## FDA U.S. FOOD & DRUG ADMINISTRATION



## Introduction

Liposomes have been used for decades as carriers of active pharmaceutical ingredients (API) resulting in enhanced drug efficacy and reduced toxicity. Phospholipids are the principal excipients in the liposomal formulations and are susceptible to hydrolysis, generating lipid degradation products such as lysophospholipids and free fatty acids. The lipid hydrolysis may alter the physical and chemical properties of the lipid bilayer, potentially affecting the drug release profiles of liposomal formulations.

## Objective

Available GC-FID and HPLC-based methods provide relatively poor selectivity and sensitivity. Therefore, we developed a rapid, sensitive and reproducible LC-QTOF based method to quantitate the phospholipid hydrolysis products, primarily lysophosphatidylcholines (LysoPCs), lysophosphatidylglycerol (LPGs) and free fatty acids (FFA) in liposomal formulations.

## Lipid Hydrolysis



Cross section of a liposome

\*Adapted from Bitounis, D., Fanciullino, R., Iliadis, A., and Ciccolini, J. (2012) Optimizing Druggability through Liposomal Formulations: New Approaches to an Old Concept, ISRN pharmaceutics 2012, 738432, http://syromonoed.com/lecithin-common-phospholipid-human-bodies **Figure 01.** Schematic representation of presence of phospholipids in liposome

### **Scheme 01.** Hydrolysis of Phosphotidylcholines (PCs) and Phosphotidylglycerols (PGs)

## Scheme 02. Schematic representation of acyl migration reaction of lysophosphatidylcholine





Spontaneous intramolecular acyl migration reaction under physiological condition can take place by yielding sn1 and sn2 regioisomers in a 9:1 ratio. The process of the formation of such regioisomer is called intra-molecular acyl migration.<sup>1</sup>

## **Table 01.** Different Types of Phospholipids Present in Liposomal Formulations<sup>2</sup>

Liposomal Formulation	Active Ingridient	Phospholipid in Fo
Doxil® (1995)	Doxorubicin	HSPC, PEG 2000-DS
DaunoXome <sup>®</sup> (1996)	Daunorubicin	DSPC
Depocyt <sup>®</sup> (1999)	Cytarabine/Ara-C	DOPC, DPPG
Myocet <sup>®</sup> (2000)	Doxorubicin	EPC
Mepact <sup>®</sup> (2004)	Mifamurtide	DOPS, POPC
Marqibo <sup>®</sup> (2012)	Vincristine	SM
Onivyde™ (2015)	Irinotecan	DSPC, MPEG-2000
Abelcet® (1995)	Amphotericin B	DMPC, DMPG
Ambisome <sup>®</sup> (1997)	Amphotericin B	HSPC, DSPG
Visudyne® (2000)	Verteporphin	DMPC, EPG
DepoDur™ (2004)	Epidural Morphine sulfate	DOPC, DPPG
Exparel <sup>®</sup> (2011)	Bupivacaine	DEPC, DPPG

\*HSPC (hydrogenated soy phosphatidylcholine); PEG (polyethylene glycol); DSPE (distearoyl-sn-glycero-phosphoethanolamine); DSPC (distearoylphosphatidylcholine); DOPC (dioleoylphosphatidylcholine); DPPG (dipalmitoylphosphatidylglycerol); EPC (egg phosphatidylcholine); DOPS (dioleoylphosphatidylserine); POPC (palmitoyloleoylphosphatidylcholine); SM (sphingomyelin); MPEG (methoxy polyethylene glycol); DMPC (dimyristoyl phosphatidylcholine); DMPG (dimyristoyl phosphatidylglycerol); DSPG (distearoylphosphatidylglycerol); DEPC (dierucoylphosphatidylcholine); DOPE (dioleoly-sn-glycero-phophoethanolamine)

## Quantification of Lysophosphatidylcholines (LPCs) and Free Fatty Acids (FFAs) in Parenteral Liposomal Formulations by Liquid Chromatography – Mass Spectrometry (LC-MS)

# <sup>b</sup> Office of Research and Standards, Office of Generic Drugs, Center for Drug Evaluation and Research, U.S. FDA, Silver Spring, MD 20993, United States

Phospholipid molecule



		Method Work	flow and	
Table (	Table 02. Method Information for FFA, LPC, and LPG			
Analyte	Column Type	Specifications	Mobile Phase A	
FFA	ACQUITY UPLC® CSH™ C8 column	100 X 2.1 mm; particle size 1.7 μm, Waters	100% H <sub>2</sub> O 1 mM Ammonium Aceta	
LPC	ACQUITY UPLC <sup>®</sup> CSH™ C18 column	100 X 2.1 mm; particle size 1.8 μm, Waters	100% H <sub>2</sub> O + 0.1 F 10 mM Ammoniu Formate	
LPG	ACQUITY UPLC <sup>®</sup> CSH™ C18 column	100 X 2.1 mm; particle size 1.8 μm, Waters	100% H <sub>2</sub> O 1 mM Ammonium Form	



**Optimization Determination of LPGs** LPG 18:0 350000 - Palmitic acid-d **Mobile Phase** - Palmitic acid y = 2419.1x - 8058.3 800000 - Stearic acid  $R^2 = 0.999$ ACN:MEOH 8:2 Internal standared (Lyso 16:0, 200pt ate 1 mM Ammonium LPG present in PF1 is Acetate 26.86 μg/g per DSPG 400000 FA + MEOH + 1.2 Time (min) um 0.1 % FA+10 mM 200000 Ammonium Formate MEOH \_\_\_\_ LPC18:0 LysoPG 18:0 (100 ppb) nate 1 mM Ammonium Formate 200 300 Time (min) Concentration (ppb) Figure 07. Chromatograms for LPG Figure 06. Calibration plot for LPG Figure 02. Chromatogram for FFA (A) and LPCs (B) 4 Time (min) **Analysis of Bulk Formulation Analysis of Phospholipid's Degradation Products** Figure 09: Chromatogram Figure 08: Chromatogram displaying presence of FFA displaying presence of LPC 18:0 and 16:0 in different 18:0 and 16:0 in different liposomal formulations liposomal formulations y = 21106x + 420590 $R^2 = 0.9967$ 800000 = 18354x + 25126  $R^2 = 0.999$ LPC 18:0 600000 16:0 4000000 FFA 16:0 FFA 16:0 • FFA 18:0 • FFA 18:0 200 300 Concentration (ppb) FFA 16:0 FFA 18:0 1.0 Time (min) **Table 05.** Detected LPCs and FFAs in different liposomal formulations as mass percentage Liposomal LPC Intra-day Measured Values <sup>n</sup> Formulation Mean SD Accuracy Precision recision (%) (%) ng/mL HSPC PF1 105.7 2.9 21.1 0.6 PF2 3.6 121.3 2.2 97.1 1.8 89.5 335.5 3.9 1.2 PF3 22.3 0.5 111.4 2.4 12.7 PF4 97.7 0.9 122.1 2.0 1.1 5.2 353.0 6.6 94.1 1.9 PF5 PF6 PF7 \*with respect to HSPC, \*\*with respect to total content of HSPC and DSPG 18000000 - y = 42919x - 83186 = 56613x + 95273 = 40314x - 126625 18000000 R<sup>2</sup> = 0.9997 R<sup>2</sup> = 0.9991 R<sup>2</sup> = 0.9997 Summary and Conclusion 1600000 = 44554x - 1839 16000000 = 62564x + 18539 = 46325x - 10839 14000000 ·  $R^2 = 0.9992$ 20000000 - $R^2 = 0.998$ R<sup>2</sup> = 0.9995 14000000 -LC-MS methods were developed and validated for the quantitation of lipid degradation products including FFAs, 12000000 12000000 LPCs, and LPGs in liposomal pharmaceutical formulations (PFs). 15000000 10000000 The limit of detection (LODs) for FFA 16:0, FFA 18:0, LPC 16:0 and LPC 18:0 are 1.70 ng/mL, 1.03 ng/mL, 2.0 8000000 8000000 1000000 600000 ng/mL, and 2.1 ng/mL, respectively. 6000000 400000 4000000 LPC 16:0 ✤ The limit of quantification (LOQs) for FFA 16:0, FFA 18:0, LPC 16:0, and LPC 18:0 are 5 ng/mL, 5 ng/mL, 6.5 LPC 16:0 LPC 16:0LPC 18:0 • LPC 18:0 • LPC 18:0 2000000 ng/mL, and 7.0 ng/mL, respectively. 300 400 200 The LOD and LOQ for LPG are 2.1 ng/mL and 7.1 ng/mL, respectively. 400 Concentration (ppb) Concentration (ppb) Concentration (ppb) Corresponding lipid degradation products (FFA and lyso-lipids) of lipid excipients were detected for all the liposomal formulations. References Laton dour N/a Intra-day Measured Values <sup>r</sup> [1] Plueckthun, A., and Dennis, E. A. (1982) Acyl and phosphoryl migration in lysophospholipids: importance in phospholipid synthesis Accuracy Precision recision Mean SD and phospholipase specificity, *Biochemistry 21*, 1743-1750. (%) (ng/mL) (%) (%) <u>+</u> [2] Bulbake, U., Doppalapudi, S., Kommineni, N., and Khan, W. (2017) Liposomal Formulations in Clinical Use: An Updated Review, Pharmaceutics 9 96.3 19.3 0.2 4.1 1.1 Acknowledgements 103.0 0.6 231.8 2.1 0.9 These studies were conducted at the NCTR/ORA Nanotechnology Core Facility (NanoCore) located on the U.S. Food & Drug Administration's 375.4 100.1 4.2 1.1 Jefferson Laboratories Campus (Arkansas). Dr. Dumindika A. Siriwardane and Dr. Siyam Ansar are ORISE fellows and this project was supported in part by an appointment to the Research Participation Program at the U.S. Food and Drug Administration administered by the 19.6 97.9 0.2 1.3 1.6 Oak Ridge Institute for Science and Education (ORISE) through an interagency agreement between the U.S. Department of Energy and the 235.0 104.5 1.9 0.8 U.S. Food and Drug Administration. Authors would like to thank the Office of Research Coordination and Evaluation (ORCE), Center for Drug Evaluation Research (CDER), ORA and Arkansas Laboratory for their support. We thank Dr. Michael Wichman, Dr. Paul Howard, Dr. Darby 371.8 99.1 3.5 0.9 0.9 Kozak, Dr. Lei Zhang, Dr. Marylin Khanna, Dr. Sean Linder and Dr. Selen Stromgren for their comments and suggestions.

endorsement of a product or manufacturer

Inter-day Measured Values <sup>m</sup>					
	QC sample	Mean	SD	Accuracy	Pı
	concentration	Concentration	±	(%)	
	(ng/mL)	(ng/mL)			
FFA	20	20.2	1.8	105	
16:0	125	126.7	7.5	112	
	375	356.5	7.6	108	
FFA	20	19.8	2.5	98.6	
18:0	125	124.0	11.1	102	
	375	366.2	8.1	98	



**Determination of FFAs Figure 04.** Calibration plots for FFA for three consecutive days 
**Table 03.** Method validation statistics for FFAs
 **Determination of LPCs Figure 05.** Calibration plots for LPC for three consecutive days Table 04. Method validation statistics for LPCs

Inter-day Measured Values "					
	QC sample concentration (ng/mL)	Mean Concentration (ng/mL)	SD ±	Accuracy (%)	F
LPC 16:0	20	19.6	0.8	98.2	
	225	225.0	1.4	100.0	
	375	374.1	3.1	99.8	
LPC 18:0	20	19.4	0.3	96.0	
	225	226.1	1.7	100.5	
	375	370.8	3.4	98.9	

The views expressed in this poster are those of the researchers and should not be interpreted as the official opinion or policy of the U.S. Food and Drug Administration, Department of Health and Human Services, or any other agency or component of the U.S. government. The mention of trades names, commercial products, or organizations is for clarification of the methods used and should not be interpreted as an





C <b>16:0</b> %)*	LPC 18:0 (%)*	FFA 16:0 (%)*	FFA 18:0 (%)*	
).00	0.00	0.00	0.00	
0.14	0.59**	0.13	1.14**	
0.04	0.05	0.15	0.00	
0.10	0.85	0.07	1.16	
0.13	1.02	0.08	1.35	
).27	2.16	0.01	2.87	
).44	4.20	0.02	4.70	
).19	1.51	0.16	2.22	