## 01-02

### Investigation of Pharmacokinetic Sensitivity to Regional Lung Deposition of **Locally-Acting Orally Inhaled Drug Products**

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### PURPOSE

The current thinking for demonstrating bioequivalence (BE) of orally inhaled drug products (OIDPs) is based on an aggregate weight-of-evidence approach - recommendations for equivalence of (i) in vitro drug delivery performance, (ii) *in vivo* systemic exposure, and (iii) *in vivo* drug delivery to the local sites of action in the lungs, in addition to qualitative and quantitative formulation sameness (same inactive ingredients within 95-105% of the reference product concentration) and device similarity.<sup>1</sup> The main challenge is establishing equivalence of drug delivery to the lungs; specifically, equivalence in the available dose, pulmonary residence time, and regional deposition of drug in the lungs.

Since there is limited understanding of how pharmacokinetics (PK) relates to drug concentrations in the lungs, and uncertainties with in vitro correlations to lung deposition and clinical efficacy, comparative pharmacodynamic (or clinical endpoint) BE studies are currently recommended to demonstrate equivalence at the sites of action. Hypothetically, PK studies can provide insights on the fate of drug in the lungs when the oral bioavailability of the drug is negligible (e.g., fluticasone propionate)<sup>2</sup> or prevented through charcoal co-treatment. Under such conditions, the area under the concentration-time curve (AUC) may indicate the dose available to the airways, while the peak concentration (Cmax) may be sensitive to the local deposition pattern (e.g., faster absorption (i.e., higher Cmax) suggests more peripheral deposition).<sup>3</sup>

This study evaluated the potential application of PK studies in assessing differences in local delivery of dry powder inhaler (DPI) formulations engineered to differ in central to peripheral lung deposition.

#### **METHODS**

Preparation of FP DPI formulations: The same batches of micronized fluticasone propionate (FP) and coarse lactose were used to prepare three DPI formulations (A, B, C) to target a similar fine particle dose (FPD), but different mass median aerodynamic diameters (MMAD) to achieve different regional (i.e., central to peripheral) deposition in the lungs; the amount and grade of lactose fines (different median volumetric particle size, D<sub>50</sub>) were varied. The following FP DPI formulation compositions were studied:

Component 9/ w/w	<b>FP DPI Formulation</b>			
Component, %w/w	Α	В	С	
Fluticasone propionate (FP, D <sub>50</sub> = 2.1 µm)	0.8	0.8	0.8	
Sieved lactose (SV003, D50 = 63.6 µm)	79.4	89.3	96.7	
Milled lactose (LH201, D <sub>50</sub> = 20-25 μm)	19.8	-	-	
Milled lactose (LH230, D <sub>50</sub> < 10 µm)	-	9.9	-	
Micro-fine lactose (LH300, D50 < 5 µm)	-	-	2.5	

In vitro characterization: The FP DPIs were filled into size 3 HPMC capsules to contain 100 µg FP in 12.4 mg of lactose and evaluated with the capsule-based Plastiape Monohaler 8<sup>®</sup> DPI.

Aerodynamic particle size distribution (APSD) was evaluated using the Next Generation Impactor (NGI) with USP induction port and pre-separator at a flow rate of 60 L/min. FPD < 3 and 5 µm were interpolated from cumulative aerosol size distributions using a Weibull function.

Realistic aerosol characterization was performed with anatomical mouth-throat (MT) models [Virginia Commonwealth University (VCU), Oropharyngeal Consortium 160

(OPC), Alberta Idealized Throat] and a realistic inhalation profile (Figure 1) to determine the *in vitro* total lung (ex-MT) dose for  $\frac{1}{2}$  120 each FP DPI formulation.

In vitro dissolution studies were performed using the UniDose apparatus<sup>4</sup> for sample collection (impactor-sized mass at 60  $\frac{1}{2}$  40 L/min) and a USP Apparatus V (paddle-over-disk) modified to house a 47 mm filter.

Pharmacokinetic study: A randomized, double-blind, singledose, 4-way crossover PK study (formulation C was replicated to Figure 1: Comparison of realistic IP with USP assess intra-subject variability) was performed in 24 healthy, profile. IP = inhalation profile adult volunteers. Two inhalations per capsule from 5 capsules Realistic IP (50th percentile of the IPs likely to be (500 µg FP) were performed to ensure complete dose delivery.

Blood samples were obtained pre-dose and up to 24 hours after the last inhalation. Non-compartmental PK analysis was performed using Phoenix WinNonlin software. Cmax, AUC0-last, and AUC0-last, and AUC0-inf (AUCinf was extrapolated) were dose normalized based on *in vitro* ex-MT determinations to adjust



for differences in the *in vivo* lung dose. ANOVA performed for statistical comparisons on log-scale with SAS software; pair-wise treatment comparisons using Bonferroni-adjusted p-values at a 5% significance level.

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# **Advancing Pharmaceutical Sciences,**

					CONCLUSIONS		
<b>Figure 2:</b> Impactor-sized mass deposition profiles for FP DPIs actuated into the NGI with a flow rate of 60 L/min. Circles are means [FP DPIs stored under long-term conditions ( $25^{\circ}C/60\%$ RH) for 12 months (n=3) and 20 months (n=3)] $\pm$ SD.					• Varying the amount and grade of lactose fines in FP DPIs, which impacted the MMADs (formulations B and C had comparable MMADs, which was different from formulation A), altered the <i>in vitro</i> APSD and dissolution performance, and <i>in vivo</i> PK parameters.		
	<ul> <li>The total amount of drug deposited on stages 2 and 3 were similar across the FP DPIs, while the mass deposited on stage 4 to micro-orifice collector (MOC) was lower for formulation A compared to formulations B and C (Refer to Table 1).</li> </ul>				<ul> <li>In vitro evaluation of the FP DPIs with comparable MIMADs (formulations B and C) were shown to have similar dissolution profiles and total ex-MT dose, while the FP DPI with a larger MMAD (formulation A) was shown to have a slower dissolution rate and lower total ex-MT dose.</li> </ul>		
	<b>Table 1:</b> Characteristics of FP DPIs differing in lactose fines. Values are mean [FP DPIs stored under long-term conditions ( $25^{\circ}C/60\%$ RH) for 12 months (n=3) and 20 months (n=3)] ± SD.				<ul> <li>The FP DPIs with comparable MMADs (formulations B and C) revealed similar PK profiles. Cmax was shown to be sensitive to</li> </ul>		
		FP	<b>DPI Formulat</b>	ion	differences in FP DPI attributes – formulation A (larger MMAD)		
	Parameter	Α	В	С	revealed a significantly lower Cmax. In addition, formulation C (smallest MMAD) was shown to have a significantly higher		
	Mass Median Aerodynamic Diameter (MMAD),	μm 4.5 ± 0.1	3.8 ± 0.1	$3.7 \pm 0.0$	AUC <sub>0-last</sub> compared to formulation A, indicating a weak trend that		
	Geometric Standard Deviation (GSD)	$1.9 \pm 0.0$	2.0 ± 0.1	2.1 ± 0.0	PK may help differentiate total and central to peripheral lung		
	<b>Fine Particle Dose (FPD) &lt; 5 μm</b> , μg	12.2 ± 1.0	18.7 ± 0.9	15.8 ± 0.9	deposition.		
5,	<b>Fine Particle Dose (FPD) &lt; 3 μm</b> , μg	$5.3 \pm 0.7$	10.0 ± 0.5	8.6 ± 0.6	PK may provide supportive information on pulmonary		
0	<b>Impactor-Sized Mass (ISM)</b> , μg Stage 2 to MOC (< 8.1 μm)	17.3 ± 1.2	23.8 ± 1.3	19.6 ± 1.1	available dose, the pulmonary residence time, and the regional deposition of drugs in the lungs). This could help generic drug		
0	Stages 2 and 3 (2.8-8.1 µm)	12.5 ± 0.7	14.4 ± 1.2	11.5 ± 0.6	development and BE evaluation of poorly soluble OIDPs.		
n	Stage 4 to MOC (< 2.8 µm)	$4.8 \pm 0.6$	$9.4 \pm 0.4$	8.2 ± 0.5			
<b>(</b> )	Normalization Factor (derived from ex-MT dose determinations)	1.00	1.32	1.21	REFERENCES		
<ul> <li>Figure 4: Plasma concentrations of FP DPIs following dose normalization based on ex-MT determinations. Symbols are means (n=24 subjects) ± SD.</li> <li>Formulation C</li> <li>Formulation C (replicate)</li> <li>Formulations B and C (similar MMAD) revealed similar PK profiles, indicating BE.</li> </ul>				<ul> <li>BA, Connor DP, et al. JAMPDD, 2010; 23(1): 1-29.</li> <li>Falcoz C, Oliver R, McDowall JE, Ventresca P, Bye A, Daley- Yates PT. Clin Pharmacokinet. 2000; 39(Suppl 1): 9-15.</li> <li>Hochhaus G, Horhata S, Hendeles L, Suarez S, Rebello J. <i>The AAPS Journal</i>. 2015; 17(3): 769-775.</li> <li>Price R, Farias G, Ganley W, Shur J. <i>RDD Asia 2018</i>. 2018; Volume 1: 265-276.</li> </ul>			
DX * C fo t nif	plot comparisons of key PK parameters: $C_{max}$ (a), AUCo-last Significantly different (p<0.05) from the estimate for formulation A was significantly lower (a). of formulation A ricantly lower than on C (b); a weak of more central of formulation A.	<ul> <li>Cmax of formulation indicating potent peripheral deposition B and C, but could slower dissolution.</li> <li>(b), and lation A.</li> <li>(b), and lation A.</li> </ul>	A (largest MMA ially higher on compared to also theoretics	AD) was lower central to o formulations ally be due to	<ul> <li>ACKNOULEDGEENELDS</li> <li>Junding for this work was supported by:</li> <li>U.S. Food and Drug Administration through contracts HHSF223201610099C, HHSF223201300479A, HHSF223201110117A, and HHSF223201000090C;</li> <li>U.S. Food and Drug Administration through grants 1001FD004950 and 1U01FD005231;</li> <li>National Center for Advancing Translational Sciences of the National Institute of Health through Award UL1TR001427.</li> <li>Views expressed in this poster are those of the authors and do not necessarily reflect the official policies of the Department of Health and Human Services, nor does any mention of trade names, commercial practices or organization imply endorsement by the United States Government.</li> </ul>		
nii So	Icant differences       Image: Second S	ormulation C Formulat	tion A Formulation B	Formulation C	UF College of Pharmacy UNIVERSITY of FLORIDA		

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