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## PURPOSE

- A comprehensive evaluation of qualitatively and quantitatively (Q1/Q2) equivalent ophthalmic ointments with manufacturing differences is challenging due to the complexity of these formulations.
- *In vitro* drug release testing and *ex vivo* transcorneal drug permeation may provide valuable information on the performance of Q1/Q2 equivalent ointments prior to any animal studies.
- Good correlation between *in vitro* and *ex vivo* drug release may be indicative of good *in vitro* and *in vivo* correlation. Accordingly, it may be useful to investigate *in vitro* as well as *ex vivo* drug release from Q1/Q2 equivalent ophthalmic ointments and evaluate any correlation between these release profiles.

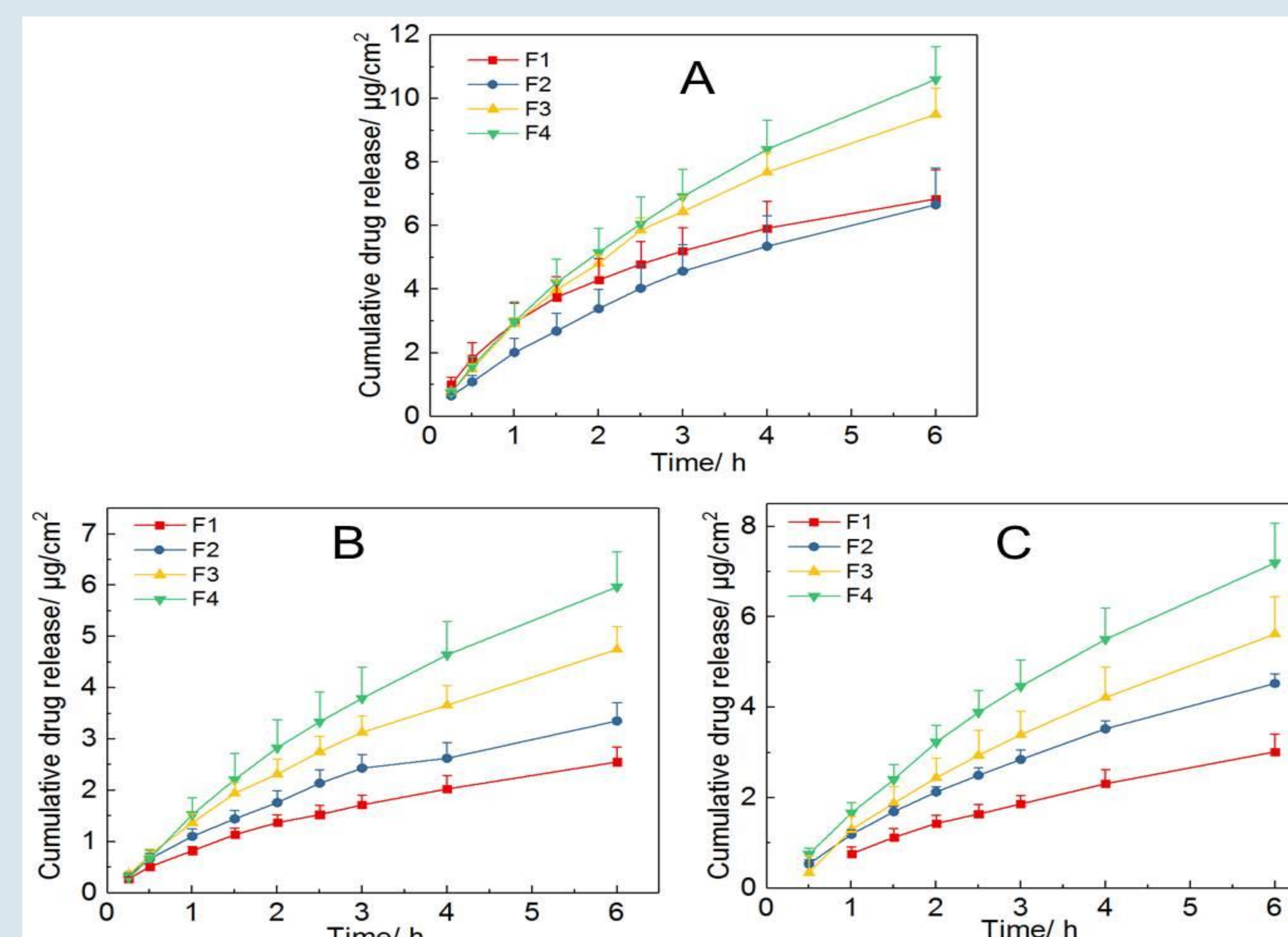
## METHODS

- Four Q1/Q2 equivalent loteprednol etabonate ointments were prepared using different processing methods and excipient sources (OWP from Fisher® and NWP from Fougera®).
- The *in vitro* drug release testing of the four ointment formulations were performed with pH 7.4 artificial tear fluid with 0.5% (w/v) SDS at 37°C using three different apparatus (Franz diffusion cells, USP apparatus 2 with enhancer cells and USP apparatus 4 with semisolid adapters). Three models (zero order, logarithmic and the Higuchi model) were used to study the release kinetics of the ointment formulations.
- The transcorneal permeation studies were performed with pH 7.4 artificial tear fluid with 9% (w/v) HP-beta-CD at 34°C using spherical joint Franz diffusion cells (area: 0.64 cm<sup>2</sup>). Fresh rabbit corneas were used and the experimental duration was 4 hours (n=6).

**Table 1.** Manufacturing methods for the Q1/Q2 equivalent loteprednol etabonate ophthalmic ointments.

Formulations	Manufacturing process	Petrolatum
F1	Hot melting with immediate cooling at -20°C	OWP
F2	Hot melting with cooling at room temperature	NWP
F3	Simple mixing at room temperature	OWP
F4	Simple mixing at room temperature	NWP

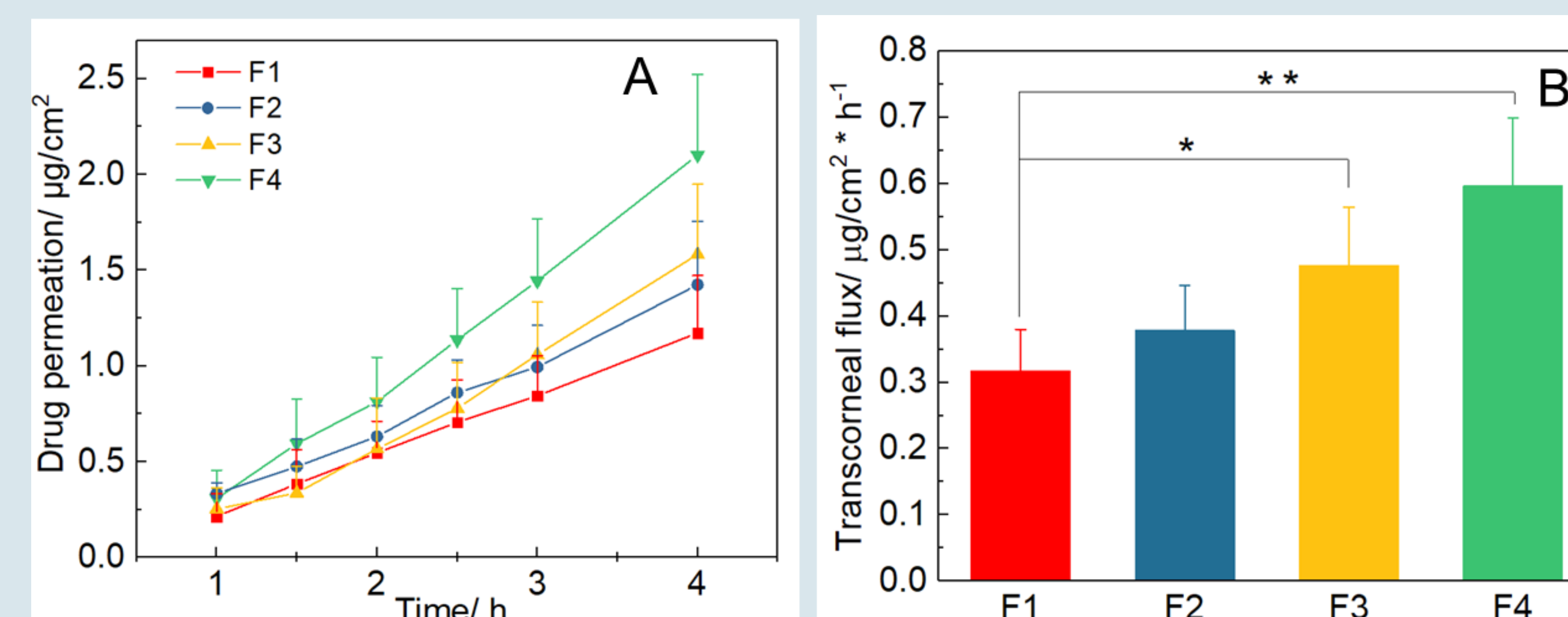
## RESULTS



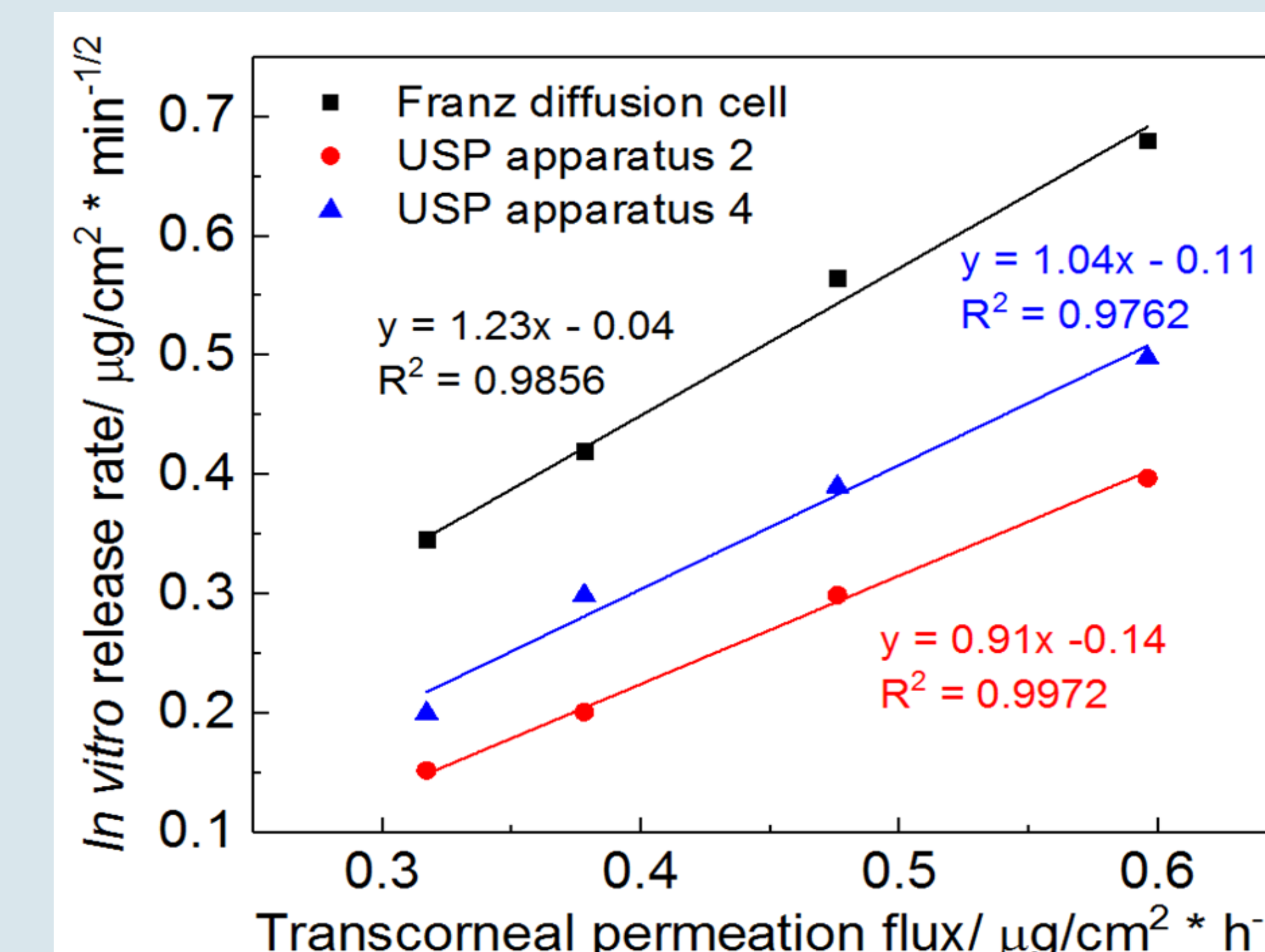
**Figure 1.** Release profiles of the loteprednol etabonate ointments obtained using: **A)** Franz diffusion cell method; **B)** USP apparatus 2 with enhancer cells; and **C)** USP apparatus 4 with semisolid adapters method (n = 6).

**Table 2.** Model fitting of release profiles of loteprednol etabonate ointments using USP app 4 with semisolid adapters (representative data, other methods had similar trends).

Formulation	Zero order model		Logarithmic model		Higuchi model	
	k <sub>0</sub> × 10 <sup>2</sup>	R <sup>2</sup>	k <sub>log</sub>	R <sup>2</sup>	k <sub>H</sub>	R <sup>2</sup>
F1	0.73 ± 0.08	0.983 ± 0.009	2.86 ± 0.28	0.978 ± 0.011	0.20 ± 0.02	0.985 ± 0.002
F2	1.10 ± 0.05	0.981 ± 0.006	4.28 ± 0.20	0.977 ± 0.008	0.30 ± 0.01	0.997 ± 0.008
F3	1.43 ± 0.18	0.987 ± 0.004	5.53 ± 0.37	0.978 ± 0.010	0.39 ± 0.05	0.983 ± 0.002
F4	1.83 ± 0.23	0.980 ± 0.004	7.14 ± 0.91	0.979 ± 0.008	0.50 ± 0.06	0.988 ± 0.002



**Figure 2.** **A)** Transcorneal permeation profiles; and **B)** Transcorneal flux of loteprednol etabonate ointments obtained using spherical joint Franz diffusion cells (n = 6).



**Figure 3.** Plot of *in vitro* release rate against *ex vivo* transcorneal permeation flux (n=6) for the four ointment formulations. Straight lines (R<sup>2</sup> > 0.98) were obtained for all three release methods.

## CONCLUSIONS

- The *in vitro* drug release profiles of Q1/Q2 equivalent ophthalmic ointments obtained using the three different release methods followed the Higuchi release kinetics.
- The compendial methods possessed better discriminatory capability compared to the non-compendial method.
- Strong correlation was established between the *in vitro* release rate and *ex vivo* release flux of the Q1/Q2 equivalent ointments with different manufacturing processes.

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