

# Development of *In Vitro-In Vivo* Correlation of Peptide Microspheres – Possibility and Challenges

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## PURPOSE

### *In vitro-in vivo* correlation (IVIVC):

- In vitro* response  $\longleftrightarrow$  **IVIVC**  $\longleftrightarrow$  *In vivo* response
- Predictive mathematical model
- Challenging for the microspheres – complex release characteristics

- However, developed Level A IVIVC (rabbit model) –



### Peptide Microspheres -

- Further explore the concept of IVIVC development
- Challenges:** Larger size, hydrophilic, high burst release –Variable

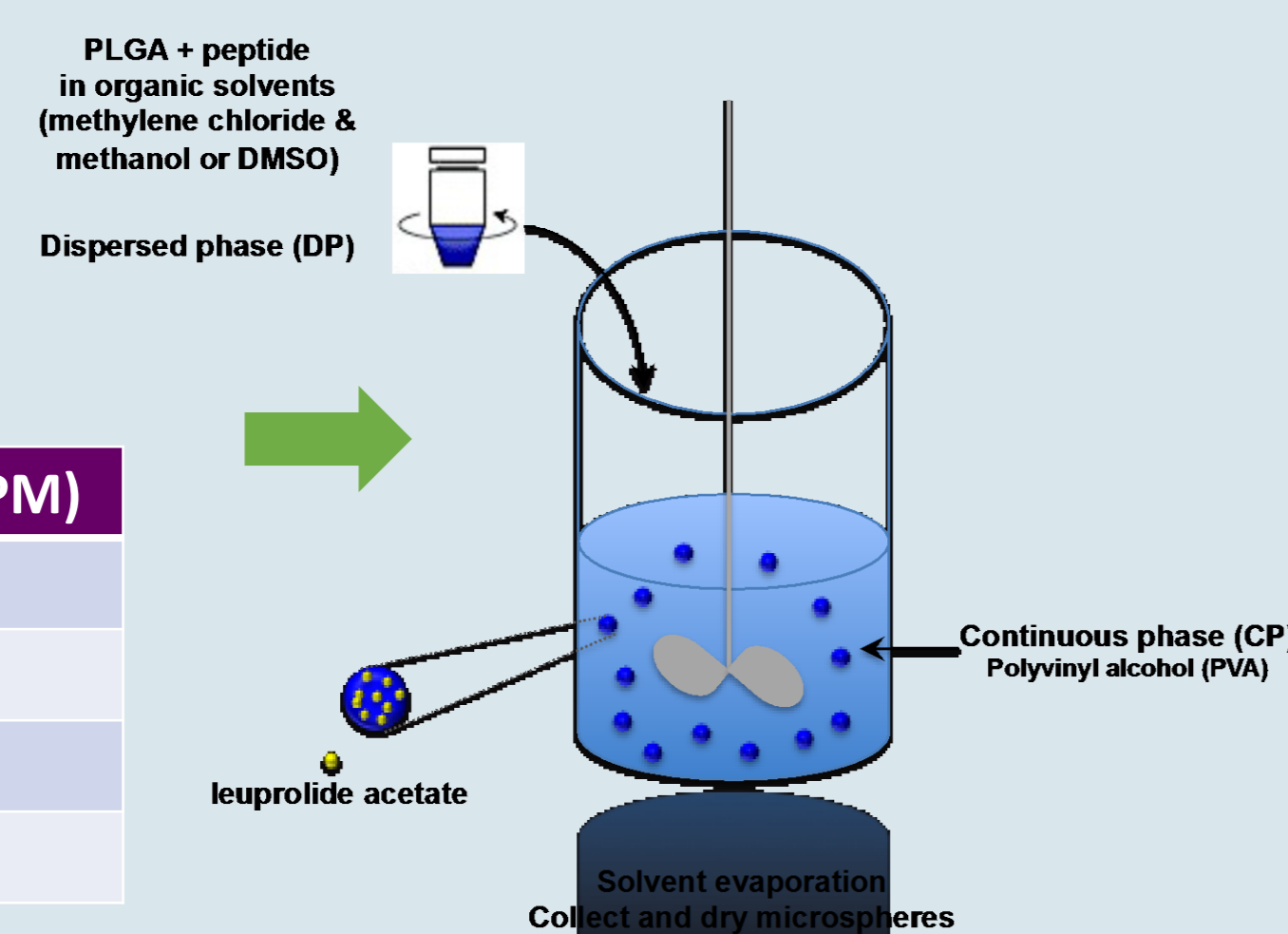
**OBJECTIVE:** To understand challenges involved and possibility of establishing level A IVIVCs (rabbit model) for peptide microsphere drug products.

## METHOD(S)

**Model Drug:** Leuprolide Acetate **Polymer:** Poly(lactic-co-glycolic acid) (PLGA)

### Preparation Method:

Single emulsion (O/W) solvent evaporation



### Process variables

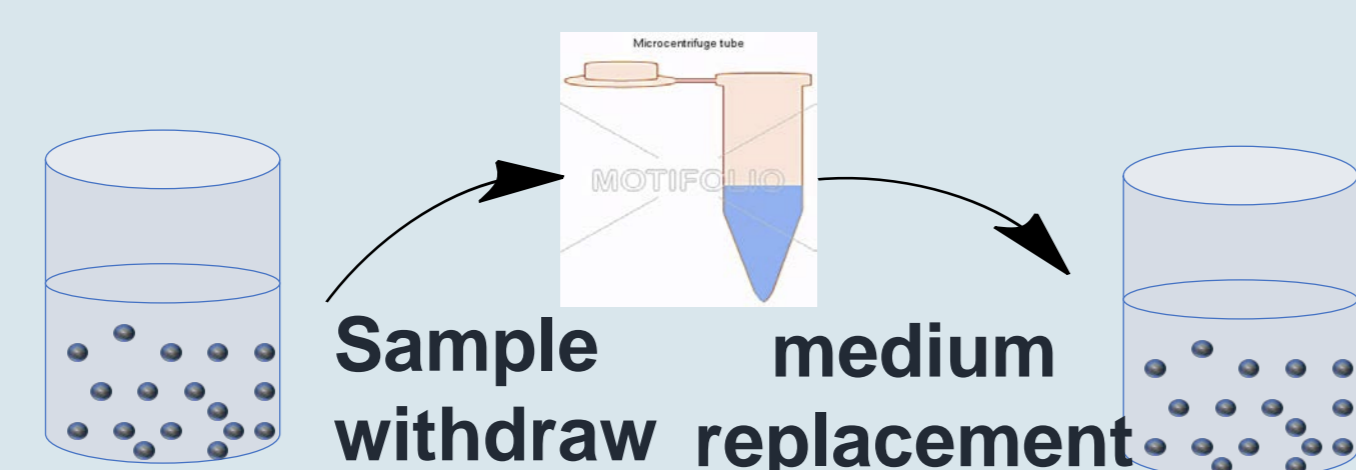
Formulations	Solvent systems	Homogenization Speed (RPM)
F1	DCM/MeOH	13 to 14 K
F2		8 to 9 k
F3	DCM/DMSO	13 to 14 K
F5		8 to 9 k

### Characterization of microspheres:

1. **Critical quality attributes:** Drug loading, particle size, porosity

### 2. *In Vitro* Release Testing:

- Sample-and-separate method
- Medium: 33 mM phosphate buffer, pH 7.4
- Testing Temperature: 37°C

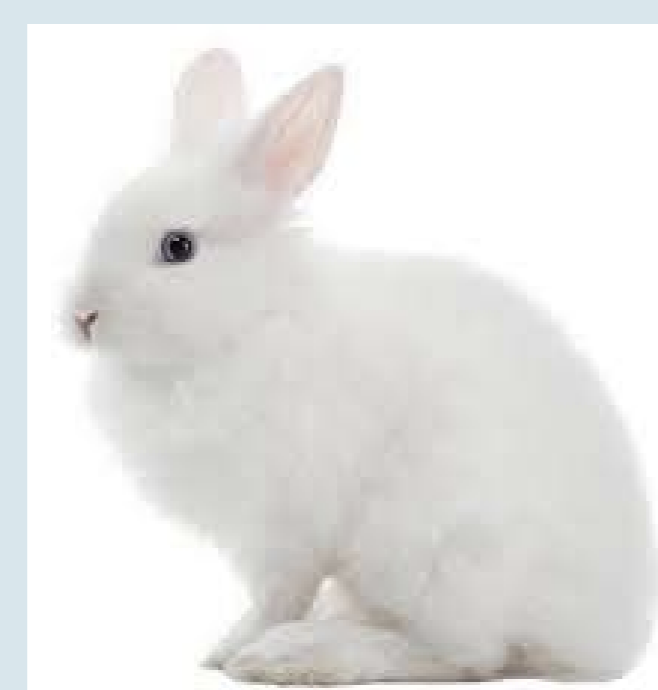


### 3. *In Vivo* Release Testing:

- Model: Rabbit
- Route: IM injection
- Blood Sample collection over the period of time

### 4. *In vitro-in vivo* correlation (IVIVC):

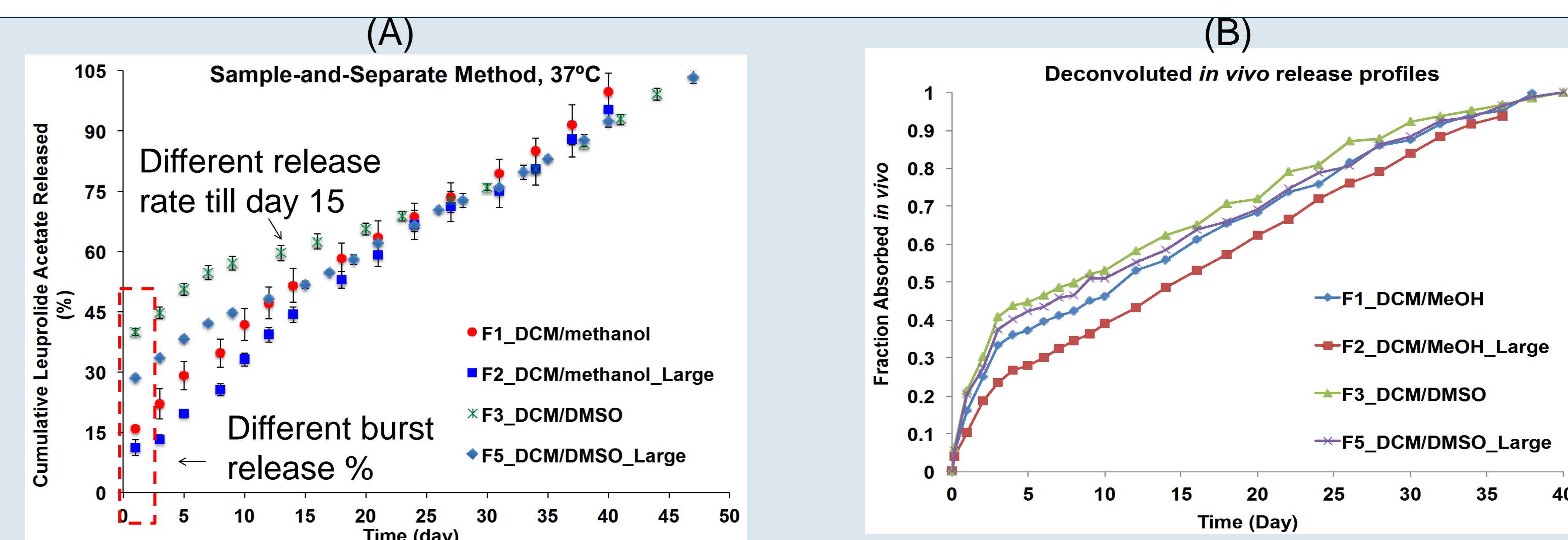
- 2- Stage deconvolution Approach (Loo-Riegelman method)
- Validation of the model: Internal as well as external
- Estimation of % Prediction Error (%PE)



## RESULT(S)

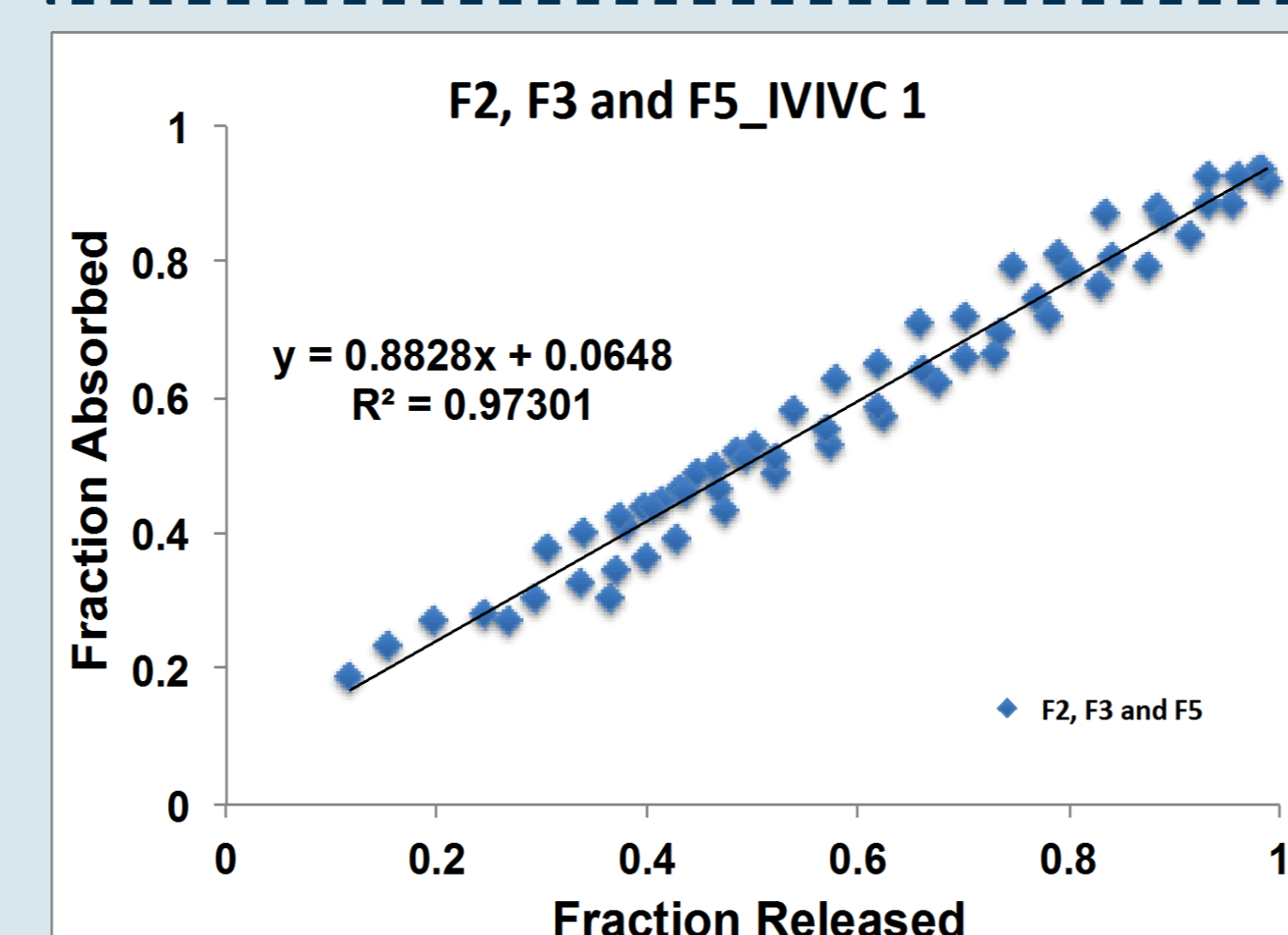
**Table 1.** Physicochemical properties of the prepared leuprolide acetate (LA) microspheres

Formulations	Drug Loading (%W/W)	Particle Size (µm)	Porosity (%)	Pore Diameter (nm)
F1	~ 8 %	45.52	57.06	814.5
F2		72.69	52.65	712.7
F3	<b>Q1/Q2</b>	40.71	61.01	964.0
F5		52.27	56.48	814.1



**Figure 1.** A) *In vitro* release profiles; and B) Deconvoluted *in vivo* release profiles of the prepared Q<sub>1</sub>/Q<sub>2</sub> equivalent LA microspheres.

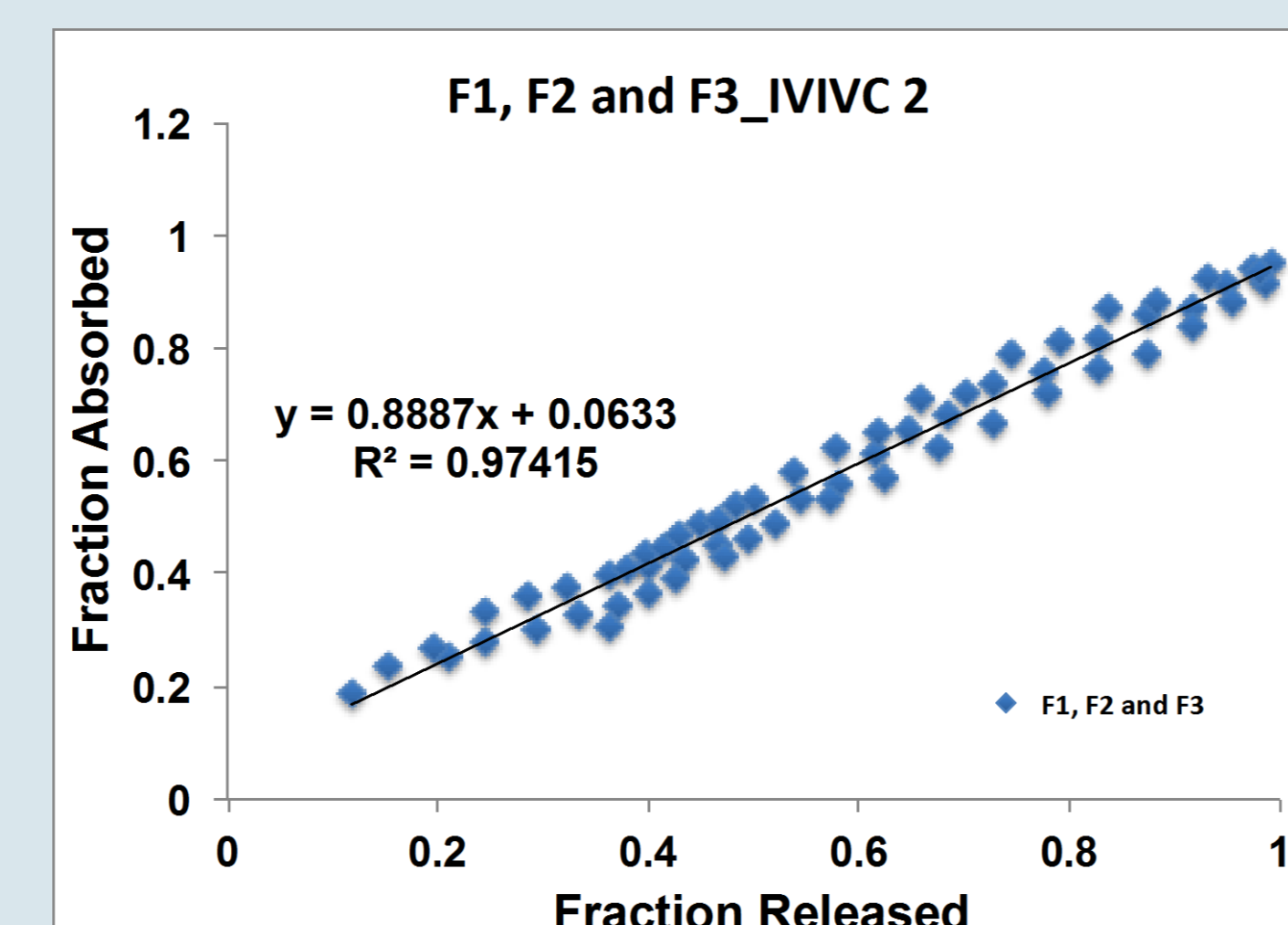
### Development of *in vitro-in vivo* correlation (IVIVC)



- 1:1 Linear Correlation – Level A**

$$T(\text{In vivo}) = (\text{Time}(\text{In vitro}) * a) + b$$

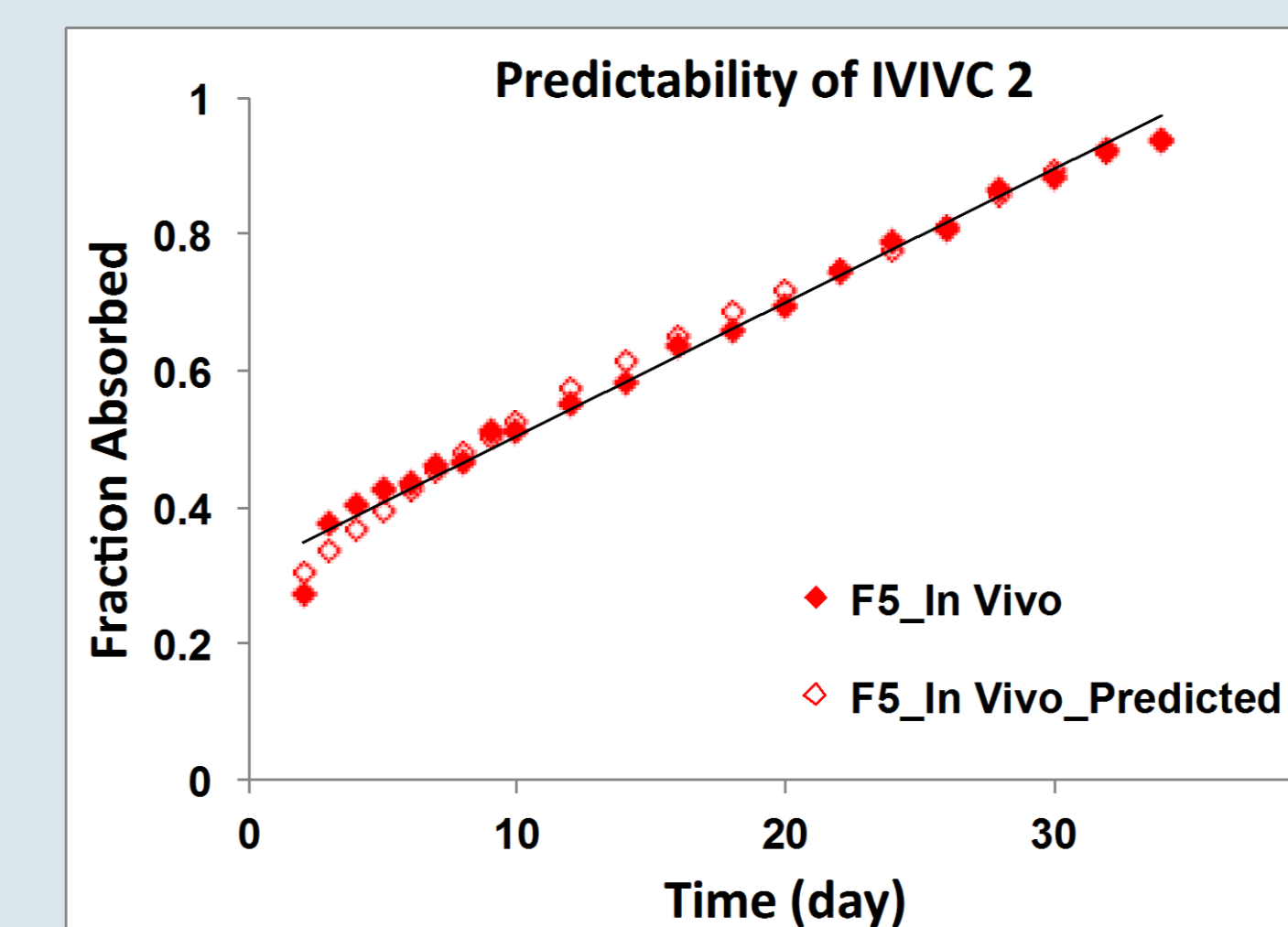
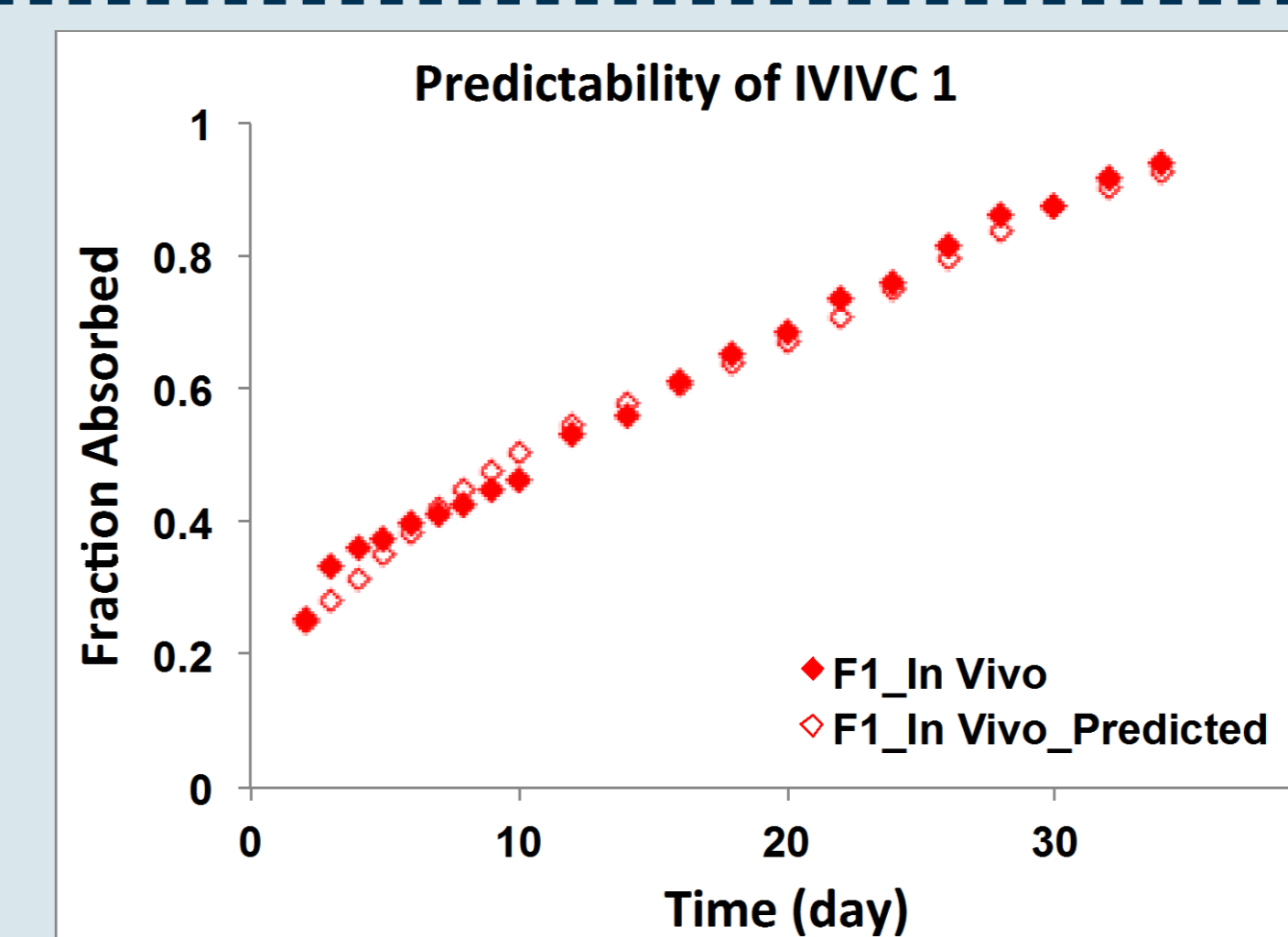
a=Time scaling factor = 1.6, b= time shifting factor = 1



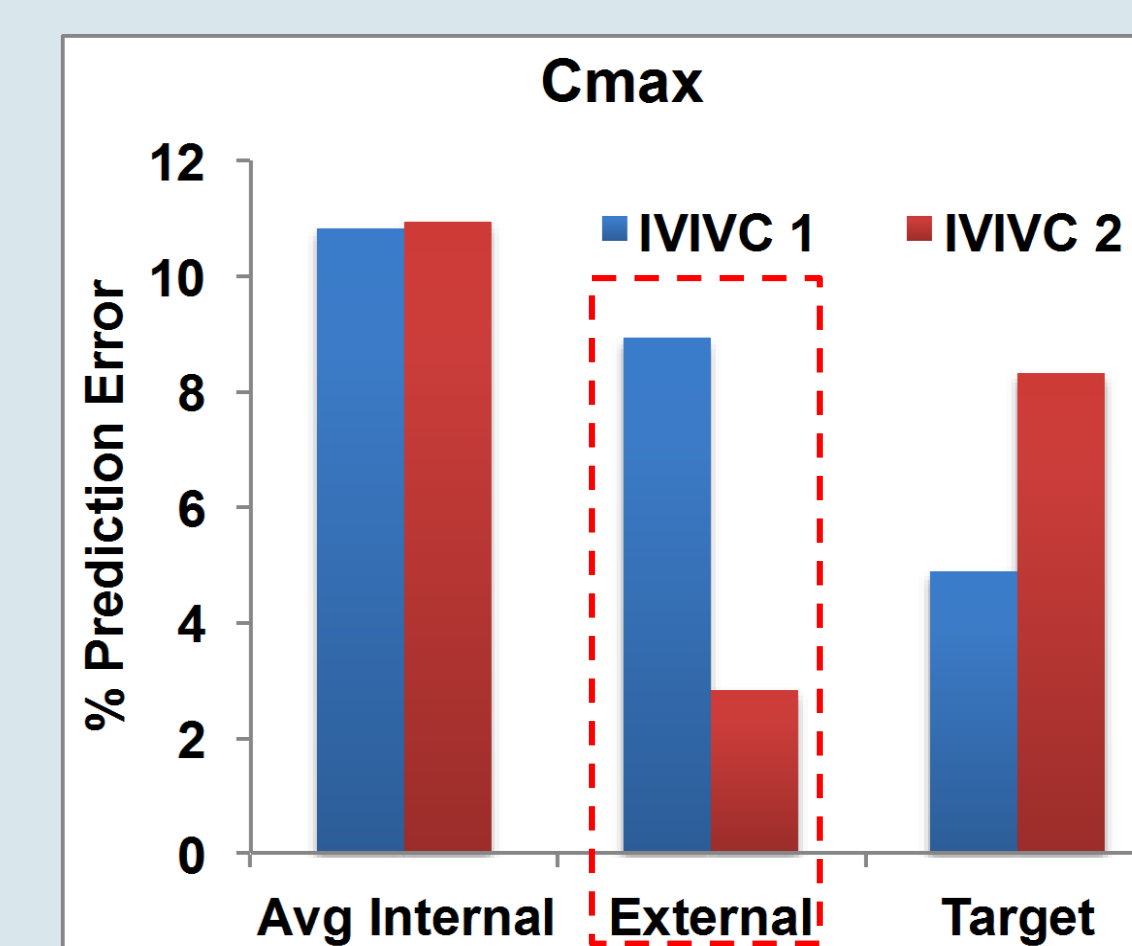
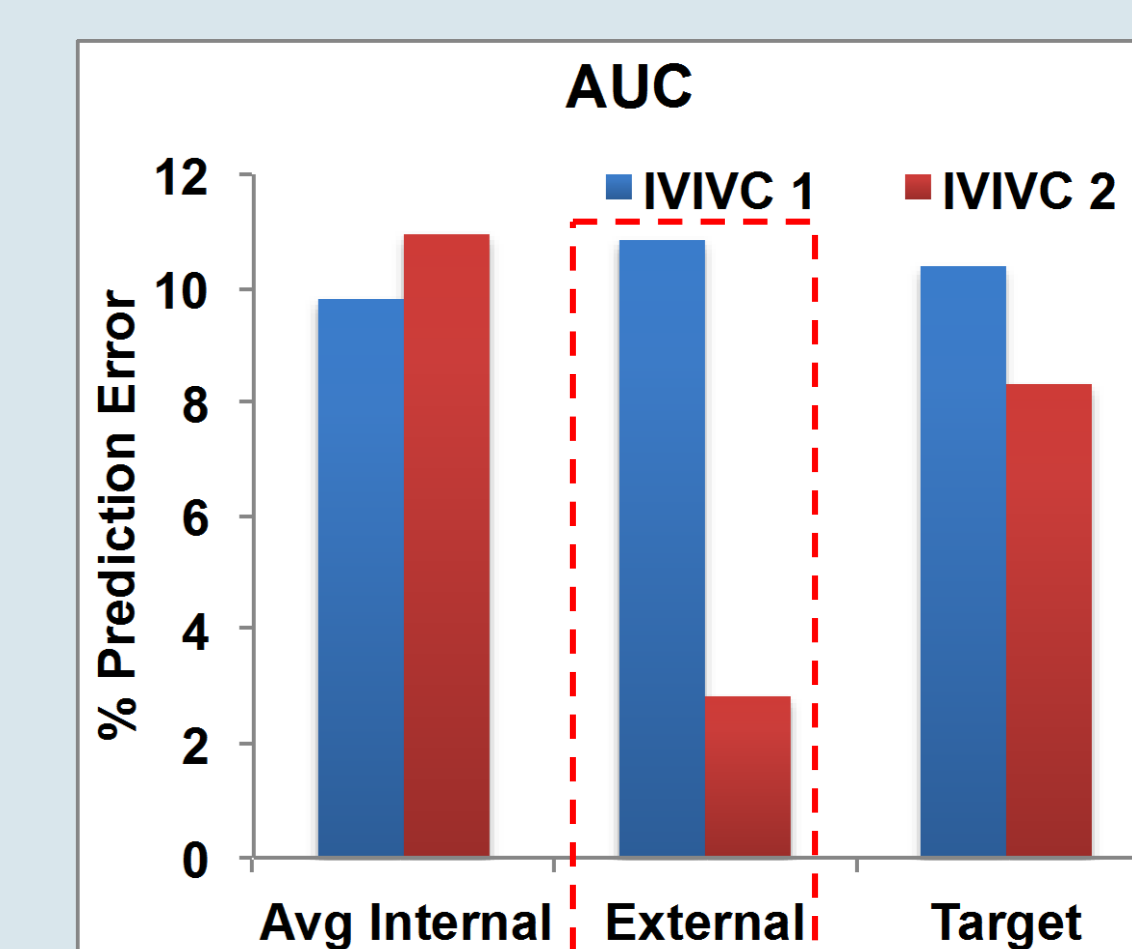
- 1:1 Linear Correlation – Level A**

$$T(\text{In vivo}) = (\text{Time}(\text{In vitro}) * a) + b$$

a=Time scaling factor = 1.6, b= time shifting factor = 1



### % Prediction Error (PE) of IVIVC 1



## CONCLUSION(S)

- The size, porosity and release characteristics of LA microspheres appeared to be sensitive to manufacturing changes.
- Despite the differences in the *in vitro* and *in vivo* release profiles (% burst release and release rate), an affirmative level A IVIVC was developed using the developed *in vitro* release testing method in a rabbit model for peptide microspheres.
- This indicates that the developed *in vitro* release testing method has the potential to predict the *in vivo* performance of microspheres.

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- Disclaimer:** This poster reflects the views of the authors and should not be construed to represent FDA'S views or policies.

**REFERENCES.** 1. Andhariya J. V., Shen J. and Burgess D. J. *et al.*, Journal of Controlled Release, 2017, 255, 27-35. 2. Shen J. and Burgess D. J. *et al.*, Journal of Controlled Release, 2015, 218, 2-12. 3. FDA Guidance for Industry: extended release oral dosage forms: development, evaluation and application of *in vitro/in vivo* correlation, Rockville, MD, 1997.