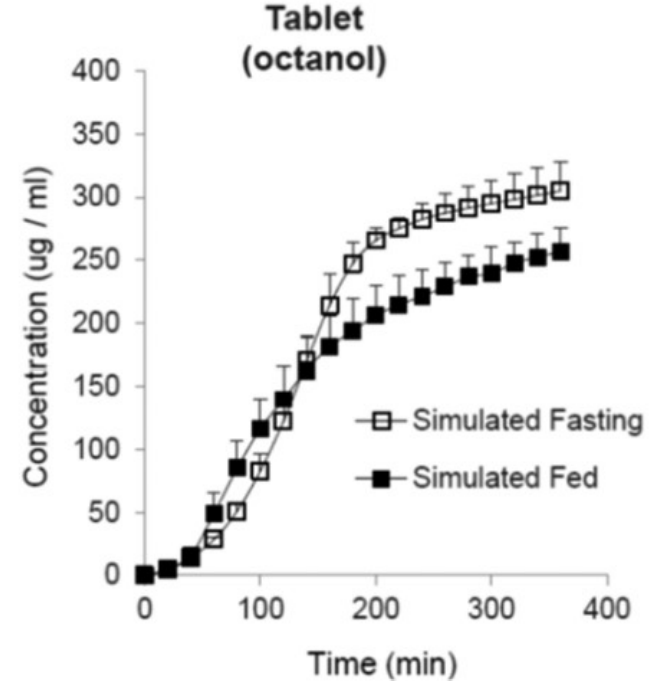
$f_{neutral}$ – fraction unionised (aqueous phase)

fu_{UBL} – free fraction UBL

Why MechPeff ?



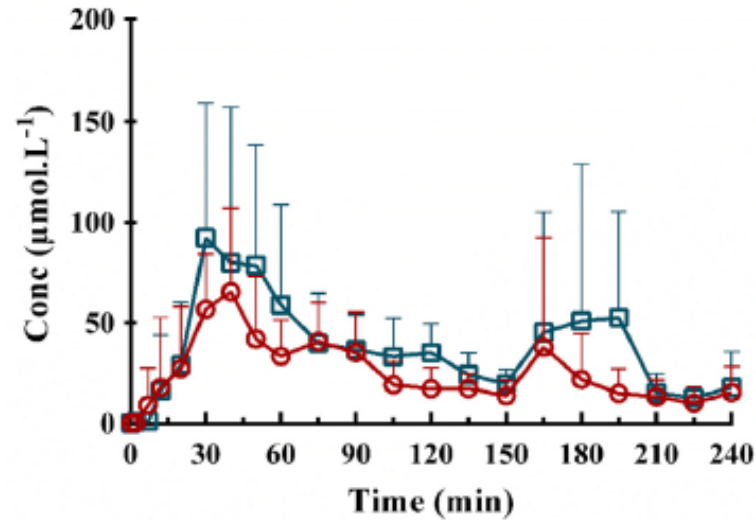
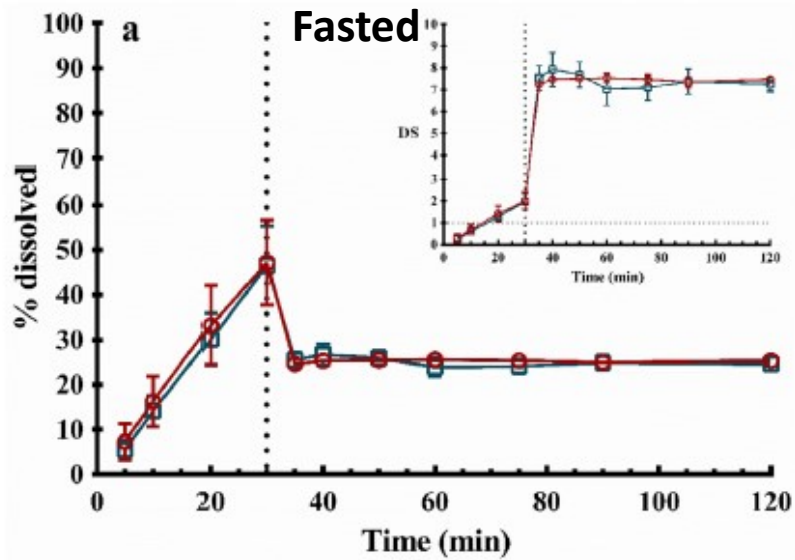
Biphasic dissolution-partition test



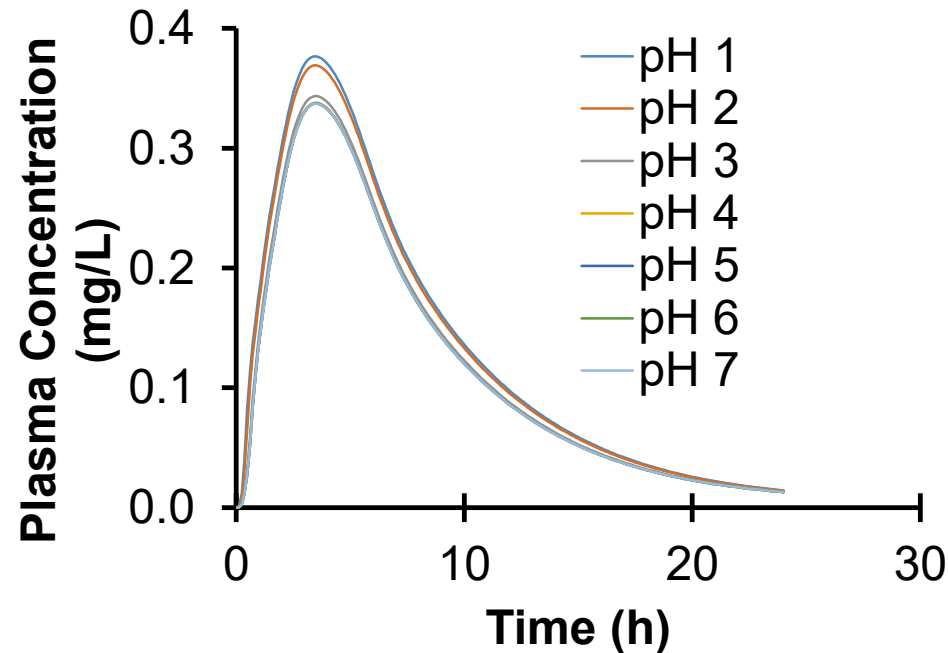
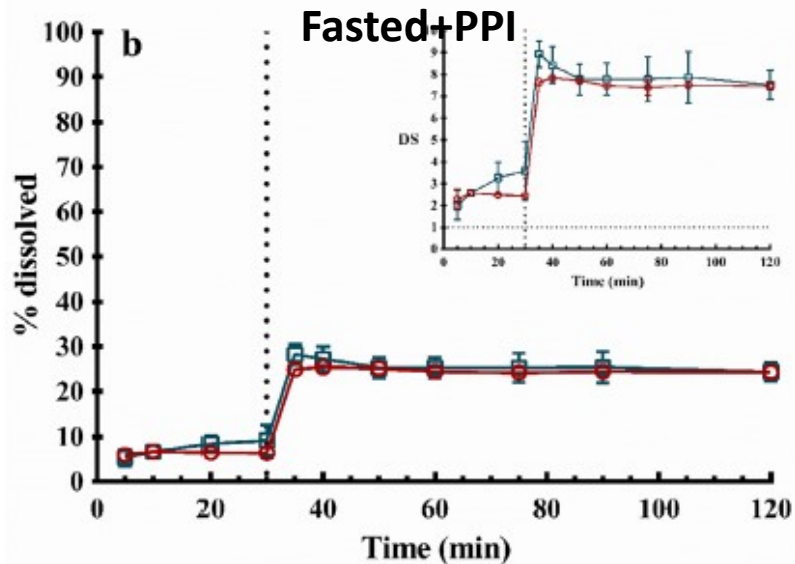
Cumulative RTV concentration-time profiles in octanol from the tablet associated with the simulated fasting and fed media.

Two factors were postulated to explain the food effects –
a. Effect of gastric pH on drug dissolution

Why gastric pH may not be the cause of observed negative food effect?



Mean (+ S.D., n = 5) dissolved drug content - time profiles in the duodenum after oral intake of one tablet of Norvir® (100 mg ritonavir) under fasted state (bullets) and fasted + PPI conditions (squares).



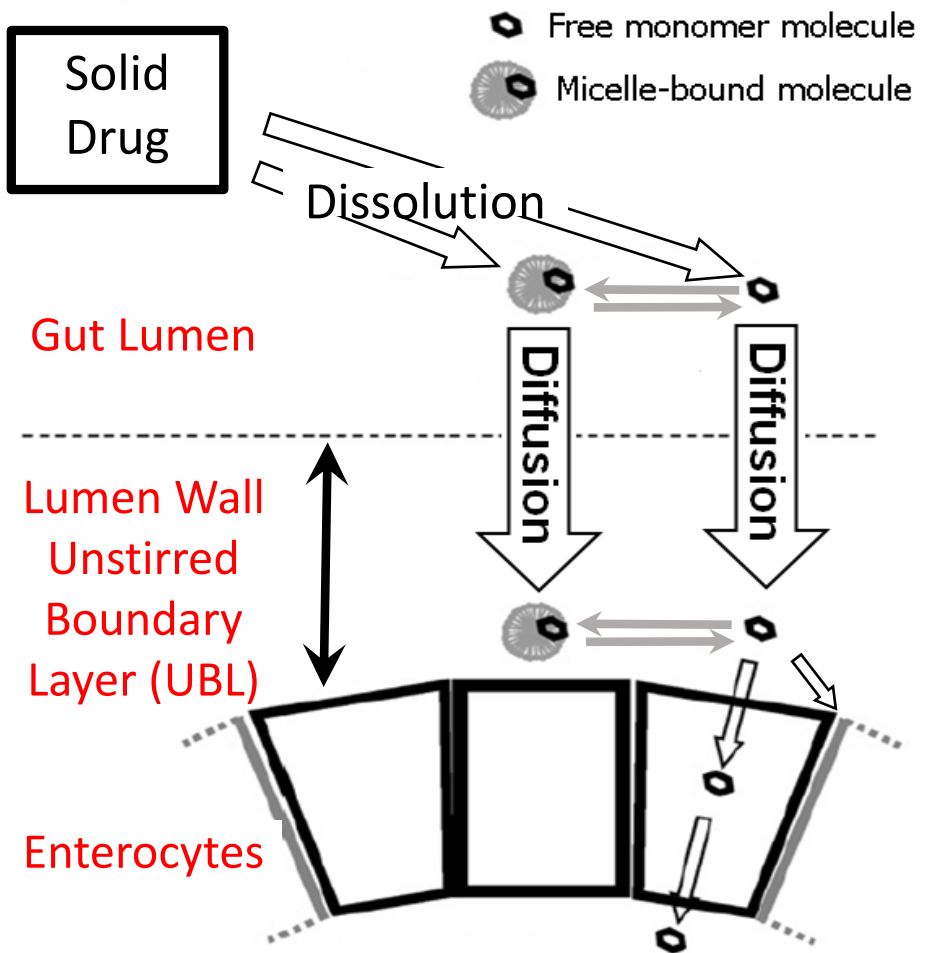
Model Sensitivity to Gastric pH

pH Shift Experiment

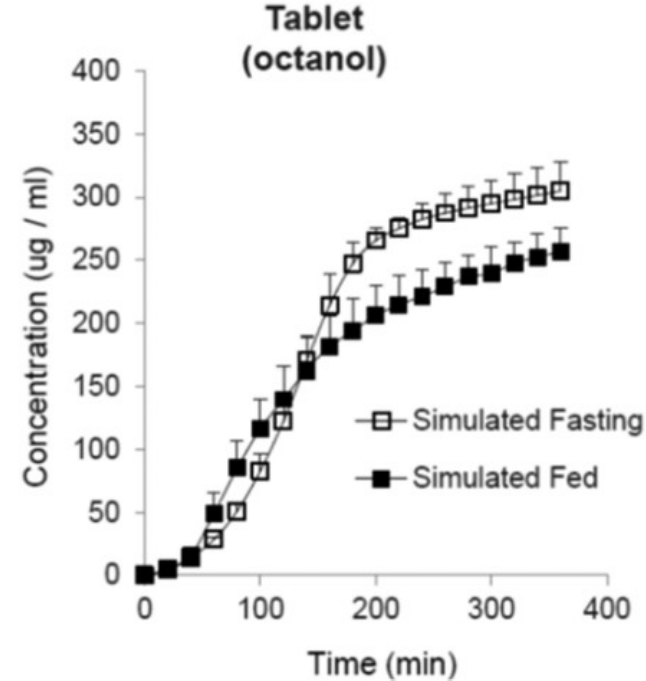
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Abeele et al. Eur J Pharm Sci. 2020 May25; 151:105377

Why MechPeff ?



Adapted from Sugano 2009



Cumulative RTV concentration–time profiles in octanol from the tablet associated with the simulated fasting and fed media.

- Two factors were postulated to explain the food effects –
- Effect of gastric pH on drug dissolution
 - Effect of free fraction of drug on its permeation

Mechanistic Permeability (MechPeff) Model in Simcyp takes into account effect of free fraction of drug available on drug permeation

Static Viscosity Model

- In most of our models, compartments are considered as well-mixed
- Viscosity of the medium affects the diffusion coefficient of the drug, thus influencing drug dissolution and diffusion
- Viscosity of GI lumen changes drastically depending upon the prandial state

Viscosity Equation

$$D_{eff,fast \mid fed} = \left(\frac{1}{\mu_{fast \mid fed}} \right)^{0.987} D_{eff,\mu=1}$$

		Stomach	Duodenum	Jejunum I	Jejunum II	Ileum I	Ileum II	Ileum III	Ileum IV	Colon
Fasted										
Apparent Viscosity	Mean (cP)	3	3	3	3	3	3	3	3	3
	CV (%)	71	71	71	71	71	71	71	71	71
Fed										
Apparent Viscosity	Mean (cP)	232	65	33	12	9	3	3	3	3
	CV (%)	32	27	36	14	2	71	71	71	71

Gastrointestinal compartment viscosity values used in Simcyp for fasted and fed state simulations (based on meta-analysis)

Distribution and Metabolism Related Parameters

- Ritonavir exhibits complex pharmacokinetic profile
- It is mainly metabolized by CYP3A4/5 with minor metabolism from CYP2D6

Parameter	Value	Comment/Reference
fu, plasma	0.015	Hsu, Clin PK, 1998
Vss (L/kg) (Full PBPK)	0.410	Umehara et al, Biopharm Drug Dispos. 2018;39:152
CL(CYP2D6) – BD Sup		Koudriakova et al. Drug Metab Dispos, 1998, 26:552
Vmax	0.93	Corrected for ISEF
Km	1	
ISEF	0.75	
CL(CYP3A4) – BD Sup		Koudriakova et al. Drug Metab Dispos, 1998, 26:552
Vmax	1.37	Corrected for ISEF
Km	0.07	
ISEF	0.24	
CL(CYP3A5) – BD Sup		Koudriakova et al. Drug Metab Dispos, 1998, 26:552
Vmax	1	Corrected for ISEF
Km	0.05	
ISEF	0.24	
Renal Clearance (L/Kg)	0.53	Denissen et al. 1997

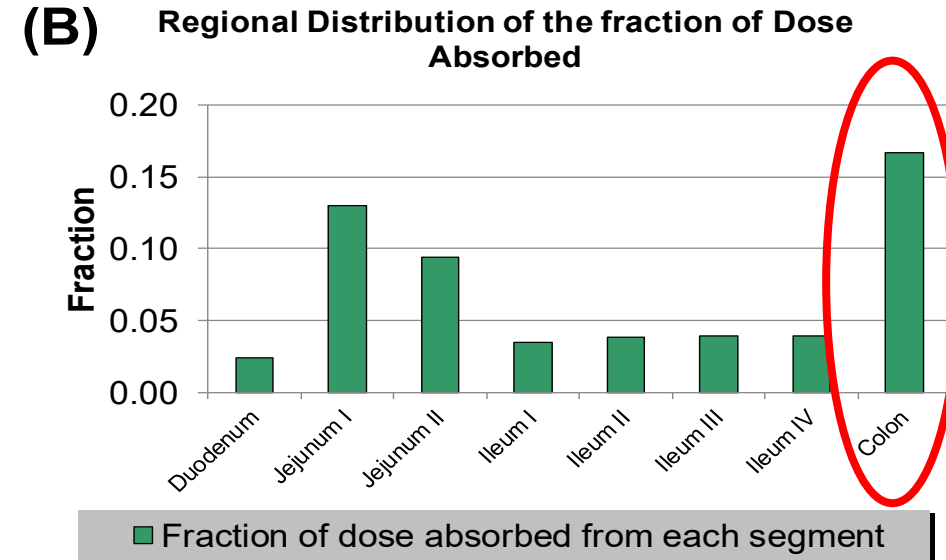
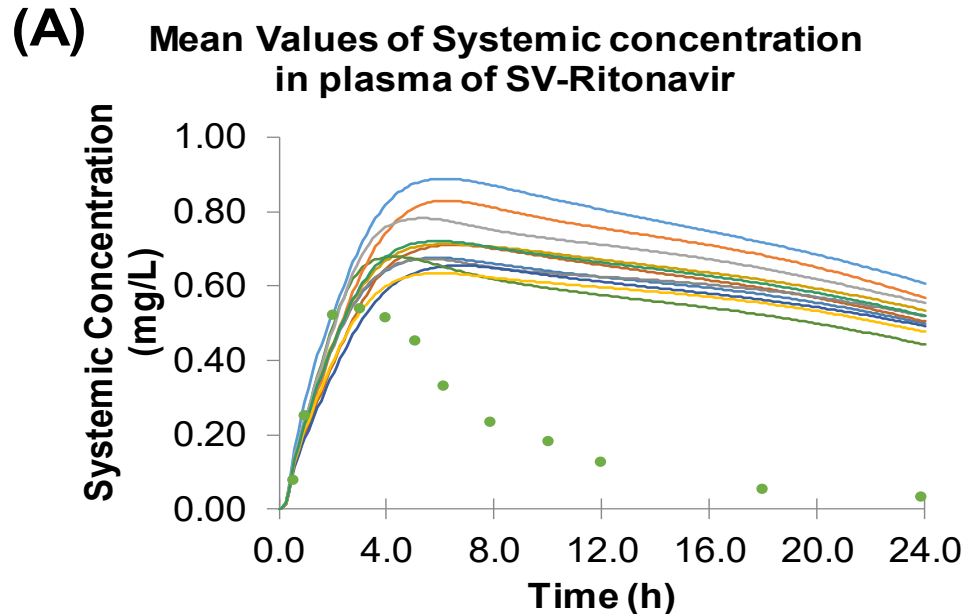
Interaction Related Parameters

- Ritonavir exhibits competitive and mechanism based inhibition of CYP3A4/5
- Ritonavir also is competitive inhibitor of CYP2D6 and Pgp
- Ritonavir is also an inducer CYP3A4/5

Parameter	Value	Comment/Reference
Mechanism Based Inhibition		
K_{app} (CYP3A4/5) (μM)	0.25	Inhibition constant, Kirby et al. ¹³
k_{inact} (CYP3A4/5) (1/h)	19.8	Inactivation rate of enzyme, Kirby et al. ¹³
$f_{u(mic)}$ (CYP3A4/5)	0.71	Kirby et. al. ¹³
Induction		
$IndC50_{(CYP3A4/5)}$ (μM)	1 (CV 30%)	Induction constant, Fahmi et al. ¹⁸
$f_{u(inc)}$ (CYP3A4)	1	Fraction unbound <i>in vitro</i> , Fahmi et al. ¹⁸
$Indmax_{(CYP3A4)}$	68.5 (30%)	
Competitive Inhibition		
K_i (CYP3A4/5) (μM)	0.0021	Inhibition constant, Shebley et al. ⁶
$f_{u_{mic}}$ (CYP3A4/5)	0.925	Fraction unbound <i>in vitro</i> , Shebley et al. ⁶
K_i (CYP2D6) (μM)	0.04	Optimized (to recover DDIs)
$f_{u_{mic}}$ (CYP2D6)	1	
K_i (Apical Efflux, Intestine P-gp) (μM)	0.03	Optimized (to recover DDIs)
$f_{u_{mic}}$ (Apical Efflux, Intestine P-gp)	1	
K_i (Liver Canalicular P-gp) (μM)	0.03	Optimized (to recover DDIs)
$f_{u_{mic}}$ (Liver Canalicular P-gp)	1	

Initial Bottom Up Predictions – Ng et al 2008 Fasted State

Simulations were carried out using 10 trials of 27 individuals per the clinical study design reported by Ng et al. in fasted state



Absorption phase to 4 h was relatively well captured, giving confidence in the absorption-related parameters

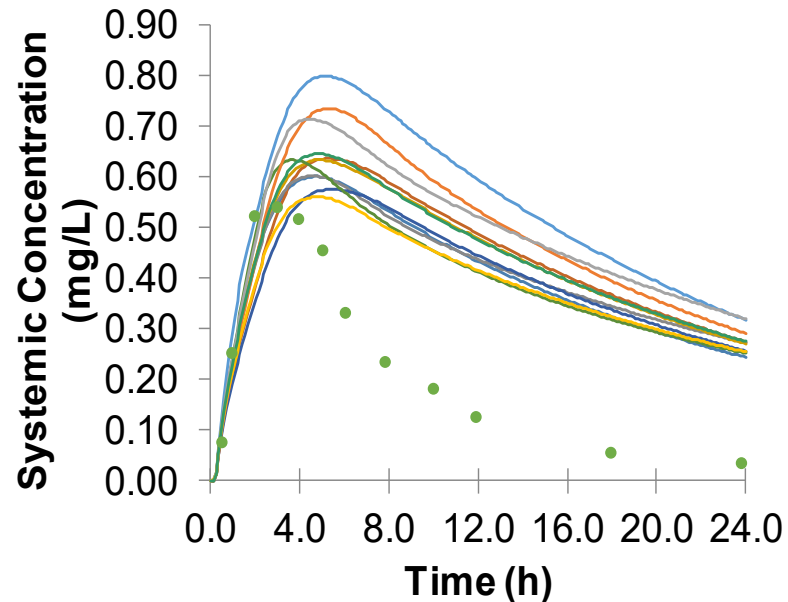
Two plausible reason for over-prediction of the terminal elimination phase:

- Over-prediction of drug absorption in the colon
- In vivo* clearance of the compound is not captured correctly by metabolic IVIVE (Please note - intravenous data for this compound are not available in literature).

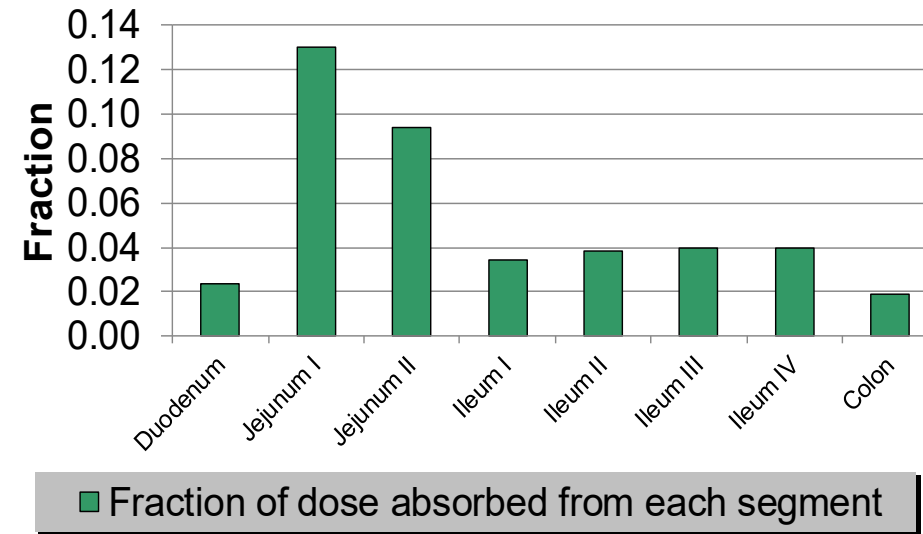
First Modification - Adjustment of Colon Absorption Scalar

- Norvir is an immediate release tablet, majority of absorption should take place in the small intestine
- Understanding of drug absorption in the colon is poor, binding to faeces etc.
- Colon absorption scalar was reduced to 0.1

(A) Mean Values of Systemic concentration in plasma of SV-Ritonavir



(B) Regional Distribution of the fraction of Dose Absorbed

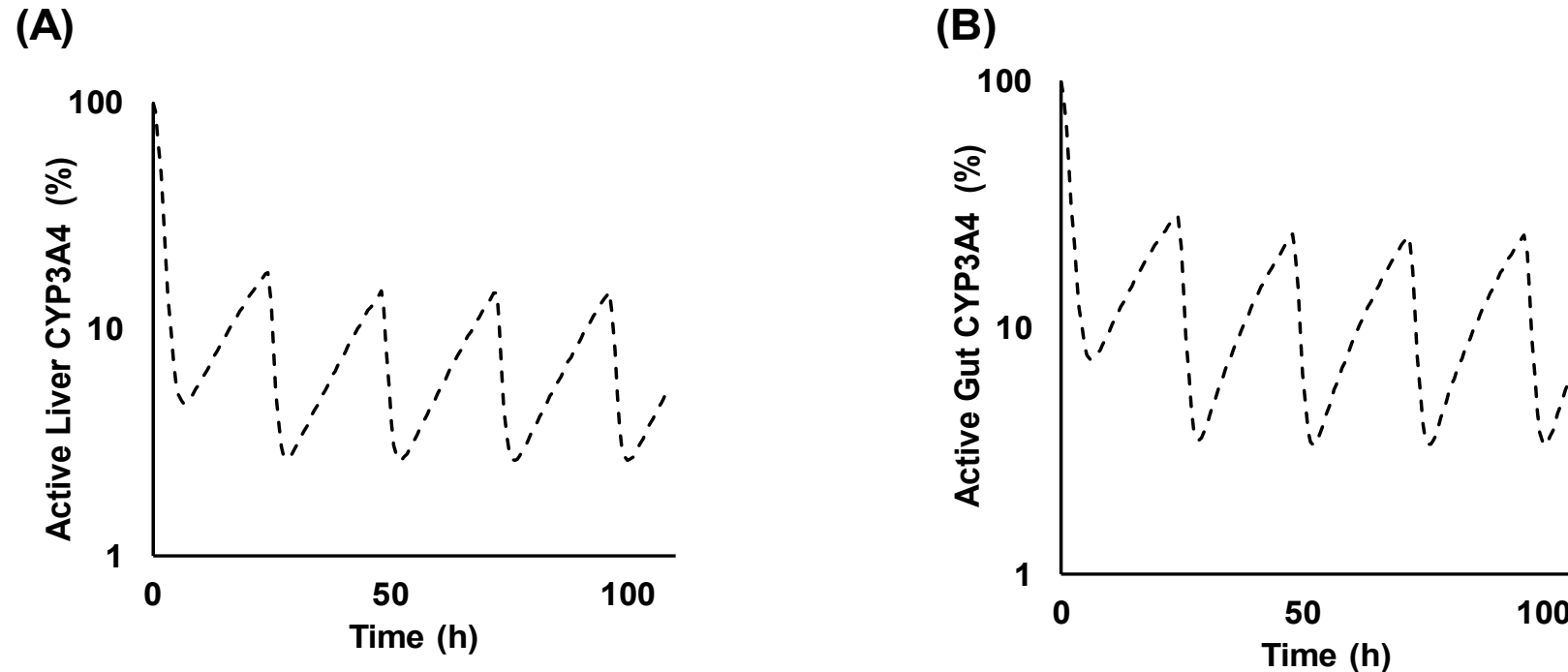


- Predictions are slightly better, C_{\max} and T_{\max} are relatively well captured
- Terminal phase is still overpredicted, AUC >two-fold over prediction suggesting that clearance is not captured correctly

Second Modification – Parameter Estimation of Additional Hepatic Intrinsic Clearance

- Ritonavir is a strong mechanism-based inhibitor of CYP3A4

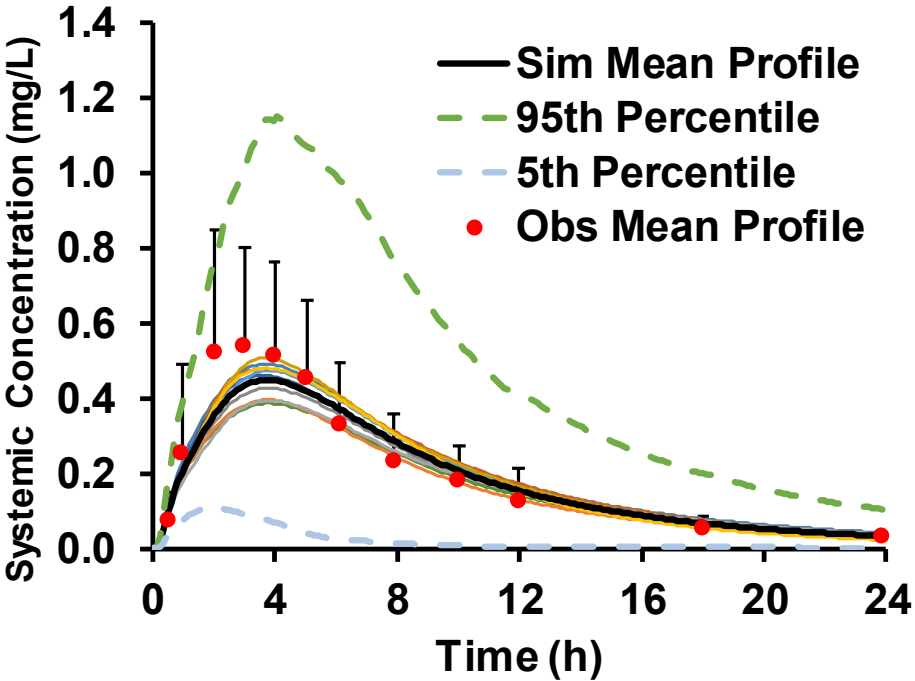
Simulated time dependent changes in hepatic (A) and gut (B) CYP3A4 abundance levels following administration of 100 mg QD oral administration of ritonavir for four days



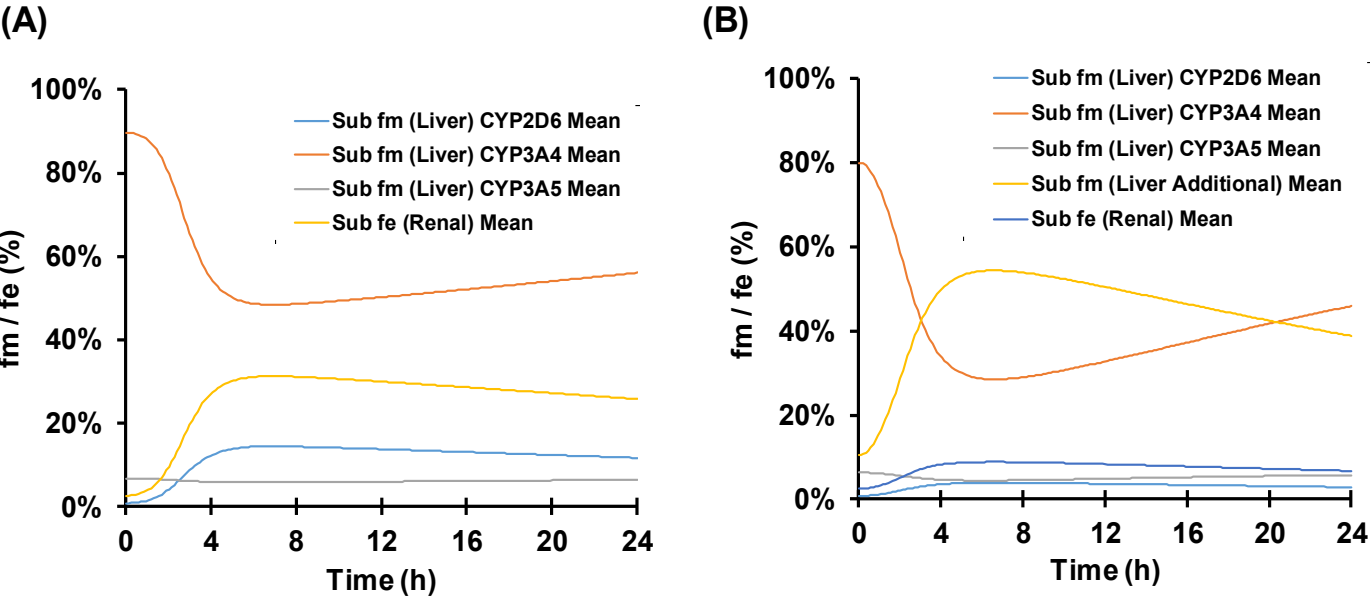
- Maximal inhibition of hepatic (4.77% activity remaining) and gut (7.59% activity remaining) CYP3A4 abundance on day 1 could be observed following administration of 100 mg of ritonavir.
- The fact that we cannot capture clearance of ritonavir suggests that there is additional clearance mechanism *in vivo*, which takes over in clearing the ritonavir from the systemic circulation once hepatic CYP3A4 is inhibited by ritonavir.

Second Modification – Parameter Estimation of Additional Hepatic Intrinsic Clearance

A value of 75 $\mu\text{L}/\text{min}/\text{mg}$ additional hepatic clearance was found to capture well the observed fasted state Ng et al 2008 data (Initially analyzed by Sensitivity Analysis)



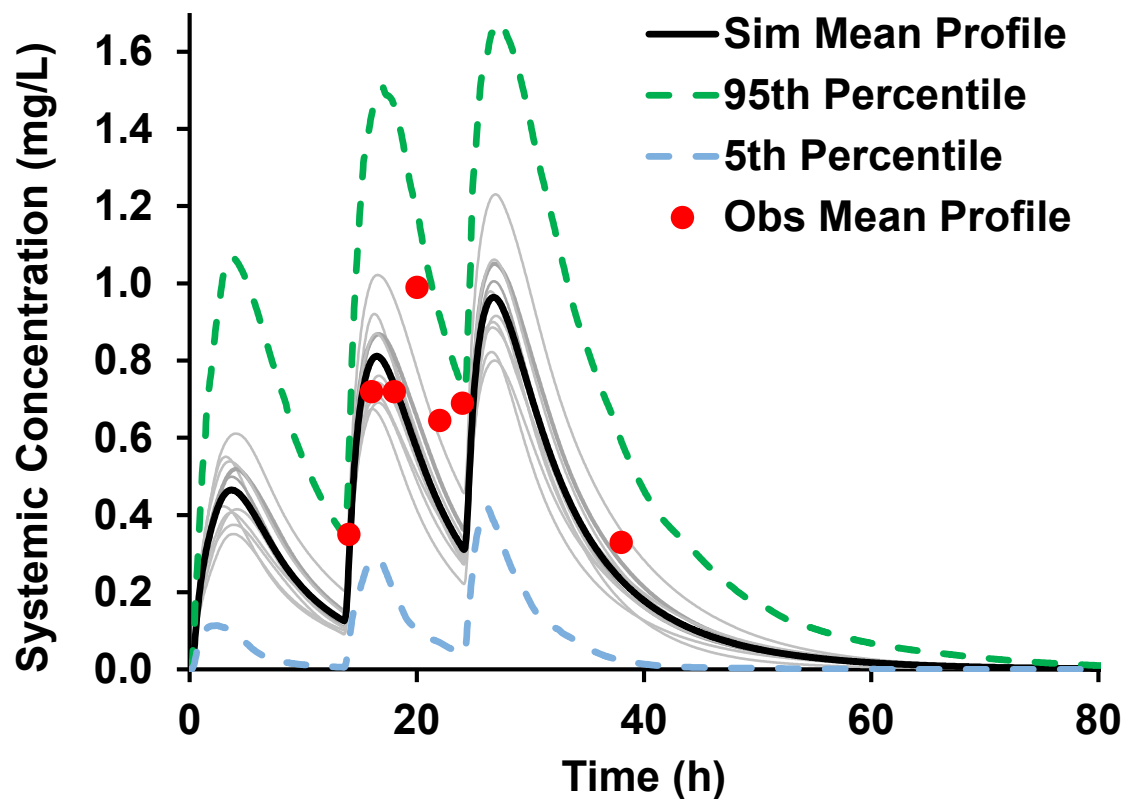
Simulated Ritonavir Concentration Time Profile of Ng et al 2008 study with additional hepatic clearance and colon absorption scalar.



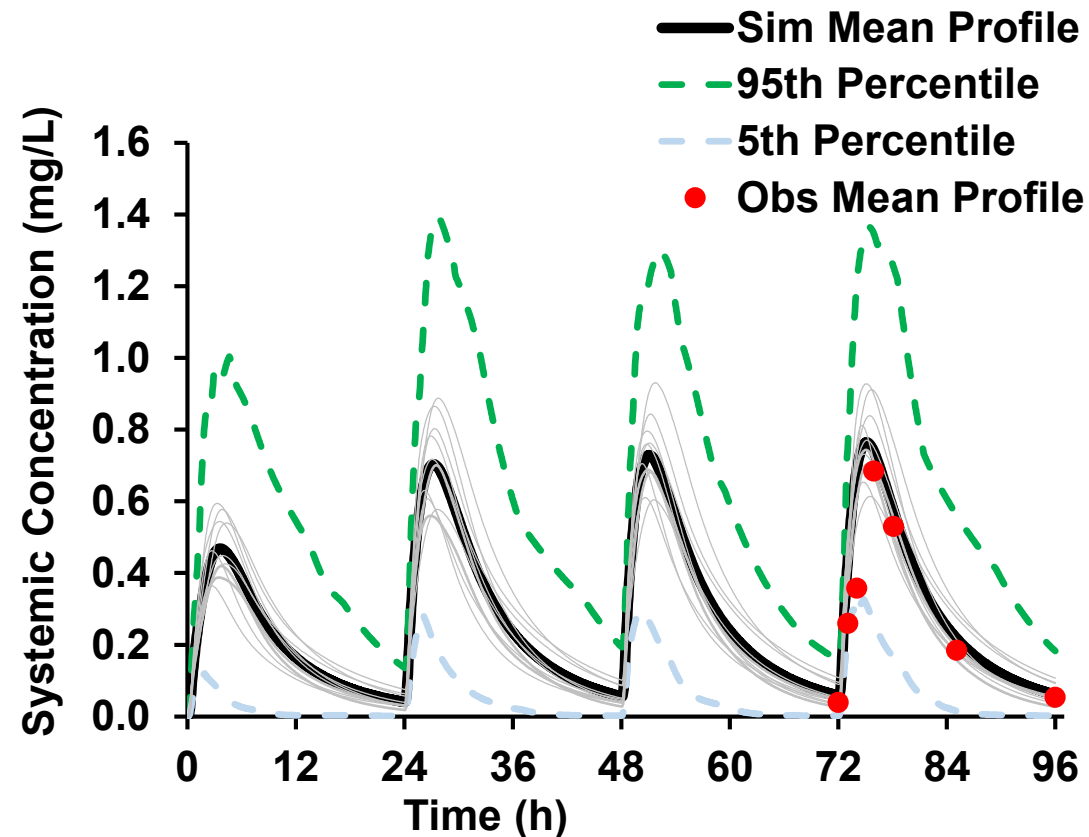
Time variant changes in percentage fraction metabolized (fm) and fraction excreted following oral administration of ritonavir (100 mg) in fasted state (A) without inclusion of additional intrinsic hepatic clearance and (B) with inclusion of additional intrinsic hepatic clearance.

Verification of the Model – Multiple Dose Studies

Greenblatt et al., 2009

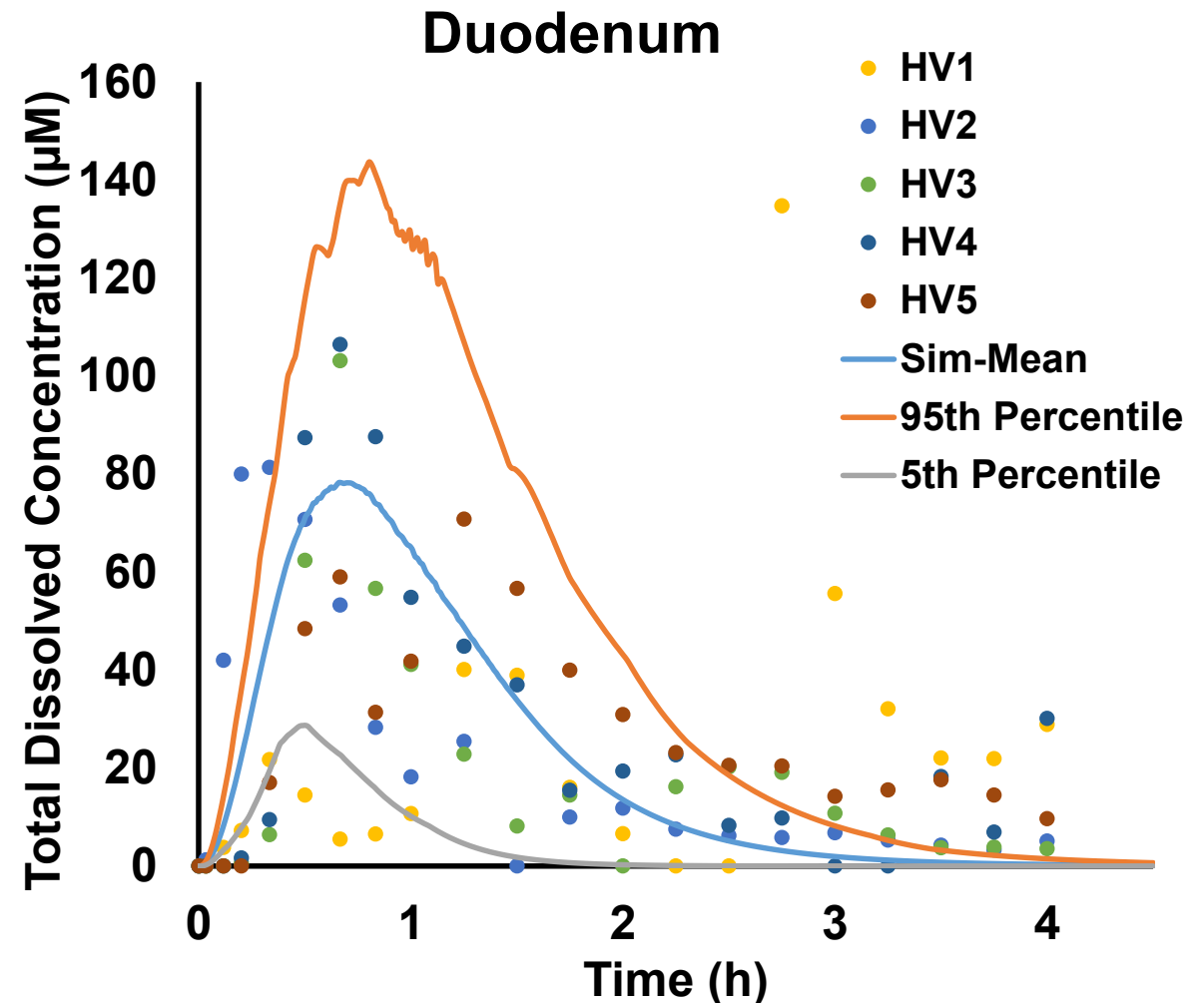
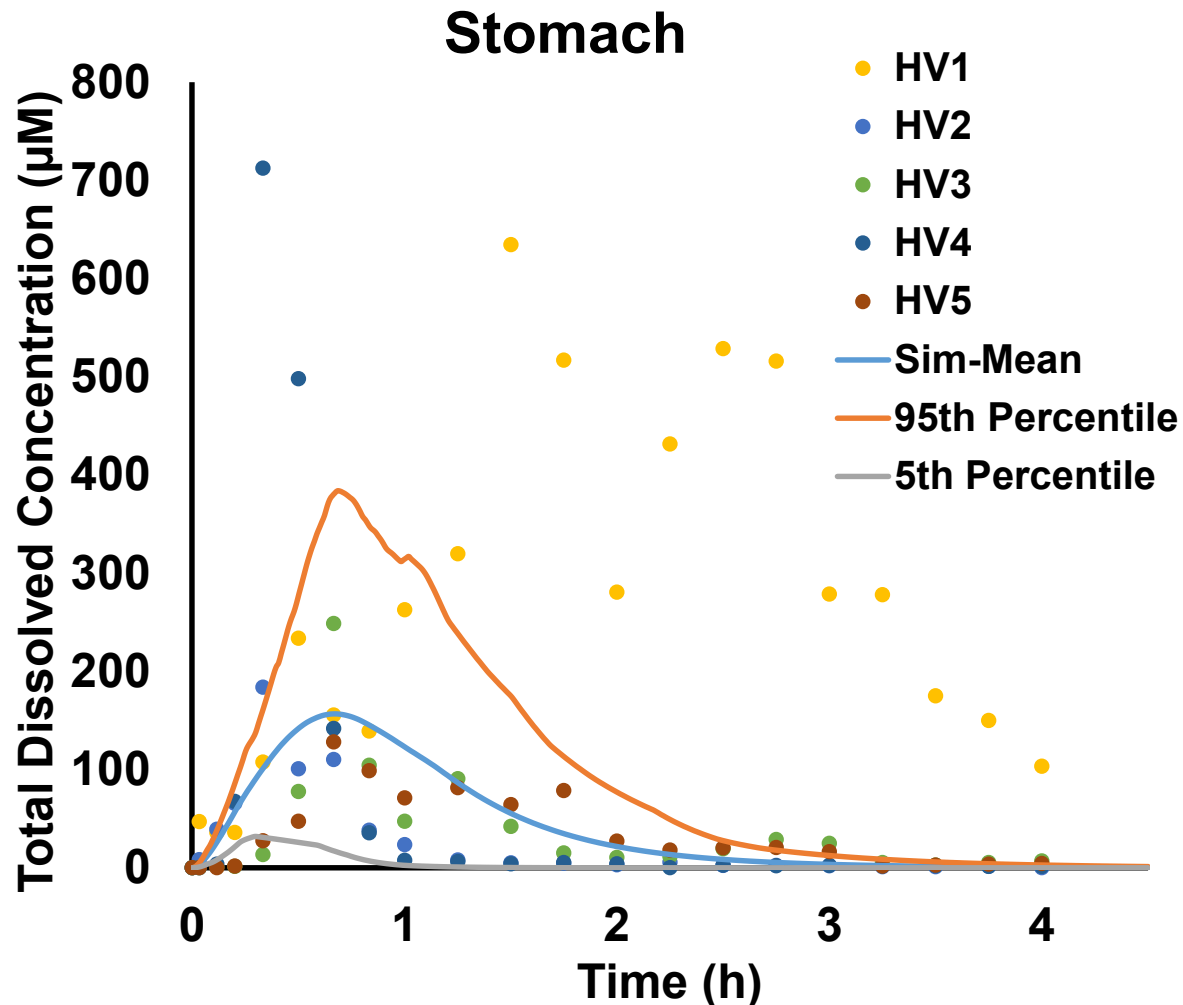


Kasserra et al., 2010



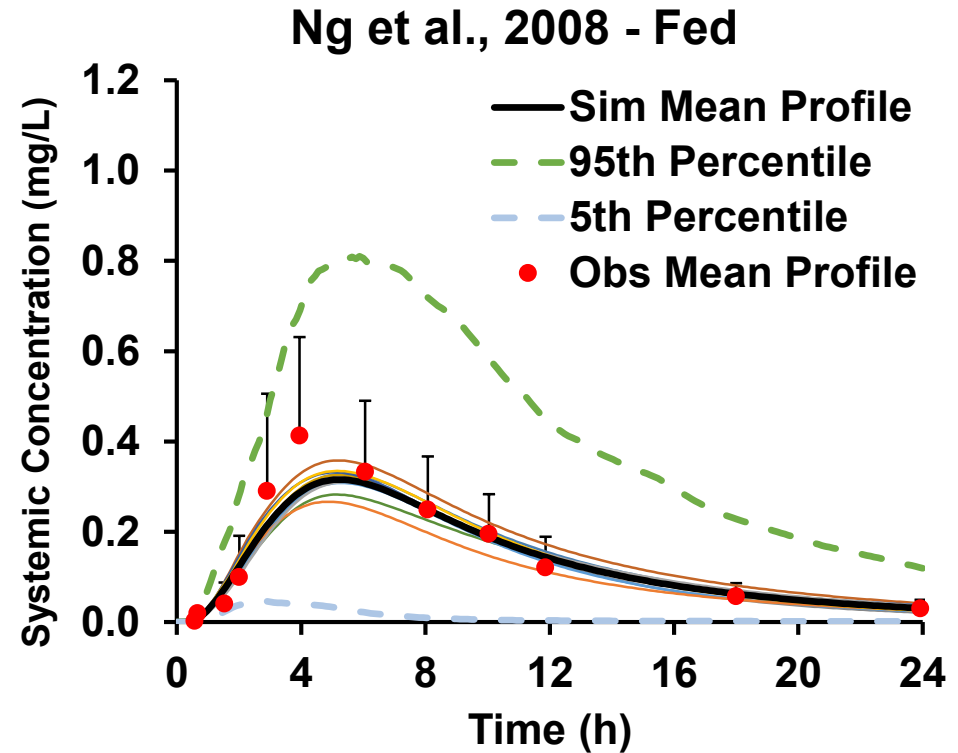
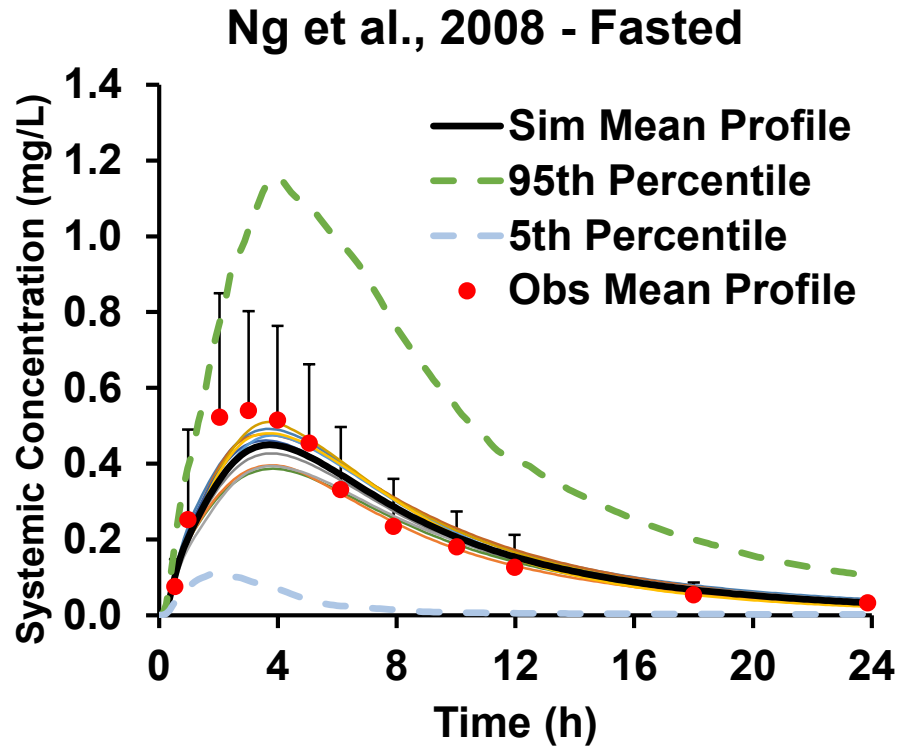
The fact that the model is able to recover multiple dose exposure following optimization of additional clearance from a single dose study also suggests that the model is able to capture the fraction metabolized by CYP3A4/5 relatively well.

Verification of the Model – Luminal Concentration Profiles



The model was able to capture both stomach and duodenal luminal profiles of ritonavir

Prediction of Negative Food Effects

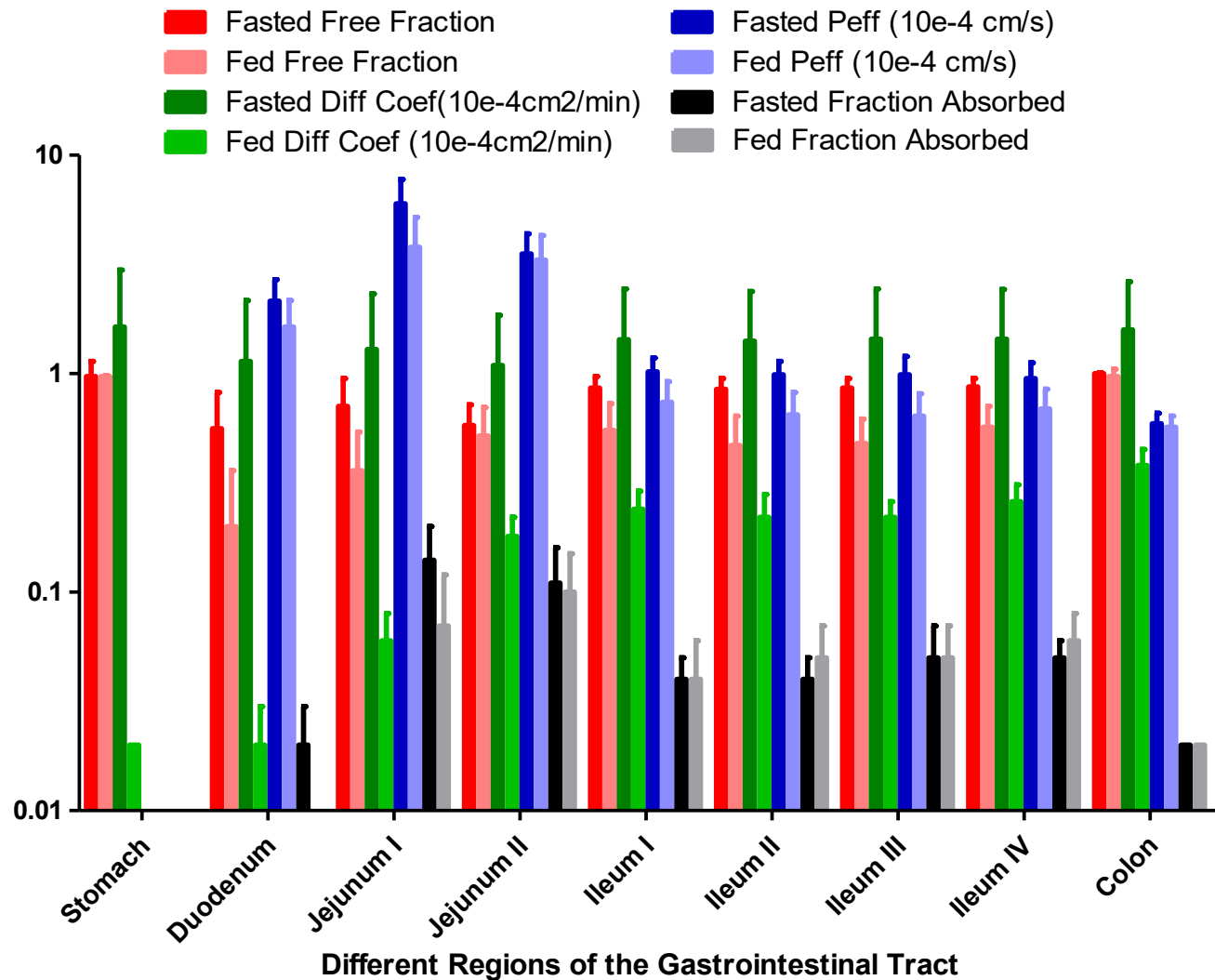


Simulated and Observed pharmacokinetic parameters (arithmetic means \pm S.D.) of ritonavir following administration of 100 mg Norvir® Tablet under fasted and high-fat fed state.

PK Parameters	Observed Fasted	Simulated Fasted	Observed High-Fat Fed	Simulated High-Fat Fed	AAFE
C_{max} ($\mu\text{g/mL}$)	0.6 ± 0.31	$0.49 \pm 0.30^{***}$	0.44 ± 0.21	$0.34 \pm 0.25^{***}$	1.25
T_{max} (h)	3.2 ± 1.2	$3.43 \pm 1.18^{***}$	4.8 ± 1.1	$4.75 \pm 1.37^{***}$	1.04
AUC_{0-t} ($\mu\text{g/mL}\cdot\text{h}$)	4.6 ± 2.0	$4.46 \pm 3.27^{***}$	3.5 ± 1.6	$3.30 \pm 2.86^{***}$	1.05

Ng *et al*, Journal of the International AIDS Society 2008, 11(Suppl 1):P247

Prediction of Negative Food Effects



Two factors were found to explain the observed negative food effect for ritonavir Norvir tablet –

A. Increased bile salt concentrations in fed state resulted in decrease in the free fraction of drug available as more drug is partitioned into bile salt micelles resulting in decreased Peff particularly in the duodenum and Jejunum I.

B. Decrease in effective diffusion coefficient of the drug in the fed state owing to increase in luminal viscosity – effect of dissolution rate

Comparison of Various Models

5 models were evaluated across 32 studies resulting in 160 workspaces

- **Model 1** – SV-Ritonavir 17 Simcyp File – First Order File
- **Model 2** – SV-Ritonavir 17 Simcyp File – Two First Order File with modified fa for fasted and fed state and Competitive Ki inhibition parameters of CYP3A4/5
- **Model 3** – ADAM Model with BD Sup ISEF Correction of V_{max} for CYP3A4/5
- **Model 4** – First order file with Fa = 0.5 (ADAM calculated) with BD Sup ISEF Correction of V_{max} for CYP3A4/5
- **Model 5** – ADAM Model with Competitive Ki inhibition parameters of CYP3A4/5

	AAFE (PK Parameters)		AAFE (DDI)	
	AUC	Cmax	AUC Ratio	Cmax Ratio
Model 1	1.32	1.38	1.83	1.42
Model 2	1.29	1.44	1.64	1.36
Model 3	1.50	1.49	1.79	1.45
Model 4	1.37	1.36	1.72	1.42
Model 5	1.49	1.53	1.52	1.31
Model 5 (with study matched formulation)	1.09	1.20	1.41	1.31
Model 3 (with study matched formulation)	1.09	1.20	1.64	1.45

AAFE – Absolute Average Fold Error
Model marked in green is the final model

Verification of systemic exposure of ritonavir following per oral administration of Norvir formulations

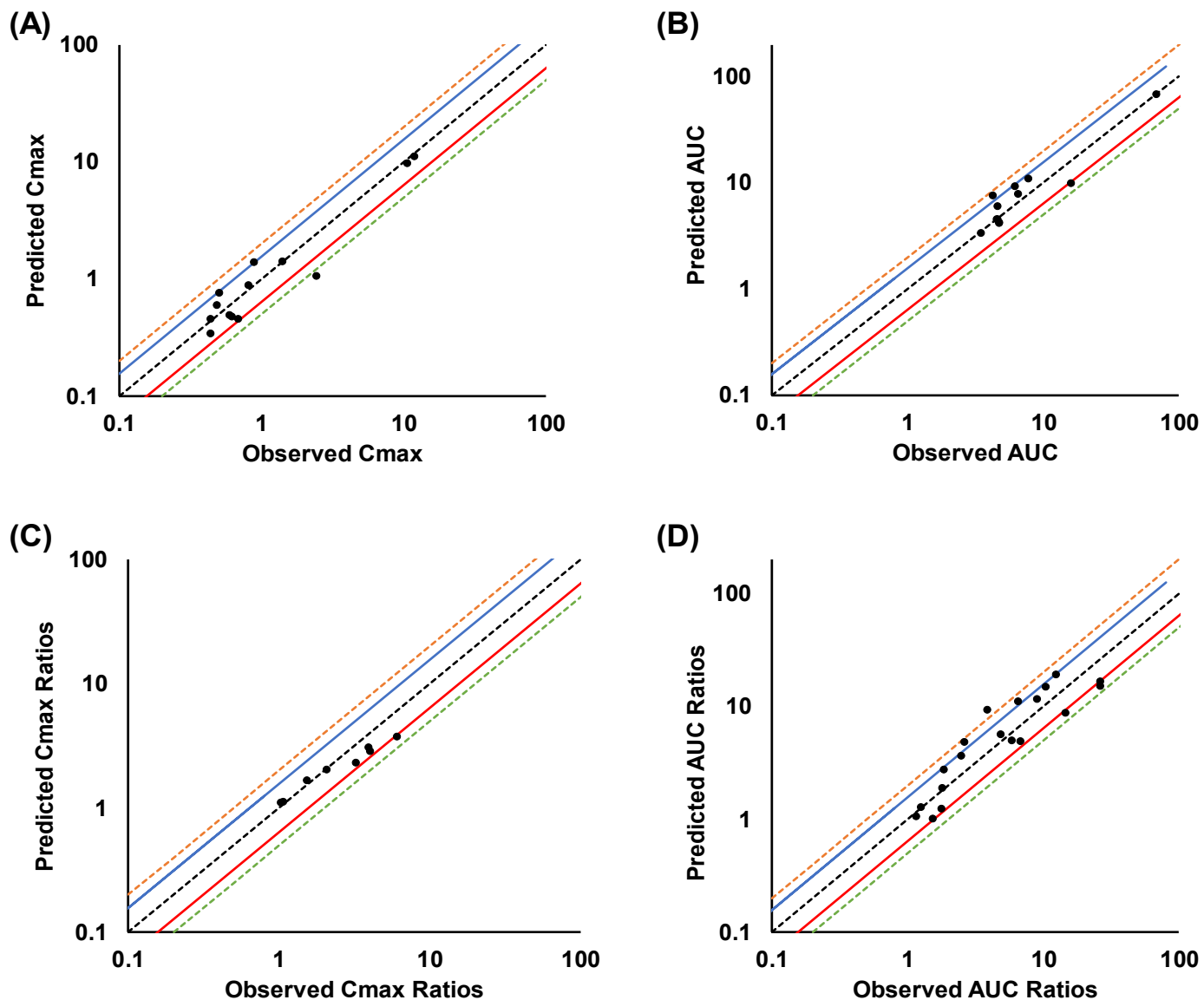
Norvir Capsule and Solution Formulation were simulated with 'Solution with Precipitation' Option

Reference	RTV Formulation	Simulation Option	C _{max} (µg/mL)		AUC _{(0-t)/(0-∞)} (µg.h/mL)	
			Observed	Predicted	Observed	Predicted
Ng et al 2008	IR Tablet 100 mg	Immediate Release	0.6 ± 0.31	0.49 ± 0.30	4.6 ± 2.0	4.46 ± 3.27
Ng et al 2008	IR Tablet 100 mg	Immediate Release	0.44 ± 0.21	0.34 ± 0.25	3.5 ± 1.6	3.30 ± 2.86
NDA Application 20-945 – Capsule	Soft Elastic Capsule 6x100mg	Solution	11.98 ± 3.33	10.99 ± 3.73	108.1 ± 33	96.56 ± 44.60
Liu 2007	Capsule 400 mg BID	Solution	10.7 ± 3.19	9.57 ± 3.51	68 ± 21	67.09 ± 31.57
Liu 2007	Capsule 100 mg BID	Solution with Precipitation	1.41 ± 0.72	1.39 ± 0.63	7.81 ± 3.87	10.79 ± 5.86
Teng 2013	Soft Elastic Capsule 100 mg SD	Solution with Precipitation	0.51 ± 0.27	0.76 ± 0.42	4.29 ± 3.07	7.51 ± 4.85
Mathias 2009	Solution 100 mg SD	Solution with Precipitation	0.81 ± 0.27	0.88 ± 0.39	6.53 ± 1.76	7.70 ± 4.23
Mathias 2009	Solution 200 mg SD	Solution with Precipitation	2.46 ± 0.27	1.05 ± 0.5	16 ± 7	9.8 ± 5.53
Brennan 2013	IR 100mg SD	Immediate Release	0.69 ± 0.31	0.45 ± 0.29	4.55 ± 1.72	4.48 ± 3.61
Brennan 2015	IR 100mg SD	Immediate Release	0.44	0.45 ± 0.29	4.74	4.22 ± 3.54
Morris 2012	Capsule 100 mg SD	Solution with Precipitation	0.49 ± 0.39	0.59 ± 0.35	4.63 ± 2.89	5.98 ± 4.58
Morcosa 2014	IR 100 mg SD	Immediate Release	0.62	0.47 ± 0.31	4.74	4.13 ± 3.30
Aarnouste 2005	Capsule 100mg BID	Solution with Precipitation	0.89	1.38 ± 0.64	6.2	9.21

DDI Verification of Ritonavir file as perpetrator of CYP3A4/5 and CYP2D6 substrate

Reference	RTV Formulation	Simulation Formulation	Cmax Ratios		AUC _(0-t) Ratios	
			Observed	Predicted	Observed	Predicted
Eichbaum et al. 2013	Solution 0.1 mg	Solution with Precipitation	Not reported	NA	1.55	1.02 ± 0.0
Eichbaum et al. 2013	Solution 0.3 mg	Solution with Precipitation	Not reported	NA	1.17	1.07 ± 0.03
Eichbaum et al. 2013	Solution 1 mg	Solution with Precipitation	Not reported	NA	1.79	1.24 ± 0.12
Eichbaum et al. 2013	Solution 3 mg	Solution with Precipitation	Not reported	NA	1.82	1.89 ± 0.43
Eichbaum et al. 2013	Solution 10 mg	Solution with Precipitation	Not reported	NA	2.63	4.83 ± 2.27
Eichbaum et al. 2013	Solution 30 mg	Solution with Precipitation	Not reported	NA	3.89	9.38 ± 7.58
Eichbaum et al. 2013	Solution 100 mg	Solution with Precipitation	Not reported	NA	6.51	11.05 ± 9.44
Eichbaum et al. 2013	Solution 300 mg	Solution with Precipitation	Not reported	NA	9.01	11.68 ± 10.14
Mathias 2009	Solution 100 mg SD	Solution with Precipitation	Not reported	NA	6.81	4.90 ± 1.67
Mathias 2009	Solution 200 mg SD	Solution with Precipitation	Not reported	NA	4.90	5.70 ± 1.92
Ancrenaz et al. 2013	IR Tablet	IR	6.10	3.76 ± 1.78	26.5	16.67 ± 12.35
Greenblatt 2009	Solution	Solution with Precipitation	3.96	3.10 ± 1.28	26.41	15.08 ± 9.61
Leri 2013	Solution	Solution with Precipitation	2.09	2.03 ± 0.46	5.9	5.04 ± 1.97
Leri 2013	Solution	Solution with Precipitation	3.26	2.31 ± 0.63	14.7	8.79 ± 4.08
Mathias 2010	Capsule	Solution with Precipitation	4.03	2.87 ± 1.16	12.5	19.22 ± 11.21
Kirby 2011	IR tablet	Immediate Release	Not reported	NA	10.5(8.7-12.7)*	14.9 (13.41-16.55)
Greenblatt 2000	Capsule	Solution with Precipitation	1.04	1.10 ± 0.05	2.48	3.64 ± 1.25
Aarnouste 2005	Capsule 100 mg BID	Solution with Precipitation	1.08	1.13 ± 0.06	1.26	1.29 ± 0.12
Ouellet 1998	Solution 200 mg TID	Solution with Precipitation	1.54	1.68 ± 0.64	1.86	2.77 ± 2.28

Model Performance Summary and Conclusions



- Mechanism-based modelling of *in vitro* biopharmaceutics experiments helped to build confidence in the quality of the key input parameters which improved the predictivity of the developed PBPK model.
- The present study highlights the importance of inclusion of formulation properties within PBPK framework which significantly improved the prediction ability of the developed model both for ritonavir systemic exposure as well as DDI risk assessment.
- **The compound file is available in Simcyp V19**

Predicted and Observed Cmax (A) and AUC (B) of systemic exposure of ritonavir reported for different formulations following oral administration. Predicted and Observed Cmax ratios (C) and AUC ratios (D) of CYP3A4 (Midazolam, Alprazolam) and CYP2D6 (Desipramine, Clarithromycin) substrates with and without ritonavir administration. All data expressed as mean (in case of systemic exposure parameters) and mean ratios (DDI liability) plotted in logarithmic x and y-axis. Black dash line represents the line of unity. Orange and Green Dashed lines represent two –fold prediction space. Solid blue and red lines represent the bioequivalence (0.8-1.25) limits.

Limitations and Gaps

1. Disintegration rate of the Norvir Tablet in the fasted state is assumed to be the same in the fed state due to lack of *in vitro* data.
2. Use of static viscosity model – more research is needed in this area to parameterize and verify the dynamic viscosity model particularly time-dependent viscosity changes following administration of food in the small intestine.
3. Further experimental exploration of additional metabolic pathways which could enable bottom up predictions of ritonavir clearance *in vivo*.

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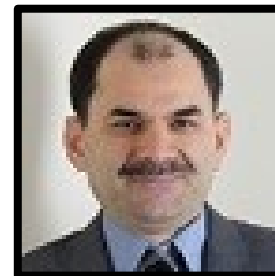
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Thank you for
listening 😊

Questions?

